

This is the accepted manuscript version of the contribution published as:

Schlüter, S., Sammartino, S., Koestel, J. (2020):

Exploring the relationship between soil structure and soil functions via pore-scale imaging
Geoderma **370** , art. 114370

The publisher's version is available at:

<http://dx.doi.org/10.1016/j.geoderma.2020.114370>

Exploring the relationship between soil structure and soil functions via pore-scale imaging¹

Steffen Schlüter^{1*}, Stephane Sammartino², John Koestel³

1. Dept. Soil System Sciences, Helmholtz-Centre for Environmental Research – UFZ, Halle, Germany

2. Avignon Université, INRAE, UMR EMMAH, F-84000, Avignon, France

3. Institute of soil and environment, Swedish University of Agricultural Sciences, Uppsala, Sweden

* corresponding author: steffen.schlueter@ufz.de

Abstract

Biogeochemical and structural heterogeneities at the pore-scale govern processes in soil in many ways. They are therefore of key importance for understanding soil functioning. Prominent examples are the stabilization of soil organic matter due to reduced bioavailability in aggregated soil structure, preferential transport of nutrients and contaminants along macropores, highly localized greenhouse gas emission around a few hotspots of microbial activity like particulate organic matter and the formation of the rhizosphere as a complex system composed of plant roots, soil and associated microorganisms.

All of these processes have in common that the underlying relevant mechanisms are fairly well understood in artificial systems with reduced degrees of heterogeneity, like soil suspensions, glass beads, micromodels with known structure and so on. However, the far more complex pore architecture of undisturbed soils leads to emergent system behavior which needs to be addressed when studying these structure-mediated processes. The opaque nature of soils predestines the use of non-invasive imaging techniques for exploring how biogeochemical and structural heterogeneities are shaping soil functions. Such imaging techniques and subsequent image analyses are now widely used to study soils. While previously many properties were defined only by averaged bulk parameters, pore-scale imaging reveals details at smaller scales and provides spatial information, in two, three or even more dimensions including time or multispectral data.

¹ This editorial contains parts of the habilitation thesis: Steffen Schlüter (2019): Exploring the relationship between soil structure and soil functions via pore-scale imaging”, Martin Luther University Halle-Wittenberg, Germany.

This virtual special issue presents fourteen contributions that employ pore scale imaging in order to highlight the role of soil structure on soil functions or reversely the effect of soil processes on soil structure or report methodological advancements in pore scale imaging. In this editorial we briefly outline the different conceptions of soil structure, demonstrate the relevance of soil structure for various soil functions with a specific example and review the different avenues along which recent advances in pore scale imaging of soil have been made, before we briefly categorize and summarize the contributions to this virtual special issue.

1. Introduction

1.1. Definition of Soil Structure

Soil structure is the three-dimensional arrangement of solid soil constituents and voids across different scales (Rabot et al., 2018), resulting from interactions of biotic and abiotic factors, including climate, mineral composition, organic matter, roots, fungal hyphae, soil fauna, and tillage. The hierarchical organization of soil structure is depicted in Figure 1, starting from soil horizons within a profile, over structural features within a horizon like macropores and soil clods in a plowed horizon down to root channels, cracks and spatial variability of matrix porosity within individual clods. Soil structure constitutes the habitat for soil organisms, provides the paths for matter fluxes and the accessibility to the chemical interfaces for reactions in soil (Figure 1). At the same time, soil structure is shaped by biological, chemical and physical processes and the feedbacks between them. It can thus evolve more or less rapidly under these processes that are also influenced by seasonal variations and climatic events such as long droughts and heavy rain. Essential ecosystem functions of soil like water storage and transport, nutrient cycling, carbon storage, plant growth and the maintenance of biodiversity are to a large degree governed by soil structure (Figure 1). In fact, soil structure can be interpreted as an integral indicator of the soil ecological status, i.e., the capacity to host organisms, to sustain the production of biomass, and to recover from external perturbation (Kibblewhite et al., 2008; Rabot et al., 2018; Young and Crawford, 2004).

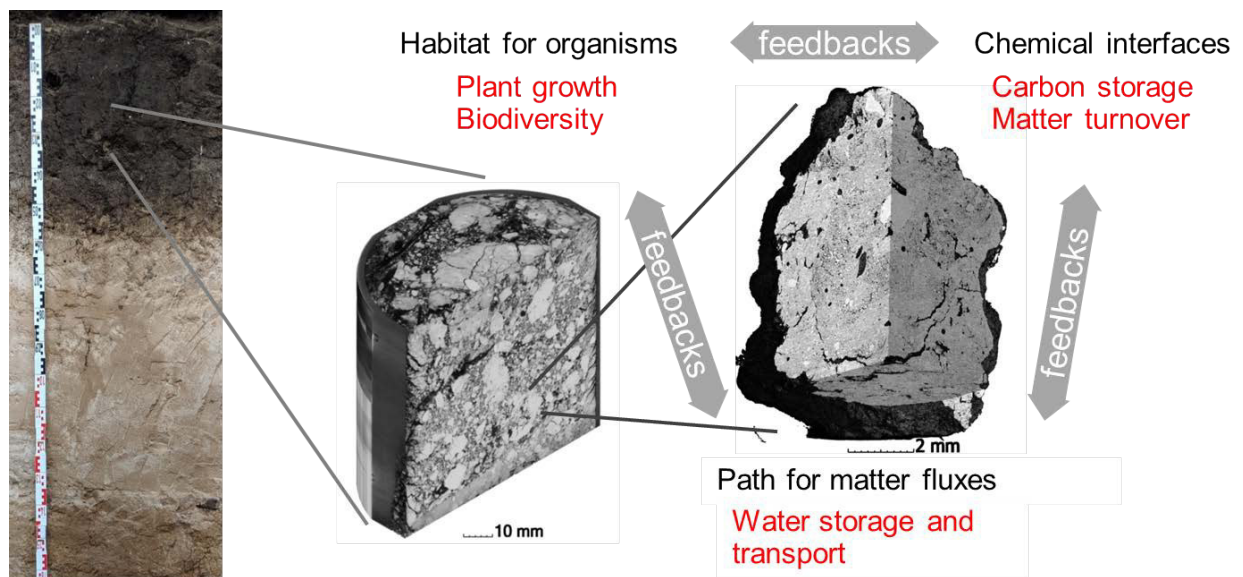


Figure 1: Schematic of the hierarchical organization of soil structure, the various properties it fulfills and the soil functions that are governed by it.

1.2. Two Perspectives on Soil Structure

There is a dichotomy in soil structure assessment through either the aggregate perspective or pore space perspective (Figure 2). A lively debate is currently ongoing in the soil science community especially about the usefulness of the aggregate perspective (Kravchenko et al., 2019a; Rabot et al., 2018; Wang et al., 2019; Yudina and Kuzyakov, 2019).

The traditional approach has always been to characterize soil structure through the size, shape, grade and stability of soil aggregates (Ad-hoc-AG Boden, 2005; Jahn et al., 2006). This is standard procedure in soil surveys worldwide and common practice for farmers to evaluate the soil ecological status from quick field observations. The fastest and cheapest soil structure assessment is achieved with drop-shatter tests, for which a spade-full of topsoil is dropped from a certain height, e.g 1 m, and a score is derived from the size distribution, and shape of aggregates as well as earthworm and root abundance (Ball et al., 2007; McKenzie, 2001; Shepherd et al., 2008). The scoring results have been shown to correlate with soil compaction and the associated decrease in gas exchange, infiltration and agricultural production (Guimarães et al., 2013; Mueller et al., 2009; Pulido Moncada et al., 2014; Shepherd, 2003). However, scoring methods are rarely used in basic research because the assessment is somewhat subjective and the results depend on texture as well as time-variant soil moisture and biological activity (Guimarães et al., 2011; Mueller et al., 2009; Newell-Price et al., 2013).

