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**On-site performance of floating treatment wetland macrocosms augmented with dye-degrading bacteria for the remediation of textile industry wastewater**

Nain Tara<sup>a,b</sup>, Muhammad Arslan<sup>b,c,d,\*</sup>, Zahid Hussain<sup>e</sup>, Mazhar Iqbal<sup>b</sup>, Qaisar Mahmood Khan<sup>b</sup>, Muhammad Afzal<sup>b,\*</sup>

<sup>a</sup> Pakistan Institute of Engineering and Applied Sciences (PIEAS), Islamabad, Pakistan

<sup>b</sup> Soil and Environmental Biotechnology Division, National Institute for Biotechnology and Genetic Engineering (NIBGE), Faisalabad

<sup>c</sup> Department Environmental Biotechnology, Helmholtz Centre for Environmental Research, Leipzig, Germany

<sup>d</sup> Institute for Biology V (Environmental Research), RWTH Aachen University, Templergraben 55, 52062 Aachen, Germany

<sup>e</sup> Interloop Limited Khurrianwala, Faisalabad, Pakistan

**Correspondence: M. Arslan**

Department Environmental Biotechnology, Helmholtz Centre for Environmental Research, Leipzig, Germany

E-mail(s): arsilan324@gmail.com; muhamamd.arslan@ufz.de

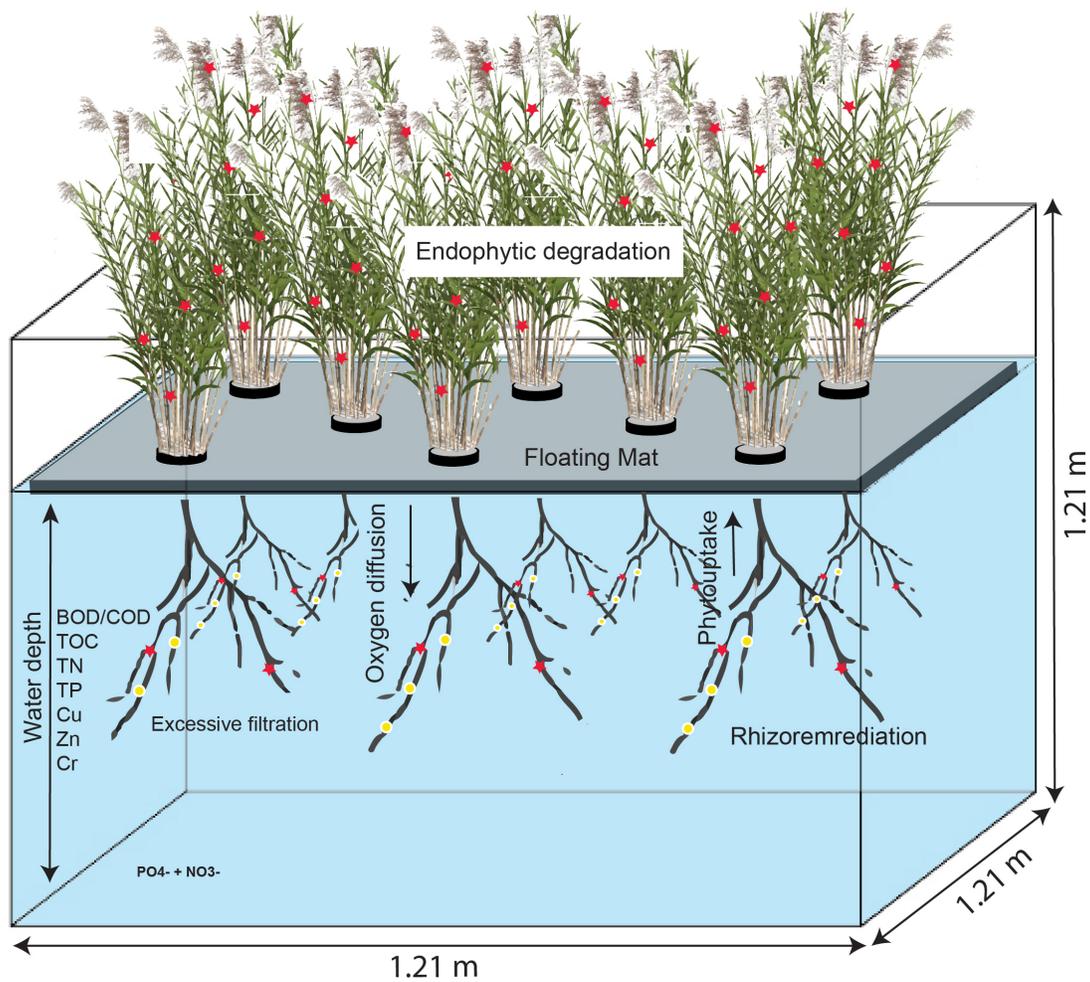
**M. Afzal**

Soil and Environmental Biotechnology Division, National Institute for Biotechnology and Genetic Engineering (NIBGE), PO Box 577, Jhang Road, Faisalabad 38000, Pakistan.

E-mail(s): manibge@yahoo.com; afzal@nibge.org.

Telephone: +92 41 920136-20

- ★ Endophytic Bacteria
- Rhizospheric bacteria



1 **ABSTRACT**

2 Floating treatment wetlands are considered cost-effective remediation options for various  
3 types of wastewater. Their effectiveness has been shown in several lab-scale and pilot-scale  
4 studies; however, there is a paucity of published data on pilot-scale systems treating genuine  
5 wastewater. This study aims to assess the performance of a pilot-scale system, carrying  
6 *Phragmites australis* in combination with three plant growth promoting and pollutant-  
7 degrading bacteria (*Acinetobacter junii* strain NT-15, *Rhodococcus sp.* strain NT-39, and  
8 *Pseudomonas indoloxydans* strain NT-38) for the treatment of textile industry wastewater  
9 (Interloop Limited, Faisalabad, Pakistan). Fifteen floating treatment wetlands macrocosms  
10 were established employing plants and bacteria separately or in combination. Each unit was  
11 capable to carry 1000-liter of wastewater and the system was operated in a batch-wise mode  
12 for the period of 2 years. After a year of installation, performance of all FTWs units was  
13 optimal. A high removal in organic and inorganic pollutants was observed in the vegetated  
14 tanks, whereas combined application of plants and bacteria further enhanced the removal  
15 performance, i.e., chemical oxygen demand was reduced to 92%, biochemical oxygen  
16 demand to 91%, color to 86%, and heavy metals to approximately 87% in the wastewater.  
17 The augmented bacteria displayed persistence in water as well as in the roots and shoots of *P.*  
18 *australis* suggesting a potential partnership with the host towards enhanced performance.  
19 Treated wastewater met the National Environmental Quality Standards of Pakistan to be  
20 discharged in the surface water without any potential risks. This pilot-scale study is a step  
21 forward towards sustainable remediation of the textile wastewater in the field.

