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1	Isotopic characterization (² H, ¹³ C, ³⁷ Cl, ⁸¹ Br) of abiotic sinks of methyl
2	bromide and methyl chloride in water and implications for future studies
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12 Abstract

Methyl bromide (CH₃Br) and methyl chloride (CH₃Cl) significantly contribute to stratospheric 13 ozone depletion. The atmospheric budgets of both compounds are unbalanced with known sinks 14 outweighing known sources. Stable isotope analysis may be capable to provide additional 15 information for better differentiation of sources and sinks, respectively, and is particularly 16 powerful if isotopes of multiple elements in each compound are used. In the current study, triple-17 element isotope analysis (²H, ¹³C, ³⁷Cl/⁸¹Br) was applied to investigate the two main abiotic 18 degradation processes of methyl halides (CH₃X) in fresh and seawater: hydrolysis and halide 19 exchange. For CH₃Br both nucleophilic substitution reactions caused significant carbon and 20 bromine isotope effects accompanied by a secondary inverse hydrogen isotope effect. Calculated 21 loss rates indicated that exchange with chloride (Cl⁻) may be the dominating abiotic sink for CH₃Br 22 in oceans. For CH₃Cl only hydrolysis was observed at significant rates causing large carbon and 23 chlorine isotope effects and a secondary inverse hydrogen isotope effect. This study demonstrates, 24 25 to our knowledge, the first triple-element isotope analyses of CH₃Cl and CH₃Br. The presented results have important implications for source apportionment of tropospheric CH_3X if viewed in 26 conjunction with previously measured isotope effects caused by other main sinks of methyl 27 halides. 28

TOC graphic



31 Introduction

Methyl chloride (CH₃Cl, chloromethane) and methyl bromide (CH₃Br, bromomethane) together 32 contribute about 30 % to halogen induced ozone loss even though atmospheric concentrations are 33 very low: 540 pptv and 7 pptv respectively¹. CH₃Cl and CH₃Br are emitted by both anthropogenic 34 and natural sources such as fumigation for quarantine and pre-shipment treatment², marine 35 macroalgae³, salt marshes⁴, soils⁵, biomass burning⁶ and tropical plants⁷. Main sinks for both of 36 these compounds are reaction with OH and Cl radicals in the troposphere⁸, uptake by oceans⁹ and 37 soils¹⁰. The atmospheric budgets of both compounds are unbalanced with known degradation 38 processes exceeding the best estimates of known sources by approximately 20 %^{1, 11}. A better 39 understanding of emission and degradation processes will be necessary in order to better quantify 40 emission and degradation of CH₃X and to improve budget estimates. 41

Previous studies suggested that ocean uptake is driven by the abiotic processes hydrolysis and halide exchange as well as microbial degradation^{9, 12, 13}. To a minor extent, hydrolysis may also contribute to degradation of CH₃Br in soils¹⁴. Hydrolysis and halide exchange of CH₃X (CH₃Cl and CH₃Br) are both nucleophilic substitution reactions (S_N2) following second order reaction kinetics. The attacking nucleophiles are either water (H₂O), hydroxide ions (OH⁻), or halide ions such as Cl⁻ and Br⁻ (Y⁻)¹⁵⁻¹⁷:

 $48 \quad CH_3X + H_2O \quad \rightarrow \quad CH_3OH + H^+ + X^- \tag{R1}$

49	$CH_3X + OH^-$	\rightarrow	$CH_3OH + X^-$	(R2)
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50 $CH_3X + Y^- \rightarrow CH_3Y + X^-$ (R3)

In principle, hydrolysis of chlorinated aliphatic compounds may occur due to neutral (R1) and/or alkaline hydrolysis (R2) depending on the pH and the reacting organic compound. For instance,

solely neutral hydrolysis (R1) was detected for CCl₄ whereas some chlorinated ethenes only 53 reacted with hydroxide ions $(R2)^{18}$. For CH₃X it was shown that alkaline hydrolysis required a 54 hydroxide concentration of more than 0.1 mol L⁻¹ and hence only neutral hydrolysis is considered 55 to be relevant in most environments¹⁶. Consequently, reactions R1 and R3 were suggested to 56 constitute large sinks for methyl bromide¹⁹⁻²¹. For methyl chloride, microbial degradation seems 57 to be more important, especially in subarctic and arctic ocean waters due to low reaction rates of 58 abiotic processes⁹. Still, the overall importance of these reactions as a sink of methyl halides 59 compared to microbial degradation in oceans and soils is generally not well understood. 60

Stable isotope analysis was suggested as a diagnostic tool to overcome the limitations of solely 61 quantifying concentration levels of these compounds²². Several studies measured the carbon 62 isotope composition of various sources and sinks and an overview of these isotopic signatures was 63 published for CH₃Br²³ and CH₃Cl²⁴, respectively. Important new information could be derived 64 from stable carbon isotope measurements but both studies also revealed the fact that the ranges of 65 the source signatures are overlapping and often coincide with the tropospheric δ^{13} C of CH₃X. For 66 that reason, stable carbon isotope measurements have not yet contributed to better budget estimates 67 of CH₃Cl and CH₃Br. Significant improvements may be expected if the isotopic compositions of 68 several elements are measured in one compound. Such multi-element isotope approaches have 69 recently become available with improved measurement techniques and were successfully applied 70 to describe, for instance, the fate of organic contaminants in groundwater^{25, 26}. For CH₃Cl, 71 hydrogen isotope measurements were presented recently²⁷⁻²⁹ and bromine isotope analysis was 72 demonstrated in two studies for CH₃Br^{30, 31} but overall no multi-element isotope studies have been 73 published yet for methyl halides. 74