Soil structure can be measured more objectively with laboratory methods. The most common method is to measure bulk density and derive porosity from the dry weight of undisturbed soil cores with a given volume. The soil core extraction in the field, might not capture the field variability representatively, is known to induce some disturbance along the wall and can become impossible for high rock content and in the presence of woody roots (Page-Dumroese et al., 1999; Schlüter et al., 2011; Vincent and Chadwick, 1994). Another common approach to characterize soil structure through aggregate size distribution and stability according to various protocols grouped into wet-sieving and dry-sieving (Díaz-Zorita et al., 2002). The results are highly sensitive to specific details of each protocol (energy, duration, repetition) and antecedent soil moisture (Almajmaie et al., 2017; Beare and Bruce, 1993). Despite these drawbacks aggregative sieving and stability tests remain being used as they can inform about the susceptibility to slaking (wet-sieving) and resistance to mechanical disturbance (dry-sieving). They are also relatively easy to perform. Moreover the fragmentation of soil into different aggregate size classes supports the concept of an aggregate hierarchy which assumes that macroaggregates ($>250\ \mu\text{m}$) form around particulate organic matter and microaggregates ($<250\ \mu\text{m}$) are released upon breakdown of macroaggregates (Angers et al., 1997; Tisdall and Oades, 1982). The macroaggregates are supposed to be relatively short-lived as their binding agents are less persistent than those within microaggregates.

This aggregate perspective on soil structure has frequently been criticized as the associated methods rather aim at measuring the stability of soil structure than soil structure itself and the outcome of these measurements highly depend on the applied energy (Baveye, 2006; Letey, 1991; Pagliai and Vignozzi, 2002; Young et al., 2001). Moreover, it is the pore space and not the solid space that constitute the spatial domain for water flow, matter fluxes and gas exchange, the habitat for soil biota and the reactor of a multitude of reactions. Therefore, the characterization of pore space attributes in undisturbed soil seems more promising to relate it to soil functions (Rabot et al., 2018). Methods for pore space characterization can be roughly grouped into indirect methods and direct methods. Indirect methods (e.g. mercury porosimetry, pressure chamber) derive pore attributes like the pore size distribution from functional behavior like water retention curves. Direct observations of pore structure are based on imaging (e.g. thin section microscopy, X-ray tomography). They allow for a qualitative assessment of pore structure according to its formation (packing voids, microcracks, root channels, earthworm burrows) as shown in Figure 2 and for a quantitative assessment through image analysis resulting in properties like pore size distribution, pore connectivity or pore distances. Both direct and indirect methods for pore structure assessment tend to be more time-consuming and labor-sensitive

than aggregate structure assessment. Imaging methods, in particular, suffer from limited access to the required hardware and from some degree of subjectivity in the image processing protocols (Baveye et al., 2010).

In summary, both approaches have their strengths and weaknesses and, more importantly, fulfill different purposes and provide complementary information. Therefore, there is a clear merit in combining both approaches for a more comprehensive picture on the links between soil structure and soil functions.

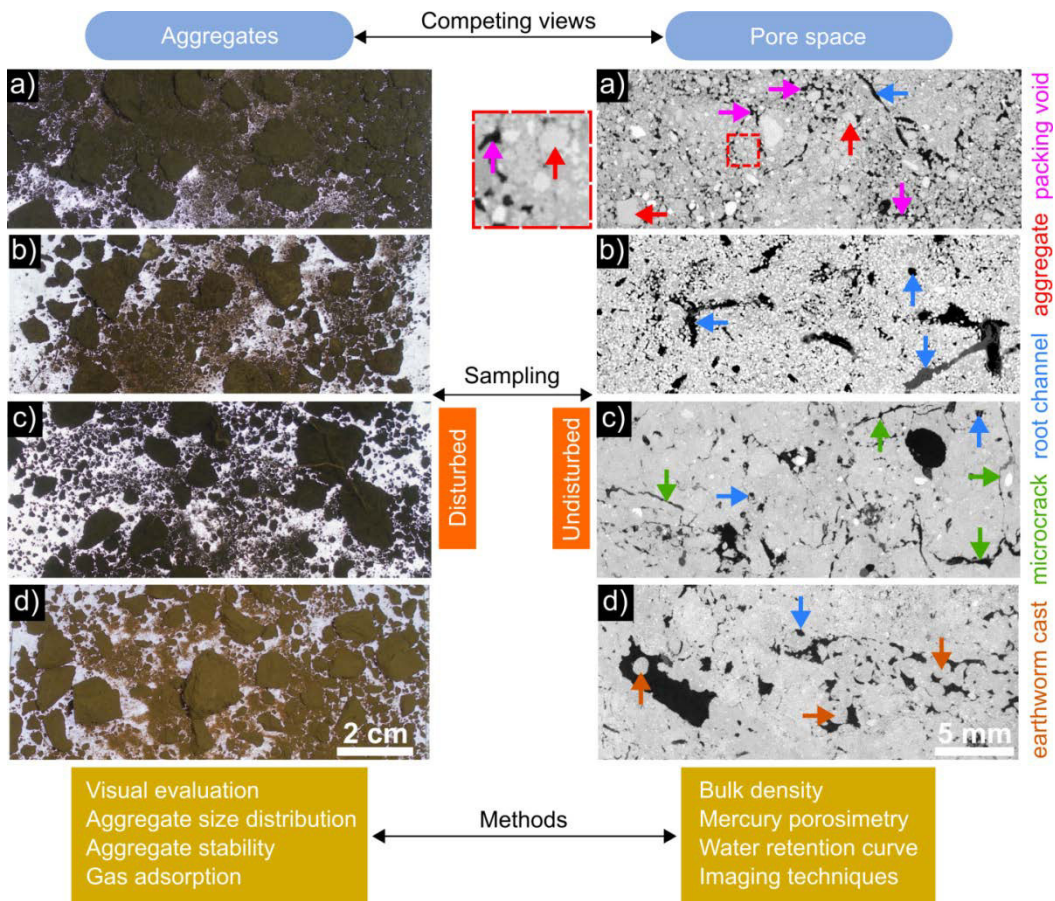


Figure 2: Summary of two competing views: the aggregate perspective and the pore space perspective. (a) Kühnfeld, Halle, Germany (continuous maize, conventional tillage, 63% sand, 25% silt, 12% clay), (b) Hadera, Israel (orchard, 65% sand, 16% silt, 19% clay), (c) Bad Lauchstädt, Germany (grassland, 12% sand, 68% silt, 20% clay), (d) Garzweiler, Germany (crop rotation, below plow layer, 5% sand, 81% silt, 14% clay); modified from (Rabot et al., 2018)

1.3. Relevance of soil structure for soil functioning

A comprehensive review of soil structure effects on these soil functions is beyond the scope of this introduction, but can be found elsewhere (Kravchenko and Guber, 2017; Rabot et al., 2018). Instead we use an illustrative example to demonstrate the role of soil structure for a few fundamental soil processes and their implications for some of the abovementioned soil functions.

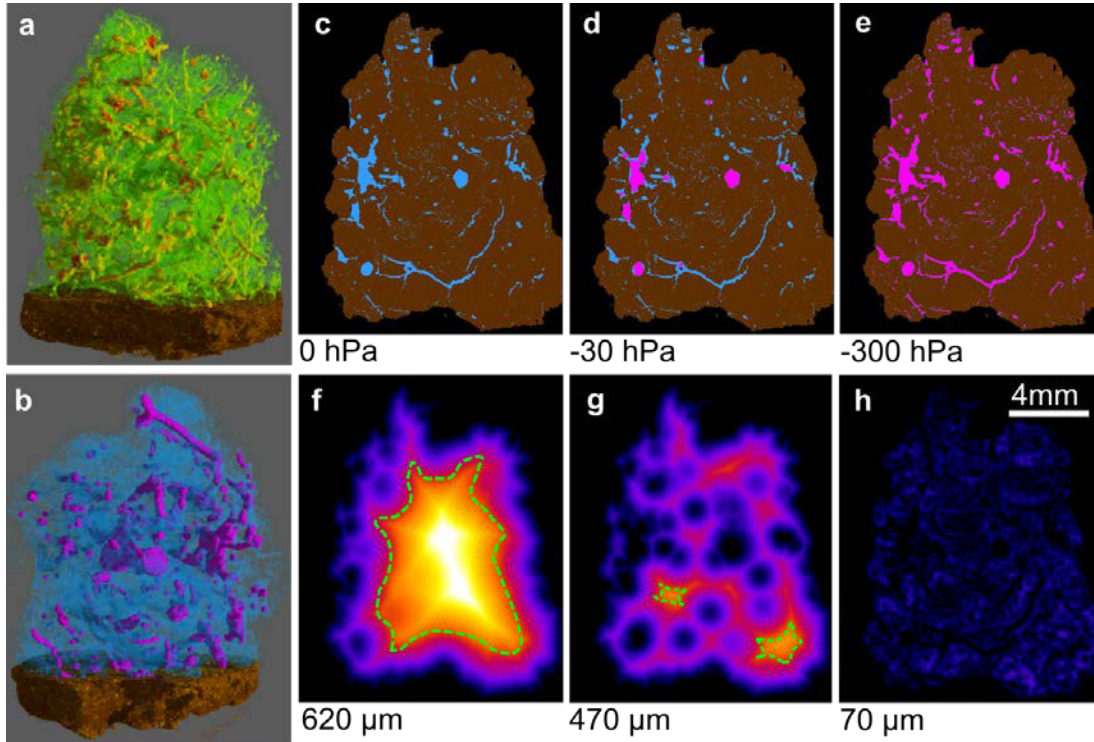


Figure 3: X-ray CT scans of a 1cm thick silt loam aggregate scanned at a resolution of 8μm: (a) the pore size distribution is depicted from small (green) to large (red) diameters; (b) the 3D distribution of air and water at -30hPa modeled with the maximum inscribed sphere method; (c-d) 2D sections of the modeling results at different matric potentials; (f-h) air distances within the aggregate at the same potentials. The values represent average distances and the green lines delineate hypothetical perimeters of anoxic centers.