22

23 **Keywords:** Textile wastewater, wastewater treatment, floating treatment wetlands, plant-  
24 bacteria partnership

25

## 26 INTRODUCTION

27 Textile industry plays a key role in the economy of many developing countries including  
28 Pakistan. However, it also causes severe water pollution due to the direct wastewater  
29 discharges in the surface water (Noreen et al. 2017). The textile industry of Faisalabad  
30 (Pakistan) represents a wide array of processes such as desizing, bleaching, mercerizing,  
31 dyeing, printing and finishing of products. This results into production of a broad range of  
32 organic and inorganic pollutants, which are discharged into the Ravi river and Chenab river  
33 through the Paharang and Madhuana drains without any pre-treatment (Azizullah et al. 2011;  
34 Imtiazuddin et al. 2012). These discharges consequently deteriorate the water quality leading  
35 to harmful effects on aquatic organisms, irrigating crops, and on human health (Ullah et al.  
36 2017; Najam-us-Sahar et al. 2017; Daud et al. 2017). Since a few years, this condition has  
37 become worse as the level of toxic pollutants in the river's body has surged dramatically.

38 Many physicochemical techniques are effective in treating textile wastewater but they  
39 demand high operational costs, engineering skills, environmental incursions, labor  
40 administration, and several other operational processes (Liu et al. 2009; Srinivasan et al.  
41 2014). Contrarily, advances in ecological engineering have made us develop environmental-  
42 friendly methods for the effective treatment of contaminated water (Giannetti et al., 2002;  
43 Ijaz et al., 2015; Saumya et al., 2015; Greenway, 2017). One such method is the installation  
44 of floating treatment wetlands (FTWs) in which emergent aquatic macrophytes are planted  
45 artificially on a floating raft that allows plants to grow hydroponically on the water body  
46 (Nahlik and Mitsch 2006; Wu et al., 2017; Shahid et al. 2018). Plant roots hang down to the  
47 pelagic zones in the water column offer both mechanical and biological filtering. Mechanical  
48 filtering is achieved through the physical effect of plant roots such as sedimentation,  
49 adsorption, filtration, etc. The biological filtering however is the result of bacterial

50 degradation and plant uptake (Zhang et al. 2106; Merkhali et al. 2015). In both situations,  
51 plant roots play several important roles such as: (1) their presence reduce the water  
52 turbulence and avoid re-suspension of sediments (Smith and Kalin 2000); (2) they attach  
53 suspended matter onto the root surface which is subsequently precipitated in the systems  
54 bottom or adsorbed on to the biofilm; and (3) when adsorbed, bacterial action results into the  
55 degradation of organic pollutants (Prajapati et al. 2017). Macrophytes can hinder the algal  
56 growth by competing for nutrition and sunlight (Li et al. 2010). Lastly, these systems also act  
57 as habitate for fish harvesting (Faulwetter et al. 2011), livestock grazing (Ladislav et al.  
58 2013), improve landscape aesthetics (Ijaz et al. 2015), and provide a significant amount of  
59 biomass to be used for bio-energy purposes (Shahid et al. 2018).

60 Floating wetlands have been traditionally employed to treat variety of wastewater  
61 such as stormwater (Headley et al. 2008), sewage effluents (Todd et al. 2003), municipal  
62 wastewater (Arshad et al. 2017; Sirage et al. 2017), industrial wastewater (Li et al. 2012),  
63 acid mine drainage and poultry processing wastewater (Smith and Kalin 2000; Todd et al.  
64 2003). These systems, however, are limited in certain contexts such as poor stability and less  
65 efficient purification performance. Among them, less potential of indigenous bacteria toward  
66 the degradation of toxic organic compounds and weak metabolic capabilities of plants are  
67 prominent ones (Arslan et al. 2017). To improve the treatment efficiency of FTWs, additional  
68 approaches have been recommended among which inoculation of plant growth promoting  
69 (PGP) and pollutant-degrading bacteria is an effective method (Shehzadi et al. 2014; Saleem  
70 et al. 2018). To date, a few studies have explored the efficiency of FTWs in the presence of  
71 bacterial partnership for the remediation of different wastewaters (Kabra et al. 2013;  
72 Watharkar et al. 2015; Rehman et al. 2018). Many of them were conducted at microcosm  
73 scale for which field-scale performance is speculated. In this study, *in situ* pilot-scale FTW  
74 system was established in combination with dye degrading bacteria for the enhanced cleanup

75 of wastewater of a textile industry, i.e., Interloop Limited, Khurrianwala, Faisalabad,  
76 Pakistan. There were enhanced pollutant removal and toxicity reduction in the bacterially  
77 assisted FTWs as a result of effective plant-bacterial partnership towards remediation of  
78 textile industry wastewater.

79

## 80 MATERIAL AND METHODS

### 81 Collection and characterization of textile wastewater

82 The untreated textile wastewater was obtained from an outlet of the wastewater equalization  
83 tank of Interloop Limited, Khurrianwala, Faisalabad, Pakistan. The wastewater was analyzed  
84 for several physicochemical parameters such as pH, color, electrical conductivity (EC),  
85 chemical oxygen demand (COD), 5-day biochemical oxygen demand (BOD<sub>5</sub>), total dissolved  
86 solids (TDS), total suspended solids (TSS), total nitrogen (TN), phosphate (PO<sub>4</sub><sup>-2</sup>), Phenols,  
87 Chlorides, Sulphates, heavy metals and toxicity by using standard methods (APHA, 2005).

### 88 Bacterial strains

89 Three bacterial strains, i.e., *Acinetobacter junii* NT-15 (NCBI accession: MF478980),  
90 *Pseudomonas indoloxydans* NT-38 (NCBI accession: MF478985), and *Rhodococcus sp.* NT-  
91 39 (NCBI accession: MF326802), were used in the present study. The strain *A. junii* NT-15  
92 was isolated from the activated sludge, whereas strain *P. indoloxydans* NT-38 and  
93 *Rhodococcus sp.* NT-39 were isolated from the root interior and rhizosphere of *Polygonum*  
94 *aviculare* and *Poa labillardierei*, respectively (Tara et al. 2018). These strains were screened  
95 because of their high dye resistance and decolorization potential. These bacterial strains were  
96 cultivated as separate cultures at 30 °C for 24 h in Luria-Bertani (LB) broth. The cells were  
97 harvested by centrifugation (×12,000 rpm) for 5 minutes at 4 °C and resuspended together at

98 ratio of 1:1:1 in 0.9% (w/v) sterile NaCl solution. The cell suspension of each pure culture  
99 was adjusted as per the guidelines of the turbid metric method (Sutton, 2011). In the end, one  
100 liter of bacterial consortium was used as an inoculum in the FTWs as per experimental  
101 design.