Future estimates of the atmospheric budget of methyl halides may significantly benefit from the 75 additional information provided by multi-element isotope analyses but before any isotope-based 76 models are applicable, source signatures and the isotope fractionation caused by sink processes 77 need to be determined. Knowledge about the isotope fractionation caused by sinks is particularly 78 important for two reasons. Firstly, it provides the link between the isotopic source signatures of 79 these compounds and the tropospheric isotopic composition of methyl halides. Secondly, the 80 extent of isotope fractionation can be used to estimate the loss rate caused by the individual 81 processes, a method commonly used to determine contaminant degradation in groundwater^{22, 32-37}. 82 Consequently the characterization of the individual sink processes is a crucial prerequisite for the 83 application of future isotope based models. 84

Hence the objective of the presented study was to characterize the isotopic enrichment factors (ϵ) 85 caused by hydrolysis and halide exchange reactions with CH₃Cl and CH₃Br, potentially important 86 abiotic sinks for these compounds. We determined the isotope fractionation of all available stable 87 isotopes in each compound, that is, hydrogen and carbon isotopes in both compounds and chlorine 88 as well as bromine isotopes for methyl chloride and methyl bromide, respectively. To our 89 knowledge, the presented data are the first three-dimensional isotope measurements for each of 90 these substances. The implications of our results for future multi-dimensional isotope studies of 91 92 methyl halides are discussed.

94 Materials and Methods

95 Chemicals. Methyl chloride and methyl bromide were purchased as compressed gases and with a 96 purity of more than 99 %. Methyl chloride was obtained from Linde (Germany) whereas methyl 97 bromide was purchased from Gerling, Holtz & Co (Germany). A commercially available sea salt 98 without any additives (Aquasale, Heilbronn, Germany, major ion composition given in Table S1) 99 was purchased to prepare brines with a concentration of 35 g kg⁻¹ (psu) which is similar to the 100 average salt content of seawater.

Preparation of samples and experiments. Stock solutions with a concentration of 10 mmol L^{-1} 101 CH₃Br and CH₃Cl were prepared for hydrolysis experiments and 5 mmol L⁻¹ for halide exchange 102 experiments by injecting the corresponding amount of gas into the headspace of a 1 L crimp-sealed 103 glass bottle filled with distilled water and brine (3.5 %), respectively. Additionally, a stock solution 104 of 0.2 mmol L⁻¹ CH₃Br was prepared to carry out a hydrolysis experiment at a lower concentration. 105 The brine was prepared by mixing distilled water with the sea salt and boiling this solution for 106 10 min. All experiments were carried out in unbuffered solution (see also Results and Discussion 107 for further explanations). After preparation, stock solutions were shaken overnight for 108 109 equilibration before further usage. For each experiment, 6 to 10 septum bottles (60 mL) were filled with 40 mL of solution, crimp-sealed and all bottles shaken for at least 3 hours. Then, the starting 110 concentration was determined by injecting aliquots of the headspace of all samples. At least three 111 112 standards were analyzed to quantify the sample concentrations via a three-point calibration (Supporting Information 3). To avoid gas-leakage through the pinched septa, the sample bottles 113 were kept upside-down throughout the entire experiment. Furthermore, samples were kept at a 114 dark place maintaining a temperature of $23 \pm 1^{\circ}$ C. Sampling of the bottles took place in different 115 time intervals. For methyl bromide, sampling occurred every 3-5 days whereas for methyl chloride 116

117 up to two months passed before another sample was collected. After sampling, each bottle was 118 frozen to -18° C for conservation and stored until the end of the experiment at this temperature. 119 Before analysis, all samples of one experiment were heated simultaneously to 25° C in a water bath 120 and subsequently shaken for two hours to ensure equal treatment of all samples and complete 121 equilibration in the sample bottles.

Stable isotope analysis of carbon, hydrogen, chlorine and bromine. Stable isotope analysis was 122 carried out by injecting aliquots of the headspace gas (50 to 1000 µL depending on concentration 123 and element) into the injector (split mode) of the gas chromatographic systems (GC) using a gas 124 tight syringe with push-button valve (VICI Precision Sampling). The GC was either connected to 125 gas source isotope ratio mass spectrometry (IRMS) for hydrogen and carbon isotope analysis or to 126 multiple collector inductively coupled plasma mass spectrometry (MC-ICPMS) for chlorine and 127 bromine isotope analyses. The analytical procedures followed closely the methods described and 128 published in previous studies for carbon³⁸, hydrogen³⁹, chlorine^{40, 41} and bromine⁴² isotopes. 129 Descriptions of the methods are provided for each method in the Supporting Information 1, 130 including also a cross calibration for bromine isotopes following previously published protocols^{43,} 131 ⁴⁴ (Supporting Information 2). 132