Figure 3(a) shows the pore architecture of a 1 cm large, silt loam aggregate scanned with X-ray CT at a voxel resolution of 8 μm. The pore size distribution is obtained with the maximum inscribed sphere method and depicted from small (green) to large (red) diameters. This pore size information can be used to model water retention and the distribution of water and air at a certain matric potential by employing Young-Laplace law and the capillary rise equation that is derived from it:

$$h = \frac{2 \gamma \cos(\alpha)}{\rho_w g r} \quad (1)$$

where h is the rise above a free water table in a cylindrical capillary with radius r , γ is interfacial tension between water and air, α contact angle, ρ_w is the density of water and g is gravitational acceleration. In hydraulic equilibrium this height above the free water table can be directly interpreted as pressure head h_m (or capillary pressure P_c or matric potential ψ_m) in that soil depth. Assuming a capillary bundle model, Eq. (1) can be recast to directly infer whether a pore with a certain radius will be water or air-filled at a certain matric potential. This pore morphology based simulation of water retention is done in Figure 3(b) for a matric potential of -30 hPa assuming pure water and perfect wettability. Note that the capillary bundle assumption is a severe oversimplification, because in order to drain a pore it is not only important whether its radius is large enough, but also whether there exists a continuous path towards the atmosphere through which air can invade (Hazlett, 1995; Hilpert and Miller, 2001). However, for the following example this difference is not important. Repeating this analysis for decreasing matric potentials resembles a drainage process (Figure 2c-e). The soil moisture characteristic (or water retention curve or moisture release curve or pF curve) could be directly estimated from the water content at each drainage step. Larger packing pores and root channels are drained first, whereas air invades microcracks and smaller intra-aggregate pores at a more negative matric potential. At a matric potential of -300 cm $\approx$ pF 2.5 all macropores ($>50\text{ }\mu\text{m}$) and narrow macropores ($>10\text{ }\mu\text{m}$) are drained and the soil has reached field capacity (Figure 2e). The unresolved mesopores ($>0.2\text{ }\mu\text{m}$) act as a reservoir for root water uptake as they hold the water against gravity by capillary forces. The visible macropores, in turn, are essential for soil aeration at field capacity or for preferential flow and solute transport when the soil is fully saturated (Rabot et al., 2018). Note that a large part of the unresolved mesopores are textural pores between primary particles, whereas all visible pores are structural pores.

In summary, water retention and soil aeration patterns are a direct imprint of the underlying pore architecture. But this also has important ramifications into matter cycles as they govern diffusion pathways, microhabitats and reaction patterns. This is demonstrated with contact distances to the closest air-filled pore at different matric potentials (Figure 2f-h). At full saturation, air is only present outside the aggregate so that on average dissolved oxygen has to diffuse $620\text{ }\mu\text{m}$ from the aggregate boundary to reach any location within the soil matrix. For sake of simplicity this contact distance is estimated by direct Euclidean distances, whereas pores are tortuous and real diffusion trajectories along a concentration gradient are not straight but chaotic due to Brownian motion. When the aggregate is drained this average air distance decreases substantially. At field capacity it already decreased by one order of magnitude ($70\text{ }\mu\text{m}$), which entails a much better supply with dissolved oxygen in the water-filled

soil matrix. This dissolved oxygen will be consumed through aerobic respiration in the soil matrix. If this oxygen consumption exceeds the oxygen supply through diffusion along the oxygen gradient, then anoxic zones may form in the aggregate center (Figure 2f-g). Their extent depends on local respiration rates and contact distances. Hence, different microhabitats within short distances that allow for a high microbial biodiversity and diverse functional behavior in soil are not just an imprint of the soil structure per se, but evolve with the saturation-dependent distribution of air and water. Finally, plant roots and soil macrofauna like earthworms are able to “engineer” their own pore space and thus modify aeration and infiltration patterns in soil (Angers and Caron, 1998; Blouin et al., 2013; Bottinelli et al., 2015). They are known to also be affected by soil structure and water distribution through the restrictions imposed e.g. by mechanical resistivity and redox conditions (Bengough et al., 2011; Capowiez et al., 2009; Hamza and Anderson, 2005; Whalley et al., 1995).

The implications of these micro-environmental conditions for carbon and nitrogen turnover are manifold. Carbon mineralization rates through anaerobic respiration are about one order of magnitude smaller than through aerobic respiration (Keiluweit et al., 2017). This leads to the well-known saturation dependence of bulk soil respiration with an optimal respiration rate at intermediate soil moisture and a decline towards full saturation and complete dryness (Moyano et al., 2013; Skopp et al., 1990). Under very dry conditions a substrate diffusion limit arises, because the continuity in the water phase is lost and microbes become separated from resources and may fall into metabolic arrest (Manzoni and Katul, 2014; Tecon and Or, 2017). Under very wet conditions an oxygen diffusion limit may arise, when oxygen consumption exceeds the diffusive flux towards the location of oxygen consumption, so that anaerobic respiration with alternative electron acceptors sets in (Linn and Doran, 1984). The susceptibility of organic matter to mineralization therefore does not only depend on the chemical structure of the organic compounds, which defines the electron donor-dependent energy gain, but also on the moisture regime and resulting redox conditions which controls the electron acceptor-dependent energy gain of the reaction (Keiluweit et al., 2016).

Soil structure does not only have an indirect effect carbon turnover through the regulation of water retention and soil aeration, but also exerts a direct control on carbon stabilization, when organic compounds are located in pores that are not accessible to microorganisms. In fact, this physical protection seems to be the main mechanism for long-term carbon storage next to protection in organo-mineral associations, whereas the importance of chemical recalcitrance might have been overstated in the past (Lehmann and Kleber, 2015; Schmidt et al., 2011). This physical protection might occur in pores that are smaller than a microorganism itself, or when they are separated by discontinuous water films,

though this limitation is more relevant for bacteria than for fungi as they can overcome these barriers via hyphae (Ritz and Young, 2004). There are indications that this physical carbon protection is most relevant in fine-textured, structured soils, but less relevant in sandy soils (Christensen, 2001). There is an important feedback loop because soil structure does not only affect carbon turnover. Organic carbon is also a key driver in the formation of soil structure by acting directly as binding agent for mineral particles and indirectly by stimulating soil biota that modify soil structure (Rillig and Mummey, 2006; Six et al., 2004; Tisdall and Oades, 1982).

Soil structure is not only important for carbon turnover, but also for nutrient cycling, in particular nitrogen. Reactive nitrogen exists in many soluble and gaseous forms in soil. Transformations between them are regulated by environmental conditions (pH, temperature, moisture) and different transformations may occur simultaneously in different niches and microsites in structured soil. This is again demonstrated with the aggregate example in Figure 2. Denitrification, i.e. the reduction of nitrate through anaerobic respiration, will occur in anoxic aggregate centers. Nitrification, i.e. the biological oxidation of ammonium to nitrite and nitrate, in turn, is an aerobic process as it requires oxygen and will therefore occur in direct vicinity to air-filled pores. The relative importance of both processes depends on soil moisture. Nitrification rates increase with soil moisture in line with overall microbial activity until it drops when aeration becomes a limiting factor. Denitrification emerges at saturations around 60-70% and peaks around 90% (Linn and Doran, 1984). It does not only depend on the moisture-dependent diffusion distances in soil, but also on the distribution of organic carbon in soil. In a seminal study Parkin (1987) demonstrated that 25-85% of denitrification activity was associated with particulate organic matter that comprised less than 1% of the soil volume. The concentration of microbial activity in microbial hotspots is therefore not only immanent to carbon cycling but also to nitrogen cycling (Kuzakov and Blagodatskaya, 2015) and their distribution in space is controlled by soil structure.