## 102 **Construction and implementation of FTWs for textile wastewater treatment**

103 Floating treatment wetlands were built in the vicinity of Interloop Limited Khurrianwala,  
104 Faisalabad (Fig. 1A-D). Plastic tanks of 1000 liters of capacity (dimensions: 1.2 meter length,  
105 1.2 meter width, and 1.2 meter height) were used to establish macrocosms. These tanks were  
106 painted from all the sides to prevent algal growth. Accordingly, floating mats of equal  
107 dimensions were prepared using polyethylene sheet as recommended earlier (Ijaz et al. 2015).  
108 Aluminum foil was used to cover all sides of the mats in order to protect them from  
109 ultraviolet radiations. Eight holes of equal diameter were drilled in each floating mat and  
110 planted with 24 healthy seedlings of *Phragmites australis*: three plants, with the height of  
111 approximately 60 cm and weight of 45 to 65 g, were inserted in each hole of the mat. *P.*  
112 *australis* was selected based on its successful application in phytoremediation of industrial  
113 wastewater as well as its ability to resist high concentrations of azo dyes (Todorovics et al.  
114 2005; Stefanakis et al. 2014; Ha and Anh 2017; Rehman et al. 2018). These seedlings were  
115 previously grown in the nursery developed in the vicinity of NIBGE, Faisalabad. Soil and  
116 coconut shaving were used to support the plant seedlings. A total of 15 macrocosms were  
117 established by placing floating mats on the tank surface. The plant seedlings were then  
118 allowed to augment roots in tap water for a period of 1-month. When the plant growth was  
119 optimum, tap water was replaced with the textile wastewater. The one-liter inoculum of  
120 previously isolated bacterial strains was introduced in the FTWs as per the experimental  
121 design. Different treatments were established to study: (1) effect of textile effluent on plant

122 growth; (2) effect of vegetation on textile effluent degradation; (3) effect of bacterial  
123 augmentation on textile effluent degradation; (4) effect of bacterial augmentation on plant  
124 growth and textile effluent degradation; and (5) plant growth in tap water (control). The  
125 sequencing fill-and-draw batch mode method was used to withdraw decontaminated water.  
126 Hydraulic retention time was set to 10 days based on lab-scale observations where a  
127 continuous decrease in organic and inorganic load was observed in initial 10-days. The  
128 system was operated for 2 years from March 2016 to March 2018. The water samples of  
129 approximately 1.5 liters were collected every two days. However, in this article, the results  
130 are reported when the system's performance was optimum/constant (after 6 months of  
131 installation). The treatment performance was established by measuring wastewater  
132 parameters such as pH, EC, TDS, TSS, COD, BOD<sub>5</sub>, TN, PO<sub>4</sub><sup>-2</sup>, color, phenols, chlorides,  
133 sulphates, and heavy metals (Fe, Ni, Cr, and Cd) immediately after sample collection;  
134 otherwise samples were stored until analysis according to standard methods (APHA, 2005).  
135 The experiment was performed in the ambient conditions of the industry for the period of 2  
136 years. In this period, precipitation on average was 29 mm (Faisalabad's climate conditions);  
137 however, during rainy period, water level was manually maintained to 1000 liters by  
138 covering the macrocosms with plastic sheets.

### 139 **Determination of persistence of inoculated bacteria**

140 The survival/persistence of inoculated bacteria in the treated wastewater and plant interior  
141 (root and shoot interior) was enumerated by viable plate count method. Samples were  
142 collected every 3 months. Shoots and roots were surface sterilized, homogenized, and the  
143 slurry was plated onto LB agar plates having 100 mg l<sup>-1</sup> of azo dyes as established previously  
144 (Ijaz et al. 2015). Similarly, treated wastewater was also plated on LB agar plates and

145 incubated at 30 °C for 24 h. Restriction fragment length polymorphism (RFLP) analysis was  
146 performed to identify the identity of inoculated strains (Afzal et al. 2012).

147

#### 148 **Plant biomass**

149 The plant tissues were harvested every 3 months to measure shoot height, root length, shoot  
150 dry weight, and root dry weight. Briefly, shoots and roots were harvested one inch above and  
151 below the mats, respectively. The root length and shoot height were measured by using hand  
152 ruler. To measure dry biomass, plant samples were first transported to the laboratory, placed  
153 in an oven at 80 °C for 72 h, and then weighed using a digital balance. Samples were stored  
154 at -80 °C until further analysis.

#### 155 **Fish toxicity bioassay**

156 Fish bioassay was performed once, during the optimal period, to test the toxicity of untreated  
157 and treated wastewater. A freshwater dwelling fish species Rohu, *Labeorohita rohita*  
158 (Family: Cyprinidae, Order: Cypriniformes) was selected for the bioassay due to the local  
159 ecological significance. Briefly, the species is present in the local streams and lakes and  
160 therefore direct discharges may lead to health issues (Khan et al. 2017). This species has  
161 previously found to accumulate heavy metals above the natural/background levels (Hamid et  
162 al. 2016). In this study, healthy specimens were obtained from Faisalabad Fish Hatchery,  
163 treated with KMnO<sub>4</sub> solution (0.05 %) for two minutes to remove dermal contamination, and  
164 then shifted to glass aquaria containing uncontaminated water. Each fish specimen had an  
165 average body length of  $\sim 8.5 \pm 0.8$  cm and weight of  $\sim 4.0 \pm 0.1$  g. The specimens were  
166 acclimatized for 15 days prior to the effluent exposure. Subsequently, four fish groups each  
167 containing 10 individuals were placed in the aquariums (dimensions: depth 30 cm, width 30

168 cm, and length 45 cm). These aquariums were filled with 30 L of treated and untreated  
169 wastewater, as per the experimental design. Finally, the survival rate was determined by  
170 counting the number of alive fish every 24 h for a period of four days.

### 171 **Long-term maintenance strategy**

172 Maintenance strategy of the system involved harvesting of the plants and inoculation of the  
173 bacterial consortium in the wetland macrocosms after every 3 months during the  
174 experimental period. Accordingly, long-term system's performance was also monitored by  
175 measuring temporal response of basic water quality parameters, i.e., COD, BOD, Color, TSS,  
176 TDS, TN, Phosphates.