133 Results from isotopic measurements are reported in delta values (δ) for all isotopes. Delta values 134 are calculated according to the following expression⁴⁵:

135
$$\delta^{i}E(Ur) = \frac{(R)sample}{(R)standard} - 1$$
 Equation 1

Here, ⁱE indicates ²H, ¹³C, ³⁷Cl, and ⁸¹Br and R the isotopic ratio ²H/¹H, ¹³C/¹²C, ³⁷Cl/³⁵Cl, and
 ⁸¹Br/⁷⁹Br for hydrogen, carbon, chlorine, and bromine, respectively. The delta values are given in
 Ur (Urey) according to recent IUPAC recommendations⁴⁶. Urey, if expressed in milli-Urey (mUr),

and the more commonly used permil scale (‰) are interchangeable: 1 mUr = 0.001 = 1 ‰, but Ur is, in contrast to permil a SI unit and hence common SI prefixes such as milli- and microbecome available. Other terms for the expression of isotopic ratios such as ppm and permeg may also be reported in Ur. Thus, the unit Urey provides a single unified approach for the expression of all stable isotope ratios. The overall uncertainties of the analytical procedures, including reproducibility, linearity, and scale normalization are usually better than 5 mUr (hydrogen), 0.5 mUr (carbon), 0.2 mUr (chlorine), and 0.1 mUr (bromine).

Enrichment factors and dual element isotope ratios. The isotopic enrichment factor describes the change of the isotopic composition between the substrate and the instantaneous product caused by a reaction or a process⁴⁷. It further characterizes the constant change of the isotopic composition of the substrate reservoir due to the preferential loss of heavy or light isotopes during a reaction or process. In the current study, isotopic enrichment factors ($\varepsilon_{\rm H}$, $\varepsilon_{\rm C}$, $\varepsilon_{\rm Cl}$, $\varepsilon_{\rm Br}$) for CH₃Cl and CH₃Br were determined by using the Rayleigh equation⁴⁸:

152
$$ln\left(\frac{\delta^{i}E+1000}{\delta^{i}E_{0}+1000}\right) \approx ln(f)\varepsilon_{\chi}$$
 Equation 2

where $\delta^{i}E$ is the isotopic signature ($\delta^{2}H$, $\delta^{13}C$, $\delta^{37}Cl$, $\delta^{81}Br$) of the organic after partial degradation, $\delta^{i}E_{0}$ indicates the initial delta value ($\delta^{2}H_{0}$, $\delta^{13}C_{0}$, $\delta^{37}Cl_{0}$, $\delta^{81}Br_{0}$), and *f* is the fraction of organic remaining after partial degradation. The procedure for quantifying *f* is provided in the Supporting Information 3. The Rayleigh equation is appropriate to derive the isotopic enrichment factors for first-order or pseudo-first-order reactions⁴⁹.

158 Λ -values (lambda) describe the ratio of the enrichment factors of isotopes of two different 159 elements⁴⁹. Λ -values are determined as the slope of a linear regression of isotopic signatures of two elements (e.g. H and C) determined from samples of the same experiment. Λ-values may also
 be estimated according to the following relationship⁴⁹:

162
$$\Lambda_{x/y} \approx \frac{\varepsilon_x}{\varepsilon_y}$$
 Equation 3

where ε_x and ε_y are the enrichment factors of two different elements determined for the same mechanism in a certain compound.

165 **Results and Discussions**

Reaction rates. Experiments were performed to investigate the abiotic degradation of CH₃Br and 166 CH₃Cl dissolved in water. Experiments in distilled water were carried out to study hydrolysis 167 reactions only. In brines (seawater) degradation may be due to both, hydrolysis and halide 168 exchange. All experiments were performed at a temperature of $23 \pm 1^{\circ}$ C (296 K). No buffer was 169 added to the stock solutions because buffer catalysis was reported as a complicating factor in 170 previous studies^{16, 50, 51}. Moreover, hydrolysis of CH₃Cl and CH₃Br is expected to primarily follow 171 reaction R1 in the environment. Alkaline hydrolysis of CH₃X (R2) only occurs at significant rates, 172 if OH⁻ concentrations exceed 0.1 mol L⁻¹¹⁶. Consequently, reaction R2 is supposed to be important 173 only at pH 10 and above⁵². Therefore all hydrolysis reactions in this study, which were performed 174 at pH values smaller than 7, were assumed to be independent of the pH and to predominantly 175 176 follow reaction R1. Results for the calculated loss rates of the individual experiments are 177 summarized in Table 1 and are compared to previously published values.