1.4. Recent Advances in Pore-Scale Imaging of Soil Systems

Pore-scale imaging studies like the example above have developed into a powerful tool to gain more insights into the complex interactions between soil structure and soil functions in intact soils. Recent advances in pore scale imaging of soils have been achieved along four major avenues that we briefly outline here without any claim to cover the recent literature completely.

1.4.1. Development of new imaging protocols

In addition to continuous progresses in imaging hardware, new image processing protocols are also being developed constantly to improve the robustness, objectivity or accuracy in microstructure characterization. The best combination of methods for image enhancement, image segmentation and image analysis needs to be found again and again for each study in order to best extract the relevant structural information from the raw data. It is still common practice to segment a raw (or filtered) gray scale image into foreground and background with histogram-based thresholding methods even though it has been shown frequently that locally-adaptive methods that take some neighborhood information into account are often superior (Iassonov et al., 2009; Schlüter et al., 2014). For some imaging tasks like root detection it is helpful to extract features based on their shape in addition to gray values (Gao et al., 2019; Schulz et al., 2013), as the cylindrical shape of roots is unique among all materials in soil, whereas the gray value is not. Other useful properties for feature extraction may include texture and local heterogeneity of materials (Andrew, 2018; Schweizer et al., 2018). With density calibrated images, a mixing law has been used to determine the thresholds for void segmentation (Sammartino et al., 2012).

The next revolution in image segmentation and feature extraction is expected to be brought about by machine learning and deep learning techniques. This class of supervised techniques requires some level of user input in terms of training data but may lead to superior results by finding for every targeted material some characteristic patterns in a higher-dimensional feature space constituted by the gray value (and other properties) at the original resolution (and other hierarchy levels). Some current examples include root segmentation (Koebernick et al., 2017; Soltaninejad et al., 2019) and pore segmentation in natural porous media (Chauhan et al., 2016; Wang et al., 2020).

Some progress in studying dynamic processes has been achieved by repetitive imaging of the same sample at different temporal scales e.g. to study the evolution of an infiltration front or solute plume within hours (Haber-Pohlmeier et al., 2010; Koestel and Larsbo, 2014; Luo et al., 2008; Sammartino et al., 2015; Tötze et al., 2017) or the soil structure dynamics under natural conditions over years (Garbout et al., 2013; Koestel and Schlüter, 2019). Depending on the research question, image registration of subsequent scans might be required for a perfect spatial alignment of consecutive images, as the sample is typically moved in between image acquisitions. This is the case when soil deformation due to compaction (Peth et al., 2010; Schlüter et al., 2016; Watanabe et al., 2012) or due to bioturbation (Koestel and Schlüter, 2019) is of interest.

The appeal of a method does not only depend on its performance but also on its availability. Therefore, the merit of developing and maintaining publicly available plugins and packages (Koestel,

2018; Legland et al., 2016; van der Walt et al., 2014) for popular imaging software like ImageJ or scikit-image cannot be appreciated enough.

1.4.2. Linking different techniques with correlative imaging

Any imaging technique is selective in the local property that is mapped and thus the kind of information which can be retrieved. For a more comprehensive view on microenvironments in soil it is appealing to combine various imaging methods through a correlative imaging approach. For instance, plant-soil interactions can be understood better with a combination of X-ray tomography to map the spatial distribution of solids and pores, neutron tomography to map water content and water uptake and magnetic resonance imaging to detect the release of gel-like root exudates in the rhizosphere (Haber-Pohlmeier et al., 2019). The method by which all images are spatially aligned with each other is called image registration or co-registration. The combination of two-dimensional microscopy data through image registration, also denoted as correlative microscopy, facilitates the combination of biological and chemical information from a whole suite of microscopic and micro-spectroscopic techniques (Baveye et al., 2018). The next frontier is to combine this biochemical information retrieved from exposed surfaces with three-dimensional, structural information of the intact sample through 2D-3D image registration (Juyal et al., 2019; Kravchenko et al., 2019b; Schlüter et al., 2019b). The required image registration is challenging due to different dimensionality and scale but also very different image content. Nevertheless, this methodological approach will progress quickly as it enables a more holistic view on microenvironments in soil that combines biotic with abiotic information.

1.4.3. Relating microstructure to soil functioning

Pore scale imaging and image analysis are now being routinely used to study a wide range of soil properties for a better understanding of soil functions summarized in Figure 1. To this category we would also add studies that investigate reverse effects, i.e. changes in soil structure due to a specific process, be it natural or management induced. Meanwhile, there is a large number of respective studies published in peer-reviewed, scientific literature. These pore-scale imaging studies are mostly carried out on targeted laboratory experiments or field sampling campaigns from selected sites in order to address a specific research question with a limited set of samples. This is a valid approach for gaining basic process understanding, but generalizations beyond the specific experimental setup are often not warranted, i.e. different behavior for different soil type, land use or environmental conditions.

In general, there is huge potential in collecting data from many studies with similar objectives in large databases to reveal general trends, dependencies or tipping points (Jorda et al., 2015; Rillig et al., 2019). Based on regression analyses with a multitude of samples so-called pedo-transfer functions can be developed to estimate some emergent soil behavior such as hydraulic conductivity (Van Looy et al., 2017) or soil respiration (Moyano et al., 2012) from a set of readily available parameters (e.g. bulk density, soil organic matter content, clay content, etc). Image-derived pore space properties have so far not been incorporated systematically for this kind of data mining due to the limited number of studies in which the targeted emergent behavior and pore space properties are measured jointly. The need to establish such databases which include microstructural properties has recently been stressed (Fatichi et al., 2020; Rabot et al., 2018; Zhang and Schaap, 2019) and the studies that investigate e.g. the impact of macropore structure on soil hydraulic conductivity are converging towards a small set of promising image-derived pore metrics that best predict flow (Koestel et al., 2018; Müller et al., 2018; Paradelo et al., 2016; Schlüter et al., 2019a; Zhang et al., 2019).

1.4.4. Combine pore scale imaging with modelling

There are two major avenues along which the combination of pore scale imaging with modelling can be beneficial.

First, a pore scale image can directly serve as a model domain for spatially resolved models. The modeling result can then be compared directly to measurements, when carried out on the same domain, e.g. saturated and near-saturated flow (Schlüter et al., 2019a), to assess measurement artifacts or the validity of assumption made during modelling. More often, direct measurements are impossible to obtain due to the small size of investigated volumes or a general lack of suitable techniques to probe a given property. In that case, image-based models are still extremely useful for explorative modelling. With this type of “what if” modelling, different scenarios of a property that cannot be readily measured are constructed and the effect on emergent system behavior is explored. Some recent examples for this hypothesis testing comprise the effect of substrate distribution within image-derived pore space geometries on biodegradation and biodiversity (Portell et al., 2018) or the effect of root hairs and local water saturation in the rhizosphere on root water uptake (Daly et al., 2016).

Second, image-derived parameters can help to constrain parameters of continuum-scale models operating on larger scales. The success of this approach depends on whether the often empirical parameters actually depend on these independently measured, microstructural properties. Sammartino

et al. (2015) derived the active macropore network of a macroporous soil using analysis of CT-images and a validation by brilliant blue staining of the preferential flow pathways. They demonstrated the strong link between the active macropore network and the percolating backbone. In a similar approach Gerke (2012) estimated the interface between macropores and the soil matrix using X-ray CT-images. Indeed, this inter-domain area is a very important parameter as it controls most of the mass exchange processes during infiltration. With some exceptions also in this virtual special issue (Haas et al., 2020; Soto-Gómez et al., 2020), analyses of CT images towards geometric parameters associated with the active macropore network has not been adopted much, but are considered very promising to make models less empirical and more physically-based.

2. Contents of the Virtual Special Issue

In the introduction we have outlined how soil structure governs soil functions and the potential of pore scale imaging to explore this relationship. This virtual special issue entitled “Recent advances in pore scale imaging of soil systems” presents fourteen contributions that applied pore scale imaging for a whole range of objectives. They can be structured into three thematic blocks.