### 177 **Statistical analysis**

178 R-statistical language was used to compare different treatments for water quality parameters  
179 (pH, EC, COD, BOD<sub>5</sub>, TDS, TSS, color, Phenols, Chlorides, and Sulphates), nutrients (TN,  
180 and PO<sub>4</sub><sup>-2</sup>), heavy metals (Fe, Cr, Ni, and Cd), plant growth parameters (root length, shoot  
181 length, fresh biomass, and dry biomass), and fish toxicity assay. The comparison was made  
182 through one-way ANOVA and Duncan's test was used after testing homogeneity of variance.

183

## 184 **RESULTS AND DISCUSSION**

### 185 **Wastewater characteristics**

186 In developing countries, unprecedented advances in textile industry have escalated the  
187 discharge of wastewater into the natural streams (Vikrant et al. 2018). In this study,  
188 physicochemical analysis of the textile industry wastewater displayed a high pollution due to  
189 COD (513 mg l<sup>-1</sup>), BOD (283 mg l<sup>-1</sup>), TDS (5251 mg l<sup>-1</sup>), TSS (324 mg l<sup>-1</sup>), Phenols (0.85 mg

190  $\text{l}^{-1}$ ), Chlorides ( $1383 \text{ mg l}^{-1}$ ), Sulphates ( $311 \text{ mg l}^{-1}$ ), Fe ( $14.4 \text{ mg l}^{-1}$ ), Ni ( $7.57 \text{ mg l}^{-1}$ ), Cr  
191 ( $9.67 \text{ mg l}^{-1}$ ), and Cd ( $0.88 \text{ mg l}^{-1}$ ). Their concentrations were higher than the established  
192 National Environmental Quality Standards (NEQS) of Pakistan (Table 1). This suggests an  
193 initial treatment of the wastewater of textile industry of Pakistan before discharging into the  
194 Ravi and Chenab rivers.

### 195 **Performance evaluation for physicochemical parameters**

196 The performance of FTWs in the presence of vegetation, bacteria, and vegetation and bacteria  
197 together, was evaluated by studying wastewater parameters at different time intervals (Table  
198 2 - 4, Fig. 2). In the beginning of the second year, the system performance was relatively  
199 optimal. Based on these observations, in-depth investigations were performed. Briefly, it was  
200 found that FTWs vegetated with *P. australis* removed a high proportion of organic and  
201 inorganic pollutants as compared to the un-vegetated FTWs. This removal was further  
202 enhanced when *P. australis* and bacterial consortium were applied in combination. Briefly, in  
203 the presence of both partners, COD was reduced to 91%, BOD to 92%, and color to 86%  
204 (Fig. 2A-C). Earlier studies have also shown highest pollution reduction of textile wastewater  
205 when plants and bacteria employed synergistically (Shehzadi et al. 2014; Watharkar et al.  
206 2015; Hussain et al. 2018a); this could be attributed to the combined enzymatic activities of  
207 both bacteria and plants to transform organic matter into simple metabolites (Kabra et al.  
208 2013; Khandare et al. 2013). These transformed products can be taken up by plants as a part  
209 of nutrient assimilation process or eliminated in the form of gases, e.g.,  $\text{CO}_2$  and  $\text{N}_2$ . With the  
210 passage of time, this reduction in organic load was increased and maximum remediation was  
211 observed in the period of 10 days. Accordingly, a positive correlation for reduction of COD  
212 and color was seen in textile wastewater. It has been reported previously that COD  
213 determines the extent of oxidizable contaminants and hence establish positive correlation

214 with the color of wastewater (Tara et al. 2018). The observation was consistent in this study  
215 as well. Likewise, pH of the remediated water moved from basic to neutral conditions (Table  
216 2). This could also be associated with the combined action of dyes transformation by bacteria  
217 and plants (Watharkar et al. 2015). Accordingly, a significant amount of phenols, chlorides,  
218 and sulphates were removed from the wastewater (Figure 2D). Earlier studies have reported  
219 similar results in different types of CWs (Shehzadi et al. 2014; Hussain et al. 2018a,b).

220 Vegetation had a prominent effect on TDS and TSS removal as compared to the  
221 unvegetated but inoculated treatments (Table 2). Briefly, vegetation reduced TDS from 5251  
222 mg l<sup>-1</sup> to 2265 mg l<sup>-1</sup> (up to 57%) and TSS from 324 mg l<sup>-1</sup> to 165 mg l<sup>-1</sup> (up to 49%). The  
223 bacterial inoculation in vegetated treatment further increased reduction in TDS and TSS up to  
224 1399 mg l<sup>-1</sup> (73%) and 148 mg l<sup>-1</sup> (54%), respectively. These results are in accordance to the  
225 previously published findings (Shehzadi et al. 2014; Tara et al. 2018; Hussain et al. 2018a,b).  
226 As of these findings, vegetated treatments also displayed better removal of nutrients (TN and  
227 PO<sub>4</sub><sup>-2</sup>) than the un-vegetated treatments whereas highest removal efficiency was seen in the  
228 presence of plant-bacteria synergism (Table 3). The removal efficiency for TN was recorded  
229 up to 60% in the vegetated FTWs, which was increased to 87% on bacterial inoculation. In an  
230 earlier study, Sun et al. (2009) reported similar results where TN removal was enhanced up to  
231 72% by the addition of bacteria in FTWs whereas only 50% removal was achieved in the  
232 treatments without bacteria. Similar results were obtained for PO<sub>4</sub><sup>-2</sup> reduction as bacterial  
233 augmentation improved the removal as compared to the vegetation only. Nevertheless,  
234 percent removal for TN was higher than the PO<sub>4</sub><sup>-2</sup>, which can be linked to the fact that  
235 nitrogen, in addition to the plant uptake, is eliminated in the form of N<sub>2</sub> gas whereas PO<sub>4</sub><sup>-2</sup>  
236 stays in the system due to precipitation (Tao and Wang 2009; Vymazal, 2010).

237 Long-term system's performance was monitored to evaluate pollutant removal during  
238 the studied period, as shown in Figure 3. It was found that the system was relatively stable  
239 throughout the experimental period; nevertheless, performance was better in the warm  
240 months.