For hydrolysis of CH₃Br two experiments were carried out at different concentrations. Reaction rates of both experiments varied between 3.5 ± 0.8 % d⁻¹ at 10 mmol L⁻¹ and 1.3 ± 0.2 % d⁻¹ at 0.2 mmol L⁻¹. The reaction rate obtained at high concentrations may, however, be influenced by

an additional equilibration effect with Br⁻ ions in the solution also represented by unusual bromine 181 isotope values (see discussion below). During hydrolytic degradation of CH₃Br, Br⁻ ions are 182 released into the solution according to reaction R1. Schwarzenbach et al⁵² (and references therein) 183 published relative nucleophilicities of nucleophiles reacting with CH₃Br. Taking into account these 184 nucleophilicities and the CH₃Br concentration of 10 mmol L⁻¹, released Br⁻ ions reached indeed a 185 concentration high enough to compete with water as a nucleophile. Both hydrolysis and Br-186 exchange are equally important when about 75 % of Br^{-} is released due to transformation of CH₃Br 187 (Supporting Information 4). A second experiment was run at a lower concentration to avoid any 188 of such additional reactions. At 0.2 mmol L⁻¹ the release of Br⁻ ions due to transformation only 189 reached about 2 % of the amount necessary to compete with hydrolysis (Supporting Information 190 4) and no additional effect on the Br isotopic composition was detected. Consequently, hydrolysis 191 of methyl bromide at 0.2 mmol L⁻¹ followed first-order kinetics (pseudo-first order reaction, Figure 192 S4), a prerequisite to reliably apply the Rayleigh equation for quantification of isotopic enrichment 193 factors. The experimentally determined loss rate of 1.3 ± 0.2 % d⁻¹ was lower than at high 194 concentrations but the magnitude was close to the rate of 2.1 ± 0.2 % d⁻¹ reported by Jeffers and 195 Wolfe¹⁶. 196

For experiments with added sea salt the loss rate was about one order of magnitude larger ($11.5 \pm 2.3 \% d^{-1}$) compared to hydrolysis in distilled water and followed first-order kinetics (pseudo-first order reaction, Figure S5). This experimentally determined loss rate agrees, within analytical uncertainty, with $15.4 \pm 6.0 \% d^{-1}$ published by a previous study¹⁷. The ten-times higher rate together with the nucleophilic strength of Cl (Supporting Information 4) suggests that halide exchange dominates the abiotic degradation of CH₃Br in seawater with hydrolysis only contributing to a minor extent to the overall combined loss rate.

For hydrolysis of methyl chloride in distilled water the experimentally determined loss rate of 204 0.15 ± 0.05 % d⁻¹ is in good agreement with 0.14 ± 0.02 % d⁻¹ reported previously⁵¹. The reaction 205 of CH₃Cl with H₂O and salt delivered a loss rate of 0.12 ± 0.02 % d⁻¹ which is indistinguishable 206 from hydrolysis in distilled water. Data from both reactions of CH₃Cl follow first-order kinetics 207 (pseudo-first order reactions, Figure S6 and S7). The similar reaction rates obtained from both 208 experiments may indicate that halide exchange of CH₃Cl at ambient temperatures does not have 209 any measurable effect on the combined loss rate and hence hydrolysis should be the main abiotic 210 degradation mechanism in natural waters. 211

Hydrolysis and associated isotope effects. Isotopic enrichment factors caused by hydrolysis were 212 determined for stable carbon, hydrogen, and bromine isotopes of CH₃Br. At high concentrations 213 (10 mmol L⁻¹) additional equilibration with Br⁻ influenced the reaction rates and the Rayleigh 214 equation may only possess a limited validity for this experiment. Despite this limitation, the carbon 215 isotopic data followed a linear regression with an $\varepsilon_{\rm C}$ of -49.6 ± 5.6 mUr (Figure S9). This value is 216 close to -58.3 ± 6.8 mUr (Figure 1b) which was determined for the low-concentration experiment 217 (0.2 mmol L⁻¹) where no significant Br⁻ exchange occurred. Both $\varepsilon_{\rm C}$ are consistent with a published 218 carbon isotope enrichment factor of $-51.0 \pm 6.0 \text{ mUr}^{53}$. The same authors investigated the pH 219 independence of carbon isotope fractionation by carrying out experiments at different pH (4.6, 7.3, 220 and 8.8) and additionally in un-buffered solutions (pH 3.6 - 6.0). No significant difference was 221 observed in that study for isotopic enrichment factors obtained from unbuffered solutions 222 compared to those with a fixed pH confirming that hydrolysis primarily occurs via one reaction 223 224 (R1). Hence enrichment factors obtained in the current study should be valid for the conditions 225 found in relevant environmental compartments such as surface waters and soils.

Hydrogen isotope enrichment factors could only be measured for samples at high concentrations. 226 CH₃Br concentrations of 0.2 mmol L⁻¹ were too low to meet the isotopic detection limit for 227 hydrogen isotope measurements. Despite the mentioned uncertainties regarding the rate law, the 228 enrichment factor of $+42 \pm 20$ mUr is consistent with a secondary isotope effect resulting from 229 reaction R1 (Figure 1a, Table 2). Secondary effects are usually smaller than primary isotope effects 230 231 and occur in elements located adjacent to a reactive position due to the changing structure of the molecule or influences of bond vibrations, for example⁵⁴. Furthermore, the positive $\varepsilon_{\rm H}$ indicates an 232 inverse isotope effect. The remaining CH₃Br in water becomes successively depleted in deuterium 233 throughout the reaction. Secondary inverse isotope effects of hydrogen are in fact a common 234 feature for nucleophilic substitution reactions of methyl derivatives and this was investigated in 235 several experimental studies in the gas phase as well as in computational studies⁵⁵⁻⁵⁸. Accordingly, 236 the inverse isotope effects may be explained with transition state theory. During S_N^2 reactions the 237 nucleophile (H₂O, Y^{-}) approaches the carbon atom from the side opposite to the halogen atom. In 238 the transition state both the nucleophile and the leaving halogen atom are partly bound to the 239 carbon atom. The tetrahedral geometry of the methyl halide molecule changes to a trigonal 240 bipyramidal geometry in the transition state where the hydrogen atoms are located in a single 241 plane⁵². This structural change is associated with an increase of the bending and stretching force 242 constants, the latter caused by a tightening of the C-H bonds⁵⁷. This increase is represented by a 243 symmetric excitation of the stretching vibration which increases the reaction probability of the 244 molecules containing a C-D bond to a larger extent than for molecules containing a C-H bond⁵⁸. 245 As a result, CH₃Br in the solution becomes enriched in ¹³C and ⁸¹Br (see below) but depleted in 246 ²H. Even though our measured enrichment factor of +42 mUr was smaller than those in the cited 247 248 articles (up to +200 mUr in gas phase experiments), it qualitatively confirms these inverse effects