2.1. Method development

One block of contributions comprises studies on methodological aspects of pore scale imaging. Torre et al. (2020) generated more realistic, synthetic test images with the truncated multi-fractal method that share common properties with X-ray CT images of real soil. Therefore, these test images do not only possess ground truth information required to validate the performance of segmentation methods per se, they are also more likely to indicate the failure of a segmentation method when X-ray CT scans of real soil are segmented. This is demonstrated by superior segmentation results of a locally-adaptive method over a global thresholding method.

Pot et al. (2020) explored the impact of choices made during different image processing steps with respect to modeled saturated hydraulic conductivity (K_{sat}) obtained with image-based Lattice-Boltzmann simulations. They showed that K_{sat} can change by two orders of magnitude, which is mainly dependent on how pore necks are segmented. Moreover, the Kozeny Carman equation based on image-derived properties was able to explain the variation in K_{sat} .

Haas et al. (2020) developed a method that is potentially able to derive the mass exchange parameter in two-domain solute transport models from independent, image-derived metrics based on the small-scale distribution of a fluorescent dye mapped on a 2-D cross-section of a soil aggregates, using digital photography in the visible light range.. The dye redistribution was traced with time-lapse imaging which serves as a proxy for the movement of adsorbing chemicals.

2.2. Soil structure dynamics

The second block of papers looks into soil structure dynamics in the laboratory or under natural conditions induced by various structure-forming processes.

Diel et al. (2019) investigated pore structure changes through crack formation and closure during wetting/drying cycles for repacked soils with different texture, organic matter content and clay mineralogy. A higher proportion of clay minerals with swell/shrinking capacity induced stronger crack formation, whereas a higher organic matter content lead to a higher density of cracks with smaller aperture. By means of a structure labeling approach with small garnet particles they showed that thorough soil structure dynamics cannot be induced by wetting/drying cycles as the cracks that are formed in the first cycles are reused in subsequent cycles.

Pires et al. (2020) also looked into soil structure changes induced by wetting/drying cycles and their effect on water retention curves measured with suction tables; an often overlooked effect that may have a considerable impact on the actual measurement. Measured water retention curves were affected by a series of up to twelve wetting/drying cycles which was explained by internal pore structure changes in terms of pore tortuosity and pore connectivity especially in the hydraulic contact region between the samples and the suction table.

Koestel and Schlüter (2019) investigated soil structure dynamics under natural conditions by time lapse imaging of an in-growth core that was repeatedly reburied in a garden soil. With this proof-of-concept study they demonstrated the temporal variability of pore space metrics induced by different biotic and abiotic process like soil settling, root growth or faunal activity. Through deformation analysis they showed the much larger extend of lateral compaction by taproot growth as compared to macro-faunal burrowing.

Lucas et al. (2019) examined soil structure formation in a 24 year “space-for-time” agricultural chronosequence established for reclamation of a lignite mining area. With a new biopore segmentation method they were able to distinguish root channels from other structural pores. Biopore density had reached equilibrium after twelve years underneath the plow layer, but only 10% of these channels are actually filled with roots each year. This study shows how fast plant roots create a stable and connected biopore system and how this is disrupted by soil tillage, which produces completely contrasting pore characteristics.

Ferreira et al. (2019) scrutinized the effect of liming on soil aggregation and pore structure in an acidic silty-clay, no-till topsoil 30 months after lime application. Only in those layers where liming affected chemical soil properties (pH, base saturation) could an increase in macroporosity with more elongated and better connected pores be observed. Increased biopore formation was observed as a secondary effect of liming.

2.3. Relationship between soil structure and soil functions

Schlüter et al. (2020) compared topsoil samples under conventional tillage and no-till with respect to saturated and near-saturated hydraulic conductivity measured with hood infiltrometers in the field, tension disk infiltrometers on intact soil cores and simulated with 3D image-based flow simulations. No-till soils exhibit higher, lower or indifferent hydraulic conductivities depending on the tension and technique. Moreover, the pore metrics that best predicted conductivity also changed with tension and technique. The critical pore diameter had very high predictive power on saturated hydraulic conductivity when simulated on no-till soils with large earthworm burrows in an otherwise compact soil matrix. In plowed soils macroporosity and pore connectivity best describe flow through the loosened soil matrix.

Piccoli et al. (2019) explored the effect of soil structure on gas transport properties with a special focus on scale effects. They showed that the bigger the volume of intact samples, the more dominant are large, continuous biopores that evoke more efficient gas transport. The smallest considered sample size (4.7 cm³) the pore network was dominated by small isolated air-filled pores that obstruct gas diffusion at field capacity (-100 hPa). While some microstructural properties showed indications for scale invariance in the investigated range of sample sizes and image resolutions, emergent system behavior in terms of gas transport is clearly not. Thus, sample size needs to be accounted for when comparing gas transport properties between studies.

Koestel et al. (2020) evaluated representative elementary volumes (REVs) for the X-ray-derived porosity and pore-connectivity in undisturbed soil columns (67 mm diameter, 60 mm height). These pore-space features are intrinsically linked to water retention and soil hydraulic properties. The existence of such REVs would support the adequacy of respective continuum-scale models. Koestel et al. (2020) detected a range of observation scales, between 15 and 65 mm, for which the mean porosity was scale-independent. Due to a lack in statistical homogeneity, it remained however unclear whether porosity REVs existed. The authors found no potential REVs for the pore-network connectivity. They recommend future studies on larger soil samples and a re-evaluation of the required of REV criteria in a continuum modelling context.

Soto-Gómez et al. (2020) investigated solute transport and colloid transport in intact soil cores and were able to show correlations between transport properties and some metrics of the percolating pore network. Moreover, the area of pore walls stained by fluorescent colloids correlated with the image-derived metrics of the percolating pore space. The morphology of the percolating backbone showed huge differences between no-till, shallow till and organic plots with high and low earthworm activity on a sandy loam.

Mawodza et al. (2020) applied neutron tomography to visualize wheat root growth and soil moisture distribution in aggregated soil. They demonstrated that in heterogeneous soil the emergence and growth of lateral roots depends on local soil structures. Moreover, local water contents were higher in larger soil aggregates suggesting that these may be able to provide the plants with water during periodic dry spells.

Menon et al. (2020) explored links between the pore space properties and the stability of soil aggregates. They could show that land use has a significant influence on both the pore size distribution and the fraction of water-stable aggregates. The pore system of stable aggregates did not undergo significant changes upon continued submergence in water, indicating that a stable pore system is crucial for aggregate stability. Supposedly, a stable pore system facilitates the transmission of fluids without trapping the air and thereby suppressing the build-up of air pressure inside an aggregate preventing it from slaking.

Acknowledgements

The guest editors like to thank Geoderma, especially Yvan Capowiez and Jan Willem van Groenigen, for the invitation to manage this virtual special issue and express their deep gratitude to all contributing authors and anonymous reviewers for their share in producing a very interesting selection of papers.