241

#### 242 **Reduction in heavy metals from wastewater and their bioaccumulation in plant tissues**

243 The studied heavy metals (Fe, Ni, Cr, and Cd) were significantly removed by FTWs (Table  
244 4). The removal efficiency was equally valid for both essential (Fe and Ni) and non-essential  
245 (Cr and Cd) heavy metals. Nevertheless, bacterial inoculation improved the bioaccumulation  
246 and phytoremediation potential of *P. australis*. The mean concentration values of heavy  
247 metals in root, shoots, and leaves were observed in the following order: Fe > Cr > Ni > Cd.  
248 Absorbed metals were partitioned among different parts of *P. australis*. Roots retained higher  
249 concentration of heavy metals than shoots and leaves with the following order:  
250 root>leaf>stem. These results are in accordance to the earlier findings demonstrating that  
251 underground part of *P. australis* shows a higher storage capacity than the aboveground  
252 organs (Štrbac et al. 2014; Kucaj and Abazi 2015; Amira and Leghouchi 2017). It has been  
253 established that roots and rhizomes of *P. australis* possess large intercellular air spaces of  
254 cortex parenchyma that allow accumulation of heavy metals far away from the metabolically  
255 active structures to avoid toxic effects (Mendonça et al. 2015). The present study confirms  
256 such a potential of *P. australis* in phytoremediation for the studied heavy metals which was  
257 further enhanced by bacterial inoculation, i.e., the system was able to remove Cr and Fe more  
258 than 90%, Ni more than 80%, and Cd more than 60%. The role of bacteria in sorbing metallic  
259 ions on their cell walls is prominent and has also been reported in terms of increasing  
260 bioavailability followed by uptake by plants (Jilani and Khan 2013; Khan et al. 2014).

**261 Inoculated bacteria displayed persistence in FTWs**

262 In phytoremediation, plant-associated microbial populations play a key role to mineralize  
263 organic pollutants (Khan et al. 2014). Many studies have reported that the phytoremediation  
264 potential of plants can be correlated with the proportion of bacteria in the surrounding  
265 environment (Yousaf et al. 2011; Shehzadi et al. 2014; Rehman et al. 2019). The artificial  
266 augmentation of bacteria in such a system establishes intimacy with their host depending  
267 upon the environmental conditions such as nutrient supply and their ability to colonize the  
268 host environment. Moreover, bacteria improve plant growth by reducing biotic and abiotic  
269 stress, followed by N<sub>2</sub> fixation, production of phytohormones, and solubilization of various  
270 essential nutrients (Glick et al. 2010; Khan et al. 2014). In this study, in order to confirm the  
271 improvement in performance of *P. australis* due to bacterial augmentation, persistence of  
272 inoculated bacteria was evaluated within the plant tissues and wastewater at the beginning of  
273 the experiment. Results showed a high persistence of inoculants in the plant interior,  
274 especially in the roots, followed by the wastewater (Fig. 4). Furthermore, RFLP analysis  
275 indicated that the relative proportion of inoculated bacterial with roots and shoots was  
276 increased significantly in the initial 3-months period, which later became stable. Although the  
277 trend was similar for both plant parts, their relative proportion was higher in the roots as  
278 compared to the shoots. This might be due to the fact that The inoculated bacteria were  
279 previously isolated from the roots and shoots of *P. australis* grown in the textile wastewater  
280 and therefore they probably have adapted necessary mechanism for proliferation in these  
281 hostile conditions (Afzal et al. 2014). By contrasts, a decline in bacterial counts was observed  
282 in the water, which might be due to the fact that contaminated water is less favorable  
283 environment for the inoculated bacteria. More specifically, rhizo- and endophytic bacteria  
284 needs a symbiotic partner such as plant roots to survive and proliferate, however, the absence  
285 can cause poor survival and growth (Arslan et al. 2014). Additionally, this might be attributed

286 to the natural selection pressure for the newly grown community that allows bacteria to  
287 migrate from less favorable environment to the more favorable environment.

288

289

### 290 **Plant biomass and growth**

291 An important parameter in phytoremediation efficiency is the plant tolerance and survival in  
292 the presence of pollutants. The growth of *P. australis* was increased sharply in the first year,  
293 which was then sustained or remained unaffected during the remaining studied period (Figure  
294 5). This could be attributed to the senescence effect. Nevertheless, when compared with the  
295 contaminated water, it was found that wastewater inhibited the plant biomass significantly  
296 (Table 5). This shows the toxic effects of textile wastewater on plant metabolism (Watharkar  
297 et al. 2015; Shehzadi et al. 2016; Ramya et al. 2017). In an earlier study, *Typha domingensis*  
298 exhibited similar behavior in which reduction in plant growth was seen in the presence of  
299 textile wastewater (Shehzadi et al. 2014). Nevertheless, inoculation with bacteria  
300 rehabilitated the plant growth with a prominent increase in root length (46%), shoot length  
301 (37%), root fresh weight (58%), shoot fresh weight (49%), root dry weight (55%), and shoot  
302 dry weight (46%) (Table 5). This increase in plant growth parameters could be associated  
303 with the pollutant degrading capabilities as well as ACC deaminase potential of the  
304 inoculated bacteria. Fatima et al. (2016) reported similar results in which plant growth was  
305 significantly improved by the augmented of endophytic bacteria due to their PGP activities  
306 such as indole acetic acid production, siderophores formation, and ACC deaminase activity.

### 307 **Detoxification of wastewater**

308 Fish toxicity bioassay suggested a successful reduction in wastewater's toxicity after  
309 treatment with FTWs. This reduction was partially effective when plant and bacteria were  
310 applied separately, i.e., death of three and five fish out of 10 specimens. Whereas, bacterial  
311 inoculation in vegetated FTWs resulted in complete detoxification as no fish died even after  
312 72 h of exposure (Table 6). These results strengthen the earlier observations in which high  
313 reduction was observed for COD, BOD<sub>5</sub>, TDS, TSS, color, and heavy metals in the presence  
314 of vegetation and bacteria. Earlier studies also reported similar findings in which use of  
315 plants in combination with bacteria was an efficient strategy to remove wastewater toxicity  
316 (Tanner and Headley 2011; Ijaz et al. 2016; Rehman et al. 2018).

317

## 318 **CONCLUSIONS**

319 This study investigated the performance of FTWs in the presence of three bacterial strains to  
320 remediate textile wastewater at pilot-scale. FTWs vegetated with *P. australis* and inoculated  
321 bacteria displayed highest pollutants removal. Bacterial persistence was prominent in the root  
322 and shoot interior suggesting optimal partnership in pollutant degradation. The system  
323 attenuated both organic and inorganic contaminants and wastewater was found to be  
324 completely detoxified. The system was operated for two years suggesting long-term potential  
325 for the on-site remediation practices. It is concluded that bacterially assisted FTWs can be  
326 exploited to treat industrial wastewaters in countries with more economic constraints like  
327 Pakistan, where capital and operational cost are of prime importance.