for different nucleophiles (see also discussion further below) reacting with methyl halidesdissolved in water.

For bromine isotopes in CH₃Br an ε_{Br} of -1.2 ± 0.4 mUr was measured for the experiment carried 251 out at 0.2 mmol L⁻¹ (Figure 1c). At high concentrations (10 mmol L⁻¹) a nonlinear behavior of the 252 δ^{81} Br values could be observed which was not in agreement with the Rayleigh equation (Figure 253 S8). At first, the δ^{81} Br of the substrate became more enriched but then started to converge toward 254 the starting value again. Apparently, the rising concentrations of bromine ions released into the 255 solution started to equilibrate with the CH₃Br substrate. At low concentrations no such effect was 256 observed because Br concentrations in solution were too low to compete with H_2O as a 257 258 nucleophile (Supporting Information 4). Hence, this equilibration is unlikely to occur in most freshwaters and soils, environments where CH₃Br and Br⁻ concentrations are low and where 259 hydrolysis may contribute to degradation. Even in ocean waters the Br⁻ content only reaches 25% 260 of the concentration required to compete with H₂O (Supporting Information 4) which might 261 explain why no indications of such an equilibration effect were observed in experiments with 262 brines, as discussed further below. 263

264 For hydrolysis of CH₃Cl, stable hydrogen, carbon, and chlorine isotope enrichment factors were 265 determined. The degradation experiment of CH₃Cl in distilled water (hydrolysis) was carried out over 232 days and sampling occurred in time steps of 1-3 months. Isotopic enrichment factors of 266 $+25 \pm 6 \text{ mUr} (^{2}\text{H}), -41.7 \pm 10.2 \text{ mUr} (^{13}\text{C}), \text{ and } -5.3 \pm 1.3 \text{ mUr} (^{37}\text{Cl})$ were derived from the 267 Rayleigh plots (Figure 2a-c). No equivalent data is available in the literature for comparison. 268 Compared to hydrolysis of CH₃Br, enrichment factors for hydrogen and carbon showed a similar 269 magnitude and direction; that is, carbon isotope effects were relatively large and normal whereas 270 hydrogen isotope effects were small and inverse due to a secondary isotope effect caused by the 271

nucleophilic substitution reaction. The chlorine isotope enrichment factor is about 3 times larger than the measured bromine isotope effect in CH_3Br which is consistent with a previous estimate based on theoretical calculations for primary kinetic isotope effects for halogens⁵⁹.

Isotope effects caused by halide exchange reactions. The nucleophilic reaction of halide ions 275 was the second abiotic degradation process investigated in this study. The enrichment factors 276 obtained from experiments with CH₃Br dissolved in brine were $+22 \pm 13$ mUr, -63.3 ± 5.1 mUr, 277 and -1.2 ± 0.2 mUr for ²H, ¹³C, and ⁸¹Br, respectively (Figure 1d-f and Table 2). The measured 278 carbon isotope enrichment factor (-63.3 ± 5.1 mUr) agrees well with -57.0 ± 5.0 mUr reported by 279 Baesman and Miller⁵³. Another study published an $\varepsilon_{\rm C}$ of -41.2 mUr for this reaction which is by 280 about 20 mUr smaller⁶⁰. Compared to the hydrolysis experiments carried out in the current study, 281 enrichment factors for halide exchange are indistinguishable if the analytical uncertainty is taken 282 into account (Table 2). Chlorine ions dominated the exchange reaction with CH₃Br because Cl⁻ 283 concentrations were about nine times higher than necessary to compete with water as a nucleophile 284 (Supporting Information 4). The product of this reaction was CH₃Cl which could be identified 285 during δ^{13} C-CH₃Br measurements (Supporting Information 5). The measured δ^{13} C values of the 286 generated CH₃Cl were indistinguishable from the δ^{13} C predicted by the Rayleigh equation for the 287 cumulative product (Table S3, Figure S10) and CH₃Cl is considered the major product of this 288 reaction. 289

Exchange of chlorine with bromine is a sink for CH₃Br but simultaneously constitutes a source for CH₃Cl. Still, oceanic concentrations of CH₃Br are very low compared to CH₃Cl. Hu et al.⁶¹ reported average concentrations of 2 pM which is much lower than the 88 pM found for CH₃Cl during the same cruise in the Atlantic Ocean. Thus the transformation of CH₃Br would only marginally (< 2%) increase the total CH₃Cl concentration in sea water even if all CH₃Br were completely transformed to CH_3Cl . Chlorine exchange in CH_3Br is therefore not a significant source of CH_3Cl despite its importance as an abiotic sink for CH_3Br .