3. References

- Ad-hoc-AG Boden, 2005. Bodenkundliche Kartieranleitung (KA 5). Schweizerbart, Stuttgart.
- Almajmaie, A., Hardie, M., Acuna, T., Birch, C., 2017. Evaluation of methods for determining soil aggregate stability. *Soil and Tillage Research* 167, 39-45.
- Andrew, M., 2018. A quantified study of segmentation techniques on synthetic geological XRM and FIB-SEM images. *Computational Geosciences* 22(6), 1503-1512.
- Angers, D.A., Caron, J., 1998. Plant-induced Changes in Soil Structure: Processes and Feedbacks. *Biogeochemistry* 42(1), 55-72.
- Angers, D.A., Recous, S., Aita, C., 1997. Fate of carbon and nitrogen in water-stable aggregates during decomposition of ¹³C¹⁵N-labelled wheat straw in situ. *European Journal of Soil Science* 48(2), 295-300.
- Ball, B.C., Batey, T., Munkholm, L.J., 2007. Field assessment of soil structural quality – a development of the Peerlkamp test. *Soil Use and Management* 23(4), 329-337.
- Baveye, P., 2006. Comment on "Soil structure and management: A review" by CJ Bronick and R. Lal. *Geoderma* 134(1-2), 231-232.
- Baveye, P.C., Laba, M., Otten, W., Bouckaert, L., Sterpaio, P.D., Goswami, R.R., Grinev, D., Houston, A., Hu, Y., Liu, J., Mooney, S., Pajor, R., Sleutel, S., Tarquis, A., Wang, W., Wei, Q., Sezgin, M., 2010. Observer-dependent variability of the thresholding step in the quantitative analysis of soil images and X-ray microtomography data. *Geoderma* 157(1-2), 51-63.
- Baveye, P.C., Otten, W., Kravchenko, A., Balseiro-Romero, M., Beckers, É., Chalhoub, M., Darnault, C., Eickhorst, T., Garnier, P., Hapca, S., Kiranyaz, S., Monga, O., Mueller, C.W., Nunan, N., Pot, V., Schlüter, S., Schmidt, H., Vogel, H.-J., 2018. Emergent Properties of Microbial Activity in Heterogeneous Soil Microenvironments: Different Research Approaches Are Slowly Converging, Yet Major Challenges Remain. *Frontiers in Microbiology* 9(1929), 1-48.
- Beare, M.H., Bruce, R.R., 1993. A comparison of methods for measuring water-stable aggregates: implications for determining environmental effects on soil structure. *Geoderma* 56(1-4), 87-104.
- Bengough, A.G., McKenzie, B.M., Hallett, P.D., Valentine, T.A., 2011. Root elongation, water stress, and mechanical impedance: a review of limiting stresses and beneficial root tip traits. *Journal of Experimental Botany* 62(1), 59-68.
- Blouin, M., Hodson, M.E., Delgado, E.A., Baker, G., Brussaard, L., Butt, K.R., Dai, J., Dendooven, L., Peres, G., Tondoh, J.E., Cluzeau, D., Brun, J.-J., 2013. A review of earthworm impact on soil function and ecosystem services. *European Journal of Soil Science* 64(2), 161-182.
- Bottinelli, N., Jouquet, P., Capowiez, Y., Podwojewski, P., Grimaldi, M., Peng, X., 2015. Why is the influence of soil macrofauna on soil structure only considered by soil ecologists? *Soil and Tillage Research* 146, 118-124.
- Capowiez, Y., Cadoux, S., Bouchant, P., Ruy, S., Roger-Estrade, J., Richard, G., Boizard, H., 2009. The effect of tillage type and cropping system on earthworm communities, macroporosity and water infiltration. *Soil and Tillage Research* 105(2), 209-216.

- Chauhan, S., Rühaak, W., Khan, F., Enzmann, F., Mielke, P., Kersten, M., Sass, I., 2016. Processing of rock core microtomography images: Using seven different machine learning algorithms. *Computers & Geosciences* 86, 120-128.
- Christensen, B.T., 2001. Physical fractionation of soil and structural and functional complexity in organic matter turnover. *European Journal of Soil Science* 52(3), 345-353.
- Daly, K.R., Keyes, S.D., Masum, S., Roose, T., 2016. Image-based modelling of nutrient movement in and around the rhizosphere. *Journal of Experimental Botany* 67(4), 1059-1070.
- Díaz-Zorita, M., Perfect, E., Grove, J.H., 2002. Disruptive methods for assessing soil structure. *Soil and Tillage Research* 64(1), 3-22.
- Diel, J., Vogel, H.-J., Schlüter, S., 2019. Impact of wetting and drying cycles on soil structure dynamics. *Geoderma* 345, 63-71.
- Fatichi, S., Or, D., Walko, R., Vereecken, H., Young, M.H., Ghezzehei, T.A., Hengl, T., Kollet, S., Agam, N., Avissar, R., 2020. Soil structure is an important omission in Earth System Models. *Nature Communications* 11(1), 522.
- Ferreira, T.R., Pires, L.F., Wildenschild, D., Brinatti, A.M., Borges, J.A.R., Auler, A.C., dos Reis, A.M.H., 2019. Lime application effects on soil aggregate properties: Use of the mean weight diameter and synchrotron-based X-ray μ CT techniques. *Geoderma* 338, 585-596.
- Gao, W., Schlüter, S., Blaser, S.R.G.A., Shen, J., Vetterlein, D., 2019. A shape-based method for automatic and rapid segmentation of roots in soil from X-ray computed tomography images: Routine. *Plant and Soil*.
- Garbout, A., Munkholm, L.J., Hansen, S.B., 2013. Temporal dynamics for soil aggregates determined using X-ray CT scanning. *Geoderma* 204-205, 15-22.
- Gerke, H.H., 2012. Macroscopic Representation of the Interface between Flow Domains in Structured Soil. *Vadose Zone Journal* 11(3).
- Guimarães, R.M.L., Ball, B.C., Tormena, C.A., 2011. Improvements in the visual evaluation of soil structure. *Soil Use and Management* 27(3), 395-403.
- Guimarães, R.M.L., Ball, B.C., Tormena, C.A., Giarola, N.F.B., da Silva, Á.P., 2013. Relating visual evaluation of soil structure to other physical properties in soils of contrasting texture and management. *Soil and Tillage Research* 127, 92-99.
- Haas, C., Horn, R., Ellerbrock, R.H., Gerke, H.H., 2020. Fluorescence imaging for mm-scale observation of macropore-matrix mass transfer: Calibration experiments. *Geoderma* 360, 114002.
- Haber-Pohlmeier, S., Bechtold, M., Stapf, S., Pohlmeier, A., 2010. Water Flow Monitored by Tracer Transport in Natural Porous Media Using Magnetic Resonance Imaging. *Vadose Zone Journal* 9(4), 835-845.
- Haber-Pohlmeier, S., Tötze, C., Lehmann, E., Kardjilov, N., Pohlmeier, A., Oswald, S.E., 2019. Combination of Magnetic Resonance Imaging and Neutron Computed Tomography for Three-Dimensional Rhizosphere Imaging. *Vadose Zone Journal* 18(1).
- Hamza, M.A., Anderson, W.K., 2005. Soil compaction in cropping systems - A review of the nature, causes and possible solutions. *Soil & Tillage Research* 82, 121-145.
- Hazlett, R.D., 1995. Simulation of capillary-dominated displacements in microtomographic images of reservoir rocks. *Transport in Porous Media* 20(1-2), 21-35.
- Hilpert, M., Miller, C.T., 2001. Pore-morphology-based simulation of drainage in totally wetting porous media. *Advances in Water Resources* 24(3-4), 243-255.
- Iassonov, P., Gebrenegus, T., Tuller, M., 2009. Segmentation of X-Ray CT Images of Porous Materials: A Crucial Step for Characterization and Quantitative Analysis of Pore Structures. *Water Resources Research* 45, W09415.
- Jahn, R., Blume, H., Asio, V., Spaargaren, O., Schad, P., 2006. Guidelines for soil description. 4th ed. FAO, Rome, Italy.

516 Jorda, H., Bechtold, M., Jarvis, N., Koestel, J., 2015. Using boosted regression trees to explore key factors
 517 controlling saturated and near-saturated hydraulic conductivity. *European Journal of Soil*
 518 *Science* 66(4), 744-756.

519 Juyal, A., Otten, W., Falconer, R., Hapca, S., Schmidt, H., Baveye, P.C., Eickhorst, T., 2019. Combination of
 520 techniques to quantify the distribution of bacteria in their soil microhabitats at different spatial
 521 scales. *Geoderma* 334, 165-174.

522 Keiluweit, M., Nico, P.S., Kleber, M., Fendorf, S., 2016. Are oxygen limitations under recognized
 523 regulators of organic carbon turnover in upland soils? *Biogeochemistry* 127(2), 157-171.

524 Keiluweit, M., Wanzek, T., Kleber, M., Nico, P., Fendorf, S., 2017. Anaerobic microsites have an
 525 unaccounted role in soil carbon stabilization. *Nature Communications* 8(1), 1771.

526 Kibblewhite, M.G., Ritz, K., Swift, M.J., 2008. Soil health in agricultural systems. *Philosophical*
 527 *Transactions of the Royal Society B: Biological Sciences* 363(1492), 685-701.

528 Koebernick, N., Daly, K.R., Keyes, S.D., George, T.S., Brown, L.K., Raffan, A., Cooper, L.J., Naveed, M.,
 529 Bengough, A.G., Sinclair, I., Hallett, P.D., Roose, T., 2017. High-resolution synchrotron imaging
 530 shows that root hairs influence rhizosphere soil structure formation. *New Phytol* 216(1), 124-
 531 135.

532 Koestel, J., 2018. SoilJ: An ImageJ Plugin for the Semiautomatic Processing of Three-Dimensional X-ray
 533 Images of Soils. *Vadose Zone Journal* 17(1).