328

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**Table 1**

Quality parameters of textile industrial wastewater collected from Interloop, Khurrianwala located in Faisalabad, compared to the National Environmental Quality Standards (NEQS), Pakistan (Hussain et al., 2018b)

Parameter	Unit	Wastewater	NEQS
Temperature	°C	38 (2.63)	40
pH	--	8.8 (0.82)	6-10
Electrical conductivity (EC)	mS cm <sup>-1</sup>	8.2 (0.64)	NG
Color	m <sup>-1</sup>	66 (4.4)	NG
Chemical oxygen demand (COD)	mg l <sup>-1</sup>	<b>513 (37.6)</b>	<b>150</b>
Total organic carbon (TOC)	mg l <sup>-1</sup>	201 (15.9)	NG
Biochemical oxygen demand (BOD <sub>5</sub> )	mg l <sup>-1</sup>	<b>283 (17.9)</b>	<b>80</b>
Phenol	mg l <sup>-1</sup>	<b>0.85 (0.06)</b>	<b>0.1</b>
Chloride	mg l <sup>-1</sup>	<b>1383 (61.8)</b>	<b>1000</b>
Sulphate	mg l <sup>-1</sup>	311 (21.9)	600
Nitrogen	mg l <sup>-1</sup>	28.7 (6.0)	40
Phosphate	mg l <sup>-1</sup>	16.4 (3.6)	NG
Total dissolved solids (TDS)	mg l <sup>-1</sup>	<b>5251 (404)</b>	<b>3500</b>
Total solids (TS)	mg l <sup>-1</sup>	5420 (395)	NG
Total settleable solids (TSeS)	mg l <sup>-1</sup>	19.9 (1.1)	NG
Total suspended solids (TSS)	mg l <sup>-1</sup>	<b>324 (29.7)</b>	<b>150</b>
Iron (Fe)	mg l <sup>-1</sup>	<b>14.4 (0.64)</b>	<b>2.0</b>
Nickel (Ni)	mg l <sup>-1</sup>	<b>7.57 (0.38)</b>	<b>1.0</b>
Chromium (Cr)	mg l <sup>-1</sup>	<b>9.67 (0.26)</b>	<b>0.1</b>
Cadmium (Cd)	mg l <sup>-1</sup>	<b>0.88 (0.02)</b>	<b>0.1</b>
Toxicity	--	Highly toxic	NG

Each value is the mean of three replicates; standard error among three replicates is presented in parenthesis. NG, not given in National Environmental Quality Standards (NEQS) list.

**Table 2**Remediation of the textile industrial wastewater by floating treatment wetlands vegetated with *Phragmites australis* and inoculated with bacterial consortium

Days	Control				T1				T2				T3			
	pH	EC	TDS	TSS												
0	8.73 (0.20)	8.22 (0.64)	5251 (404)	324 (30)	8.74 (0.16)	8.34 (0.66)	5252 (404)	325 (31)	8.75 (0.19)	8.27 (0.61)	5251 (404)	325 (29)	8.74 (0.20)	8.25 (0.61)	5252 (404)	324 (29)
2	8.49 (0.18)	7.72 (0.64)	4934 (411)	319 (29)	8.43 (0.17)	7.27 (0.55)	4635 (354)	275 (19)	8.19 (0.12)	6.64 (0.55)	4250 (355)	256 (18)	8.55 (0.08)	7.34 (0.67)	4699 (430)	310 (28)
4	8.38 (0.17)	7.36 (0.69)	4677 (442)	306 (28)	8.14 (0.13)	6.42 (0.43)	4103 (279)	241 (18)	7.83 (0.13)	5.73 (0.43)	3632 (277)	225 (12)	8.35 (0.13)	6.84 (0.61)	4356 (389)	296 (29)
6	8.58 (0.09)	6.7 (0.72)	4295 (462)	300 (29)	7.67 (0.10)	5.35 (0.53)	3397 (339)	224 (16)	7.46 (0.10)	4.35 (0.49)	2735 (314)	206 (11)	8.58 (0.09)	6.37 (0.69)	4016 (441)	290 (27)
8	8.88 (0.08)	6.2 (0.70)	3974 (447)	294 (29)	7.41 (0.10)	4.42 (0.43)	2843 (278)	199 (16)	7.14 (0.10)	3.49 (0.46)	2183 (295)	171 (12)	8.81 (0.11)	5.67 (0.62)	3632 (394)	286 (26)
10	9.04 (0.04)	5.7 (0.61)	3655 (389)	285 (28)	7.35 (0.06)	3.57 (0.35)	2265 (222)	165 (12)	7.12 (0.09)	2.26 (0.30)	1399 (185)	148 (10)	8.87 (0.09)	5.13 (0.46)	3292 (296)	271 (24)

Floating treatment wetlands vegetated with *P. australis* (T1), *P. australis* and bacteria (T2), and bacteria only (T3). Each value is the mean of three replicates; the standard error of three replicates is presented in parenthesis. TDS and TSS are mentioned in mg l<sup>-1</sup>, whereas EC in mS cm<sup>-1</sup>.

**Table 3**

Nutrients (TN and  $\text{PO}_4^{2-}$ ) removal from textile industrial wastewater by floating treatment wetlands vegetated with *Phragmites australis* and inoculated with bacterial consortium

Days	TN ( $\text{mg l}^{-1}$ )				$\text{PO}_4^{2-}$ ( $\text{mg l}^{-1}$ )			
	Control	T1	T2	T3	Control	T1	T2	T3
0	28.7 (6.0)	28.7 (6.0)	28.9 (5.6)	28.9 (5.7)	16.4 (3.6)	16.4 (3.6)	16.5 (3.6)	16.5 (3.6)
2	27.0 (5.4)	22.6 (4.5)	19.3 (4.1)	25.9 (5.1)	16.1 (3.6)	15.7 (3.4)	15.1 (3.4)	15.9 (3.3)
4	25.9 (5.1)	20.3 (3.9)	15.7 (2.9)	23.6 (4.7)	15.9 (3.4)	14.5 (3.1)	13.2 (2.8)	15.6 (3.5)
6	24.5 (4.9)	17.4 (3.2)	12.0 (1.9)	22.2 (4.4)	15.8 (3.4)	13.1 (2.8)	11.8 (2.7)	15.2 (3.2)
8	23.2 (4.6)	14.8 (2.7)	8.7 (1.8)	20.1 (3.7)	15.6 (3.4)	11.3 (2.3)	10.0 (2.3)	14.8 (3.0)
10	22.3 (4.5)	11.3 (2.0)	3.7 (0.6)	19.0 (3.5)	15.3 (3.3)	10.1 (2.1)	8.2 (1.9)	14.2 (2.9)

Floating treatment wetlands vegetated with *P. australis* (T1), *P. australis* and bacteria (T2), and bacteria only (T3). Each value is the mean of three replicates; the standard error of three replicates is presented in parenthesis.