The degradation of CH₃Cl dissolved in brine generated similar enrichment factors as for hydrolysis 297 of CH₃Cl in distilled water: $+24 \pm 19$ mUr for hydrogen, -40.6 ± 13.9 mUr for carbon and -298 5.2 ± 1.0 mUr for chlorine (Figure 2d-f, Table 1). In contrast to CH₃Br, the addition of sea salt did 299 not increase the reaction rates of CH₃Cl (see discussion above) and it can be assumed that only 300 hydrolysis took place. Furthermore, chlorine isotope measurements did not deliver any evidence 301 for equilibrium exchange of Cl⁻ with CH₃Cl. The CH₃Cl dissolved in the brine had a starting δ^{37} Cl 302 of $+6.02 \text{ mUr SMOC}^{40}$ whereas Cl⁻ in the added sea salt should be close to 0.0 mUr SMOC⁶². In 303 our experiments we observed a shift of δ^{37} Cl toward more enriched values following clearly first-304 order kinetics (Figure S7). In contrast, exchange of CH₃Cl with Cl⁻, if occurring at ambient 305 temperatures, should have caused a shift of δ^{37} Cl in the direction of the lighter values of the added 306 sea salt. Thus, halide exchange in CH₃Cl may be considered negligible at ambient temperatures 307 and should not affect the δ^{37} Cl of methyl chloride in most environments. 308

Overall, the results of the current study give further insights into the importance of these two 309 reactions for degradation of CH₃X in water and provide first isotopic enrichment factors for 310 311 hydrogen and halogens. For CH₃Br, halide exchange should be the dominant abiotic degradation mechanism in seawater which confirms the findings of previous studies^{15, 53, 63}. Hydrolysis may 312 have a rather minor role in degrading CH₃Br in freshwaters and soils. Hydrolysis of CH₃Br also 313 occurs in oceans at a ten times slower rate and isotopic enrichment factors are similar to those 314 obtained from halide exchange reactions due to the same reaction mechanism (S_N2 reaction). 315 Consequently, the two processes cannot be individually characterized and quantified with isotopic 316 methods and both processes may be included as a combined abiotic sink in future isotope-based 317

budget estimates. Still, if necessary, degradation by the individual processes can be estimated from
the relative rate difference of these abiotic processes which should be constant at relevant
temperatures in seawater. For CH₃Cl, only hydrolysis may occur as an abiotic mechanism in most
environments with reaction rates being one order of magnitude lower than for hydrolysis of CH₃Br.
Consequently, hydrolysis should only marginally contribute to degradation of CH₃Cl in oceans,
freshwater, and soils as confirmed by previous studies^{51, 64}.

Dual-element isotope ratios. A-values provide a more precise parameter than ε values to 324 characterize and compare reaction mechanisms because these ratios are insensitive to masking and 325 rate limitation by additional processes⁶⁵. For the current study, lambda values were derived from 326 dual-element isotope plots provided in Figure S11–S14. The resulting $\Lambda_{H/C}$, $\Lambda_{C/CI}$, and $\Lambda_{C/Br}$ are 327 given in Table 2. $\Lambda_{H/C}$ values ranged from -0.3 to -0.7 for both compounds and both reaction 328 pathways. The $\Lambda_{C/CI}$ for CH₃Cl ranged from 6.9 to 7.3 and the $\Lambda_{C/Br}$ of CH₃Br ranged from 46.1 to 329 48.2. No other lambda values have been reported yet for both CH₃Cl and CH₃Br. Some $\Lambda_{H/C}$ for 330 reactions of CH₃Cl could be calculated from published ε_H and ε_C according to Equation 3 because 331 these enrichment factors were derived from the same experiment. The resulting $\Lambda_{H/C}$ are given in 332 Table S4. For CH₃Br no dual-element isotope studies are available yet and therefore lambda values 333 of other reaction pathways could not be calculated. 334

The $\Lambda_{H/C}$ of -0.6 for hydrolysis of CH₃Cl shows a similarly small absolute value as degradation by methylotrophic bacteria²⁷ ($\Lambda_{H/C} = 0.7$, Table S4), differing by the algebraic sign due to an inverse ε_{H} for hydrolysis. Consortia of soil microbes showed slightly larger $\Lambda_{H/C}$ which ranged from 1.3 up to 4.6 due to a decreasing ε_{C} with decreasing CH₃Cl concentrations⁶⁶. These small lambdas for abiotic and biotic reactions in water are the result of relatively small secondary hydrogen isotope effects (<-50 mUr) due to rupture of the C-X bond. Abiotic and biotic reactions in water are, however, still distinguishable by opposing positive and negative $\Lambda_{H/C}$.