534 Koestel, J., Dathe, A., Skaggs, T.H., Klakegg, O., Ahmad, M.A., Babko, M., Giménez, D., Farkas, C., Nemes,
 535 A., Jarvis, N., 2018. Estimating the Permeability of Naturally Structured Soil From Percolation
 536 Theory and Pore Space Characteristics Imaged by X-Ray. *Water Resources Research* 54(11),
 537 9255-9263.

538 Koestel, J., Larsbo, M., 2014. Imaging and quantification of preferential solute transport in soil
 539 macropores. *Water Resources Research* 50(5), 4357-4378.

540 Koestel, J., Larsbo, M., Jarvis, N., 2020. Scale and REV analyses for porosity and pore connectivity
 541 measures in undisturbed soil. *Geoderma* 366, 114206.

542 Koestel, J., Schlüter, S., 2019. Quantification of the structure evolution in a garden soil over the course of
 543 two years. *Geoderma* 338, 597-609.

544 Kravchenko, A., Otten, W., Garnier, P., Pot, V., Baveye, P.C., 2019a. Soil aggregates as biogeochemical
 545 reactors: Not a way forward in the research on soil-atmosphere exchange of greenhouse gases.
 546 *Global Change Biology* 25(7), 2205-2208.

547 Kravchenko, A.N., Guber, A.K., 2017. Soil pores and their contributions to soil carbon processes.
 548 *Geoderma* 287, 31-39.

549 Kravchenko, A.N., Guber, A.K., Razavi, B.S., Koestel, J., Blagodatskaya, E.V., Kuzyakov, Y., 2019b. Spatial
 550 patterns of extracellular enzymes: Combining X-ray computed micro-tomography and 2D
 551 zymography. *Soil Biology and Biochemistry* 135, 411-419.

552 Kuzyakov, Y., Blagodatskaya, E., 2015. Microbial hotspots and hot moments in soil: Concept & review.
 553 *Soil Biology and Biochemistry* 83, 184-199.

554 Legland, D., Arganda-Carreras, I., Andrey, P., 2016. MorphoLibJ: integrated library and plugins for
 555 mathematical morphology with ImageJ. *Bioinformatics* 32(22), 3532-3534.

556 Lehmann, J., Kleber, M., 2015. The contentious nature of soil organic matter. *Nature* 528, 60.

557 Letey, J., 1991. The study of soil structure - Science or art. *Soil Research* 29(6), 699-707.

558 Linn, D., Doran, J., 1984. Effect of water-filled pore space on carbon dioxide and nitrous oxide
 559 production in tilled and nontilled soils. *Soil Science Society of America Journal* 48(6), 1267-1272.

560 Lucas, M., Schlüter, S., Vogel, H.-J., Vetterlein, D., 2019. Soil structure formation along an agricultural
 561 chronosequence. *Geoderma* 350, 61-72.

562 Luo, L., Lin, H., Halleck, P., 2008. Quantifying soil structure and preferential flow in intact soil using X-ray
 563 computed tomography. *Soil Science Society of America Journal* 72(4), 1058-1069.

- Manzoni, S., Katul, G., 2014. Invariant soil water potential at zero microbial respiration explained by hydrological discontinuity in dry soils. *Geophysical Research Letters* 41(20), 7151-7158.
- Mawodza, T., Burca, G., Casson, S., Menon, M., 2020. Wheat root system architecture and soil moisture distribution in an aggregated soil using neutron computed tomography. *Geoderma* 359, 113988.
- McKenzie, D.C., 2001. Rapid assessment of soil compaction damage I. The SOILpak score, a semi-quantitative measure of soil structural form. *Soil Research* 39(1), 117-125.
- Menon, M., Mawodza, T., Rabbani, A., Bland, A., Lair, G.J., Babaei, M., Kercheva, M., Rousseva, S., Banwart, S., 2020. Pore system characteristics of soil aggregates and their relevance to aggregate stability. *Geoderma* 366, 114259.
- Moyano, F.E., Manzoni, S., Chenu, C., 2013. Responses of soil heterotrophic respiration to moisture availability: An exploration of processes and models. *Soil Biology and Biochemistry* 59, 72-85.
- Moyano, F.E., Vasilyeva, N., Bouckaert, L., Cook, F., Craine, J., Curiel Yuste, J., Don, A., Epron, D., Formanek, P., Franzluebbers, A., Ilstedt, U., Kätterer, T., Orchard, V., Reichstein, M., Rey, A., Ruamps, L., Subke, J.A., Thomsen, I.K., Chenu, C., 2012. The moisture response of soil heterotrophic respiration: interaction with soil properties. *Biogeosciences* 9(3), 1173-1182.
- Mueller, L., Kay, B.D., Hu, C., Li, Y., Schindler, U., Behrendt, A., Shepherd, T.G., Ball, B.C., 2009. Visual assessment of soil structure: Evaluation of methodologies on sites in Canada, China and Germany: Part I: Comparing visual methods and linking them with soil physical data and grain yield of cereals. *Soil and Tillage Research* 103(1), 178-187.
- Müller, K., Katuwal, S., Young, I., McLeod, M., Moldrup, P., de Jonge, L.W., Clothier, B., 2018. Characterising and linking X-ray CT derived macroporosity parameters to infiltration in soils with contrasting structures. *Geoderma* 313, 82-91.
- Newell-Price, J.P., Whittingham, M.J., Chambers, B.J., Peel, S., 2013. Visual soil evaluation in relation to measured soil physical properties in a survey of grassland soil compaction in England and Wales. *Soil and Tillage Research* 127, 65-73.
- Page-Dumroese, D.S., Brown, R.E., Jurgensen, M.F., Mroz, G.D., 1999. Comparison of Methods for Determining Bulk Densities of Rocky Forest Soils. *Soil Science Society of America Journal* 63(2), 379-383.
- Pagliai, M., Vignozzi, N., 2002. The soil pore system as an indicator of soil quality. *Advances in Geoecology* 34, 69-80.
- Paradelo, M., Katuwal, S., Moldrup, P., Norgaard, T., Herath, L., de Jonge, L.W., 2016. X-ray CT-derived soil characteristics explain varying air, water, and solute transport properties across a loamy field. *Vadose Zone Journal* 15(4).
- Parkin, T.B., 1987. Soil microsites as a source of denitrification variability. *Soil Science Society of America Journal* 51(5), 1194-1199.
- Peth, S., Nellesen, J., Fischer, G., Horn, R., 2010. Non-invasive 3D analysis of local soil deformation under mechanical and hydraulic stresses by μ CT and digital image correlation. *Soil and Tillage Research* 111(1), 3-18.
- Piccoli, I., Schjønning, P., Lamandé, M., Zanini, F., Morari, F., 2019. Coupling gas transport measurements and X-ray tomography scans for multiscale analysis in silty soils. *Geoderma* 338, 576-584.
- Pires, L.F., Auler, A.C., Roque, W.L., Mooney, S.J., 2020. X-ray microtomography analysis of soil pore structure dynamics under wetting and drying cycles. *Geoderma* 362, 114103.
- Portell, X., Pot, V., Garnier, P., Otten, W., Baveye, P.C., 2018. Microscale Heterogeneity of the Spatial Distribution of Organic Matter Can Promote Bacterial Biodiversity in Soils: Insights From Computer Simulations. *Frontiers in microbiology* 9, 1583-1583.