**Table 4**Heavy metals removal from textile industrial wastewater and their accumulation in different parts of *Phragmites australis*

Metal	Control		T1			T2			T3	
	Initial conc. (mg l <sup>-1</sup> )	Water (mg l <sup>-1</sup> )	Root (mg kg <sup>-1</sup> )	Shoot (mg kg <sup>-1</sup> )	Leaves (mg kg <sup>-1</sup> )	Water (mg l <sup>-1</sup> )	Root (mg kg <sup>-1</sup> )	Shoot (mg kg <sup>-1</sup> )	Leaves (mg kg <sup>-1</sup> )	Water (mg l <sup>-1</sup> )
<b>Fe</b>	14.4 (0.64)	2.63 (0.27)	5.90 (0.45)	1.97 (0.15)	3.94 (0.30)	1.05 (0.11)	6.69 (0.37)	2.23 (0.12)	4.46 (0.25)	9.0 (0.47)
<b>Ni</b>	7.57 (0.38)	1.72 (0.12)	2.92 (0.19)	0.97 (0.06)	1.95 (0.13)	1.08 (0.09)	3.25 (0.18)	1.08 (0.06)	2.16 (0.12)	5.43 (0.35)
<b>Cr</b>	9.67 (0.26)	1.50 (0.14)	4.08 (0.09)	1.36 (0.03)	2.72 (0.06)	0.40 (0.10)	4.63 (0.16)	1.54 (0.05)	3.09 (0.10)	6.53 (0.38)
<b>Cd</b>	0.88 (0.06)	0.29 (0.06)	0.24 (0.03)	0.08 (0.01)	0.16 (0.02)	0.26 (0.03)	0.31 (0.01)	0.10 (0.01)	0.21 (0.01)	0.72 (0.04)

Floating treatment wetlands vegetated with *P. australis* (T1), *P. australis* and bacteria (T2), and bacteria only (T3). Each value is the mean of three replicates; the standard error of three replicates is presented in parenthesis.

**Table 5**

Effect of bacterial inoculation on biomass and root and shoot length of *Phragmites australis* vegetated in floating treatment wetlands

Treatment	Length (cm)		Fresh biomass (g)		Dry biomass (g)	
	Root	Shoot	Root	Shoot	Root	Shoot
Tap water	67.8 (2.6)	407 (19.1)	1189 (52.1)	2194 (62.0)	152 (6.1)	510 (13.9)
Wastewater	43.6 (2.5)	294 (14.8)	936 (64.2)	1803 (48.9)	84 (7.9)	370 (13.9)
Wastewater and bacteria	54.8 (4.9)	336 (6.9)	1083 (48.9)	1994 (94.4)	122 (7.5)	434 (26.5)

Each value is the mean of three replicates; the standard error of three replicates is presented in parenthesis.

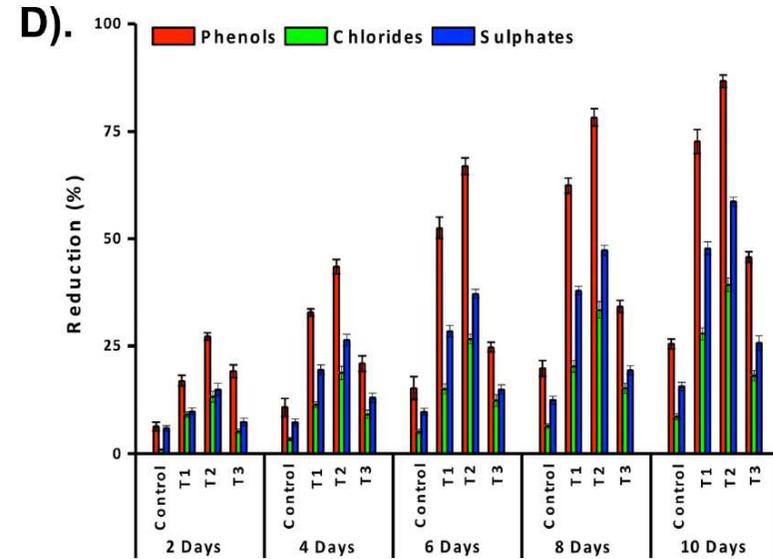
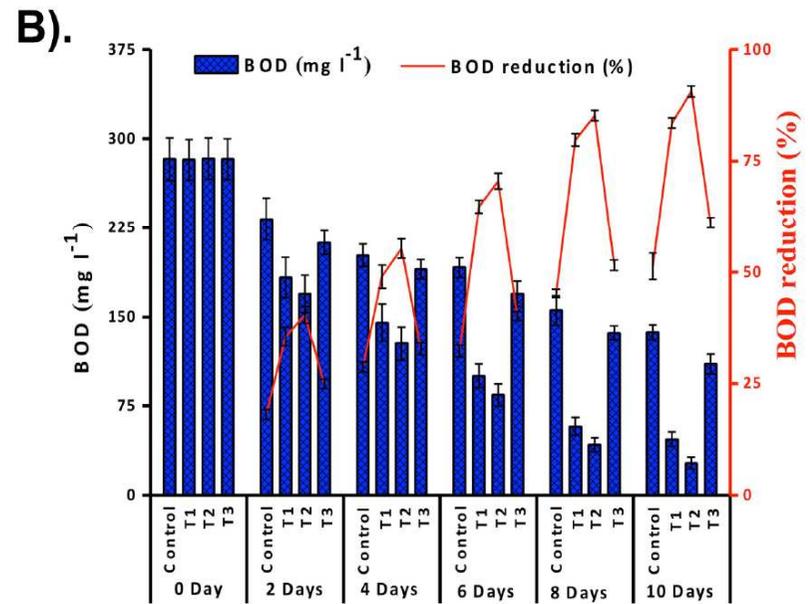
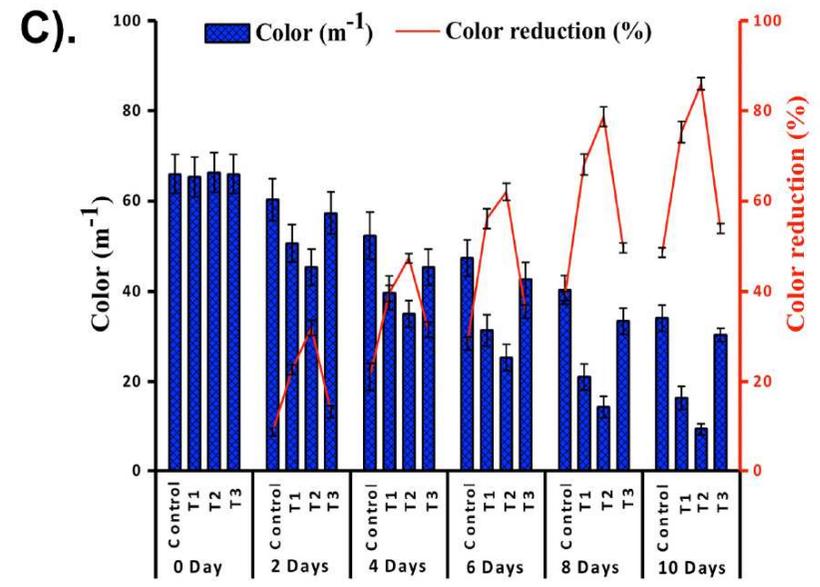
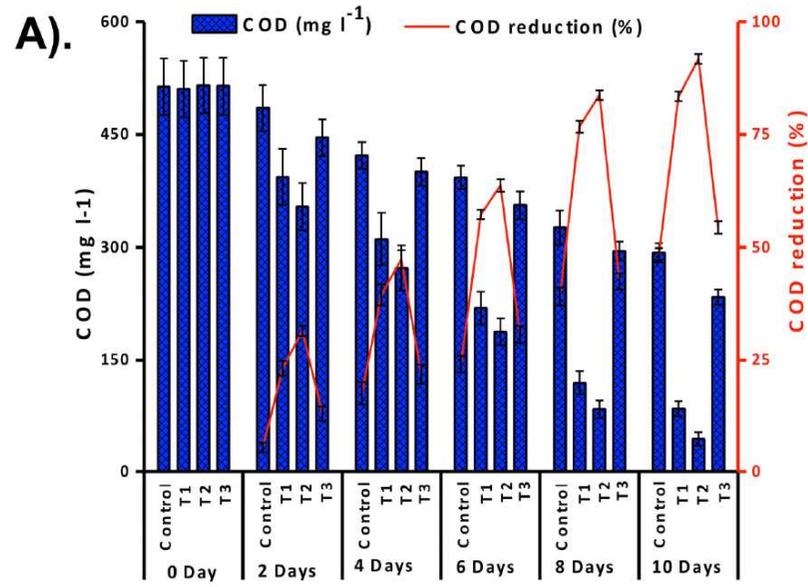
**Table 6**