In contrast, the main abiotic degradation pathways of CH₃Cl in the gas phase (OH·, Cl· radical reactions) are characterized by larger $\Lambda_{H/C}$ of 23.6 to 27.5 (Table S4) based on the enrichment factors published by two recent studies^{28, 67}. Radical reactions cleave the C-H bond and therefore primary isotope effects for both C and H can be observed causing an overall larger $\Lambda_{H/C}$ which is clearly distinguishable from reactions in water, even though only two isotopic systems are used.

Implications for future isotope-based studies of CH₃X. The results presented in this paper 347 provide a first glimpse of the capabilities of triple-element isotope analysis of CH₃X for 348 identification and characterization of sinks of methyl halides. Specifically, the pattern of isotopic 349 350 shifts defined by enrichment factors and lambda values will be a useful tool to distinguish abiotic degradation processes from other sinks of CH₃X. A comparison of isotope effects measured for 351 352 degradation mechanisms of CH₃Cl demonstrates that basically all known relevant abiotic and 353 biotic degradation mechanisms in water (and soils) are due to a C-Cl bond cleavage in CH₃Cl (Figure 3). Resulting $\Lambda_{C/Cl}$ are relatively similar for all these reactions but $\Lambda_{H/C}$ may still be used 354 to distinguish abiotic from biotic degradation due to opposing inverse and normal hydrogen 355 356 fractionation, respectively (Table 2, Table S4). Methylotrophic bacteria, for instance, were identified as the main biotic sink in water and soils because these organisms are capable of 357 degrading large amounts of methyl halides⁶⁸. Large carbon isotope enrichment factors (-38 to -358 41 mUr) were reported for this biotic sink but rather small secondary hydrogen isotope enrichment 359 factors (-27 to -29 mUr) due to cleavage of the C-Cl bond²⁷. Halogen isotope effects for aerobic 360 microbial degradation of CH₃Cl (or CH₃Br) have not been measured yet but it is conceivable that 361 they show a similarly large isotopic fractionation as reported for other halogenated alkanes. 362

Aerobic microbial degradation of 1,2-dichloroethane, for instance, caused an ε_{Cl} of at least -3.8 mUr⁶⁹ which was similar to the ε_{Cl} reported for anaerobic microbial degradation (-4.2 mUr)²⁶.

In contrast, CH_3X in the troposphere mainly degrades *via* OH and Cl radical reactions¹ which 365 cause a C-H bond dissociation. Consequently, reported hydrogen isotope effects for CH₃Cl were 366 large²⁸ (>-264 to -280 mUr) and carbon isotope effects were moderate (-10.2 to -11.2 mUr)⁶⁷. 367 Halogens present in CH₃X should only show small secondary isotope effects because they are not 368 involved in radical reactions⁵⁴. Consequently, gas phase reactions of CH₃Cl show a completely 369 different pattern of isotopic shifts compared to reactions in water and soils (Figure 3). This 370 separation of fractionation patterns of aqueous and gas phase reactions may also be conceivable 371 for CH₃Br because biotic and abiotic degradation mechanisms are largely the same¹. The 372 possibility to clearly distinguish degradation in water from degradation in the gas phase may 373 considerably simplify the characterization and quantification of sinks of methyl halides when 374 triple-element isotope analysis is applied. 375

376 In order to fully benefit from the advantages of triple-element isotope analysis in the future, however, it will be necessary to determine three-dimensional isotopic fingerprints of atmospheric 377 378 samples and of the largest sources (macroalgae, salt marshes, biomass burning, plants, soils). 379 Moreover, full sets of isotopic enrichment factors and lambda values for the main sinks (OH radical reactions, microbial degradation in oceans and soils) must be determined. Once these tasks are 380 completed, the isotopic data can be fed into models. Previous models relied on upscaled emission 381 data, atmospheric concentrations and corresponding stable carbon isotopic signatures to create 382 isotopic mass balances for both CH₃Cl and CH₃Br^{24, 67, 70}. The future use of two additional isotopic 383 systems for each compound may substantially improve this mass balance approach because of the 384 different sensitivity of hydrogen and halogen isotopes for radical reactions in the atmosphere and 385

degradation in water/ soil respectively. Hence, three different isotopic mass balances may be 386 created which have to yield matching results for emissions and degradation rates thus providing a 387 tool for verification of this mass balance approach. Apart from these bottom-up approaches, 388 inverse top-down models using isotopic data might be appropriate methods for source 389 apportionment of atmospheric compounds⁷¹. These inverse models estimate source and sink terms 390 from variations in the atmospheric composition but require long-term monitoring using a relatively 391 dense network of sampling stations⁷². Inverse models may, however, provide an alternative route 392 to calculate the atmospheric budget but for CH₃X the application of these models is currently still 393 out of reach because of the challenges to regularly measure the isotopic composition of 394 atmospheric CH₃X. Once these challenges are overcome triple-element isotope analysis may 395 provide a realistic chance to better quantify the unbalanced atmospheric budgets of CH₃X and to 396 identify the putatively missing sources. 397

399 Associated content

400 The Supporting Information is available free of charge on the ACS publications website:

Additional information on analytical methods (isotope analysis, quantification), cross-calibration of standards for bromine isotope analysis, relative nucleophilicities of halogens and water, graphical determination of the rate laws of the different reactions, isotopic measurements of carbon and bromine for hydrolysis of CH₃Br at high concentrations, determination of lambda values from the current and previous studies, and isotopic information of product-CH₃Cl