- Pot, V., Zhong, X., Baveye, P.C., 2020. Effect of resolution, reconstruction settings, and segmentation methods on the numerical calculation of saturated soil hydraulic conductivity from 3D computed tomography images. *Geoderma* 362, 114089.
- Pulido Moncada, M., Gabriels, D., Lobo, D., Rey, J.C., Cornelis, W.M., 2014. Visual field assessment of soil structural quality in tropical soils. *Soil and Tillage Research* 139, 8-18.
- Rabot, E., Wiesmeier, M., Schlüter, S., Vogel, H.J., 2018. Soil structure as an indicator of soil functions: A review. *Geoderma* 314, 122-137.
- Rillig, M.C., Mummey, D.L., 2006. Mycorrhizas and soil structure. *New Phytologist* 171(1), 41-53.
- Rillig, M.C., Ryo, M., Lehmann, A., Aguilar-Trigueros, C.A., Buchert, S., Wulf, A., Iwasaki, A., Roy, J., Yang, G., 2019. The role of multiple global change factors in driving soil functions and microbial biodiversity. *Science* 366(6467), 886-890.
- Ritz, K., Young, I.M., 2004. Interactions between soil structure and fungi. *Mycologist* 18, 52-59.
- Sammartino, S., Lissy, A.-S., Bogner, C., Van Den Bogaert, R., Capowicz, Y., Ruy, S., Cornu, S., 2015. Identifying the Functional Macropore Network Related to Preferential Flow in Structured Soils. *Vadose Zone Journal* 14(10).
- Sammartino, S., Michel, E., Capowicz, Y., 2012. A Novel Method to Visualize and Characterize Preferential Flow in Undisturbed Soil Cores by Using Multislice Helical CT. *Vadose Zone Journal* 11(1).
- Schlüter, S., Albrecht, L., Schwärzel, K., Kreiselmeier, J., 2019a. Long-term effects of conventional tillage and no-tillage on saturated and near-saturated hydraulic conductivity – Can their prediction be improved by pore metrics obtained with X-ray CT? *Geoderma*, 114082.
- Schlüter, S., Albrecht, L., Schwärzel, K., Kreiselmeier, J., 2020. Long-term effects of conventional tillage and no-tillage on saturated and near-saturated hydraulic conductivity – Can their prediction be improved by pore metrics obtained with X-ray CT? *Geoderma* 361, 114082.
- Schlüter, S., Eickhorst, T., Mueller, C.W., 2019b. Correlative Imaging Reveals Holistic View of Soil Microenvironments. *Environmental Science & Technology* 53(2), 829-837.
- Schlüter, S., Leuther, F., Vogler, S., Vogel, H.-J., 2016. X-ray microtomography analysis of soil structure deformation caused by centrifugation. *Solid Earth* 7(1), 129-140.
- Schlüter, S., Sheppard, A., Brown, K., Wildenschild, D., 2014. Image processing of multiphase images obtained via X-ray microtomography: A review. *Water Resources Research* 50(4), 3615-3639.
- Schlüter, S., Weller, U., Vogel, H.-J., 2011. Soil-structure development including seasonal dynamics in a long-term fertilization experiment. *Journal of Plant Nutrition and Soil Science* 174(3), 395-403.
- Schmidt, M.W.I., Torn, M.S., Abiven, S., Dittmar, T., Guggenberger, G., Janssens, I.A., Kleber, M., Kögel-Knabner, I., Lehmann, J., Manning, D.A.C., Nannipieri, P., Rasse, D.P., Weiner, S., Trumbore, S.E., 2011. Persistence of soil organic matter as an ecosystem property. *Nature* 478(7367), 49-56.
- Schulz, H., Postma, J.A., van Dusschoten, D., Scharr, H., Behnke, S., 2013. Plant Root System Analysis from MRI Images. In: G. Csurka, M. Kraus, R.S. Laramée, P. Richard, J. Braz (Eds.), *Computer Vision, Imaging and Computer Graphics. Theory and Application: 7th International Joint Conference, VISIGRAPP 2012, Rome, Italy, February 24-26, 2012, Revised Selected Papers*. Springer Berlin Heidelberg, Berlin, Heidelberg, pp. 411-425.
- Schweizer, S., A., Hoeschen, C., Schlüter, S., Kögel-Knabner, I., Mueller, C., W., 2018. Rapid soil formation after glacial retreat shaped by spatial patterns of organic matter accrual in microaggregates. *Global Change Biology* 24(4), 1637-1650.
- Shepherd, G., Stagnari, F., Pisante, M., Benites, J., 2008. Visual Soil Assessment–Field guide for annual crops. FAO, Rome, Italy.
- Shepherd, T.G., 2003. Assessing soil quality using visual soil assessment. In: L.D. Currie, J.A. Hanly (Eds.), *Tools for nutrient and pollutant management: : Applications to agriculture and environmental quality*. Fertilizer and Lime Research Centre, Massey University, Palmerston North, pp. 153-166.

- Six, J., Bossuyt, H., de Gryze, S., Denef, K., 2004. A history of research on the link between (micro)aggregates, soil biota, and soil organic matter dynamics. *Soil & Tillage Research* 79, 7-31.
- Skopp, J., Jawson, M.D., Doran, J.W., 1990. Steady-State Aerobic Microbial Activity as a Function of Soil Water Content. *Soil Science Society of America Journal* 54(6), 1619-1625.
- Soltaninejad, M., Sturrock, C.J., Griffiths, M., Pridmore, T.P., Pound, M.P., 2019. Three Dimensional Root CT Segmentation using Multi-Resolution Encoder-Decoder Networks. *bioRxiv*, 713859.
- Soto-Gómez, D., Vázquez Juárez, L., Pérez-Rodríguez, P., López-Periago, J.E., Paradelo, M., Koestel, J., 2020. Percolation theory applied to soil tomography. *Geoderma* 357, 113959.
- Tecon, R., Or, D., 2017. Biophysical processes supporting the diversity of microbial life in soil. *FEMS Microbiology Reviews* 41(5), 599-623.
- Tisdall, J.M., Oades, J.M., 1982. Organic matter and water-stable aggregates in soils. *European Journal of Soil Science* 33(2), 141-163.
- Torre, I.G., Martín-Sotoca, J.J., Losada, J.C., López, P., Tarquis, A.M., 2020. Scaling properties of binary and greyscale images in the context of X-ray soil tomography. *Geoderma* 365, 114205.
- Tötze, C., Kardjilov, N., Manke, I., Oswald, S.E., 2017. Capturing 3D Water Flow in Rooted Soil by Ultra-fast Neutron Tomography. *Scientific Reports* 7(1), 6192.
- van der Walt, S., Schönberger, J.L., Nunez-Iglesias, J., Boulogne, F.c., Warner, J.D., Yager, N., Gouillart, E., Yu, T., 2014. scikit-image: Image processing in Python. *PeerJ PrePrints* 2, e336v332.
- Van Looy, K., Bouma, J., Herbst, M., Koestel, J., Minasny, B., Mishra, U., Montzka, C., Nemes, A., Pachepsky, Y.A., Padarian, J., Schaap, M.G., Tóth, B., Verhoef, A., Vanderborght, J., van der Ploeg, M.J., Weihermüller, L., Zacharias, S., Zhang, Y., Vereecken, H., 2017. Pedotransfer Functions in Earth System Science: Challenges and Perspectives. *Reviews of Geophysics* 55(4), 1199-1256.
- Vincent, K.R., Chadwick, O.A., 1994. Synthesizing Bulk Density for Soils with Abundant Rock Fragments. *Soil Science Society of America Journal* 58(2), 455-464.
- Wang, B., Brewer, P.E., Shugart, H.H., Lerda, M.T., Allison, S.D., 2019. Building bottom-up aggregate-based models (ABMs) in soil systems with a view of aggregates as biogeochemical reactors. *Global Change Biology* 25(8), e6-e8.
- Wang, Y.D., Armstrong, R.T., Mostaghimi, P., 2020. Boosting Resolution and Recovering Texture of 2D and 3D Micro-CT Images with Deep Learning. *Water Resources Research* 56(1), e2019WR026052.
- Watanabe, Y., Lenoir, N., Otani, J., Nakai, T., 2012. Displacement in sand under triaxial compression by tracking soil particles on X-ray CT data. *Soils and Foundations* 52(2), 312-320.
- Whalley, W.R., Dumitru, E., Dexter, A.R., 1995. Biological effects of soil compaction. *Soil and Tillage Research* 35(1), 53-68.
- Young, I.M., Crawford, J.W., 2004. Interactions and Self-Organization in the Soil-Microbe Complex. *Science* 304, 1634-1637.
- Young, I.M., Crawford, J.W., Rappoldt, C., 2001. New methods and models for characterising structural heterogeneity of soil. *Soil & Tillage Research* 61, 33-45.
- Yudina, A., Kuzyakov, Y., 2019. Saving the face of soil aggregates. *Global Change Biology* 25(11), 3574-3577.
- Zhang, Y., Schaap, M.G., 2019. Estimation of saturated hydraulic conductivity with pedotransfer functions: A review. *Journal of Hydrology* 575, 1011-1030.
- Zhang, Z., Liu, K., Zhou, H., Lin, H., Li, D., Peng, X., 2019. Linking saturated hydraulic conductivity and air permeability to the characteristics of biopores derived from X-ray computed tomography. *Journal of Hydrology* 571, 1-10.