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619 Figures and Tables



Figure 1. Rayleigh plots for abiotic reactions of CH₃Br in water. Panel (a), (b), and (c) show results of the reaction CH₃Br + H₂O (hydrolysis). Panel (d), (e), and (f) demonstrate isotope effects due to CH₃Br + H₂O + Y⁻ (hydrolysis and halide exchange combined). The slope of the regression indicates the enrichment factor in Ur. Error bars represent the analytical uncertainty of 5 mUr $(\delta^{2}$ H), 0.5 mUr (δ^{13} C), and 0.1 mUr (δ^{81} Br). For carbon isotopes, error bars are smaller than the used symbols. The quantification was carried out with an uncertainty of usually better than 5 %.



Figure 2. Rayleigh plots for reactions of CH₃Cl. Panel (a), (b), and (c) show results of the reaction CH₃Cl + H₂O (hydrolysis). Panel (d), (e), and (f) represent isotope effects due to CH₃Cl + H₂O + Y- (combined hydrolysis and halide exchange). The slope of the regression indicates the enrichment factor in Ur. Error bars represent the analytical uncertainty of 5 mUr (δ^{2} H), 0.5 mUr (δ^{13} C), and 0.2 mUr (δ^{37} Cl). For carbon isotopes, error bars are smaller than the used symbols. The quantification was carried out with an uncertainty of usually better than 5 %.



Figure 3. Illustration of fractionation effects of CH₃Cl in water and in the gas phase (troposphere). 635 The δ^2 H, δ^{13} C, and δ^{37} Cl are expressed as the relative difference from δ^2 H₀, δ^{13} C₀, and δ^{37} Cl₀ of 636 the undegraded sample. All isotopic values for abiotic reactions in water are taken from the current 637 study whereas carbon and hydrogen isotope values are taken from the literature (Table S4). 638 Fractionation for chlorine isotopes in biotic reactions has been estimated assuming a minimum 639 isotope effect of -4 mUr based on previous studies on chlorinated alkanes as described in the text 640 (average of aerobic/ anaerobic reactions). For gas phase reactions an insignificant secondary 641 chlorine isotope effect was assumed being smaller than analytical uncertainty of 0.2 mUr. The 642 figure demonstrates a clear distinction between isotopic fractionation in the gas phase 643 (troposphere) and fractionation in water. 644

	loss rates ¹ [%·d ⁻¹]	loss rates (previous studies) [%·d ⁻¹]
$CH_3Br + H_2O$	1.3 ± 0.2	2.1 ± 0.2^{a}
$CH_3Br + H_2O + Y^2$	11.5 ± 2.3	15.4 ± 6.0^{b}
$CH_3Cl + H_2O$	0.15 ± 0.05	$0.14 \pm 0.02^{\circ}$
$CH_3Cl + H_2O + Y^-$	0.12 ± 0.03	-

Table 1: Loss rates for hydrolysis and halide exchange of CH₃Br and CH₃Cl

¹ Loss rates were determined graphically from plots of $\ln[f]$ versus time and indicate the loss rate

at 23±1°C (296 K). The errors are given as the 95% confidence interval. Literature values are

taken from ^a Jeffers and Wolfe¹⁶, ^b King and Saltzman¹⁷, ^c Elliot and Rowland⁵¹;

649 Table 2: Isotopic enrichment factors (ϵ) and lambda values (Λ) of abiotic reactions in water

	ε _H mUr	ε _C mUr	ε _{Cl} mUr	ε _{Br} mUr	$\Lambda_{\rm H/C}$	$\Lambda_{C/Cl}$	$\Lambda_{C/Br}$
$CH_3Br + H_2O (10 \text{ mmol}L^{-1})$	$+42 \pm 20$	-49.6 ± 5.6	-	n.d.			
$CH_3Br + H_2O (0.2 \text{ mmol } L^{-1})$	n.d.	-58.3 ± 6.8	-	-1.16 ± 0.42	$-0.7 \pm 0.2*$		46.1 ± 16.1
$CH_3Br + H_2O + Y^2$	$+22 \pm 13$	-63.3 ± 5.1	-	-1.22 ± 0.23	-0.3 ± 0.2		48.2 ± 6.5
$CH_3Cl + H_2O$	$+25\pm6$	-41.7 ± 10.2	-5.3 ± 1.3	-	-0.6 ± 0.3	7.3 ± 0.9	
$CH_3Cl + H_2O + Y^-$	$+24 \pm 19$	-40.6 ± 13.9	-5.2 ± 1.0	-	-0.6 ± 0.2	6.9 ± 2.3	

Enrichment factors are derived from the Rayleigh plots in Figure 1 and Figure 2. Lambda values are determined graphically from dual-elemental isotope plots (Figure S11-S14, Supporting Information). Errors are given as the 95% confidence interval of the regressions. Values in italics indicate that these reactions might not strictly follow first-order kinetics and epsilons only serve as an approximation despite acceptable correlation coefficients; n.d. means "not determined"; * indicates that this value was calculated using Equation 3.