Fish toxicity assay of textile industrial wastewater detoxified by floating treatment wetlands vegetated with *Phragmites australis* and inoculated with bacterial consortium (BC)

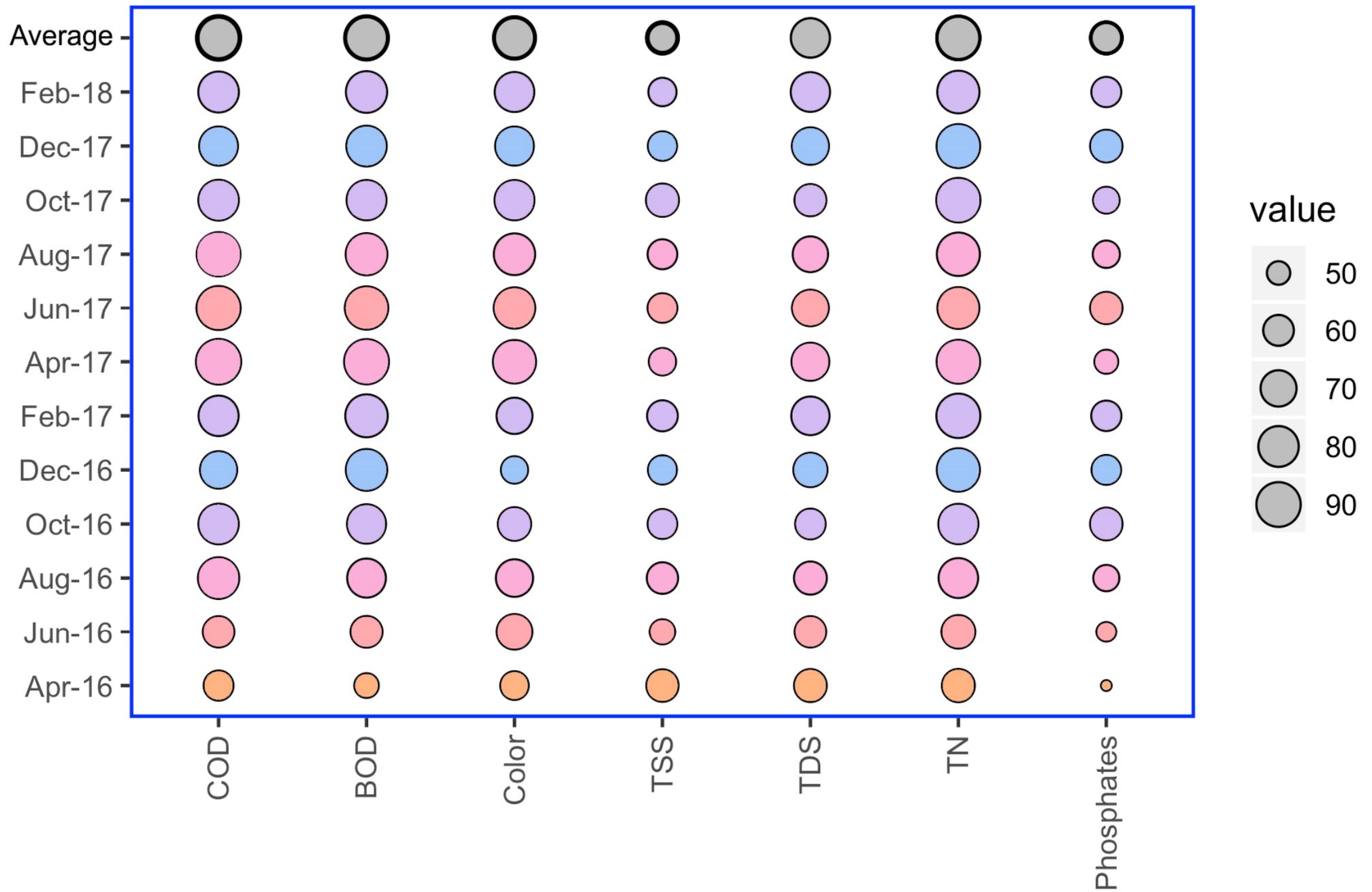
Treatment	Death over time										Total deaths	Detoxification status
	0 h	1 h	2 h	4 h	8 h	12 h	24 h	48 h	72 h	96 h		
<i>P. australis</i> (T1)	0	0	0	0	0	0	0	0	1	2	3	Partial
<i>P. australis</i> + BC (T2)	0	0	0	0	0	0	0	0	0	0	0	Complete
BC (T3)	0	0	0	0	0	0	2	1	1	1	5	Partial
Control	0	2	0	1	1	1	4	0	0	0	9	Negligible



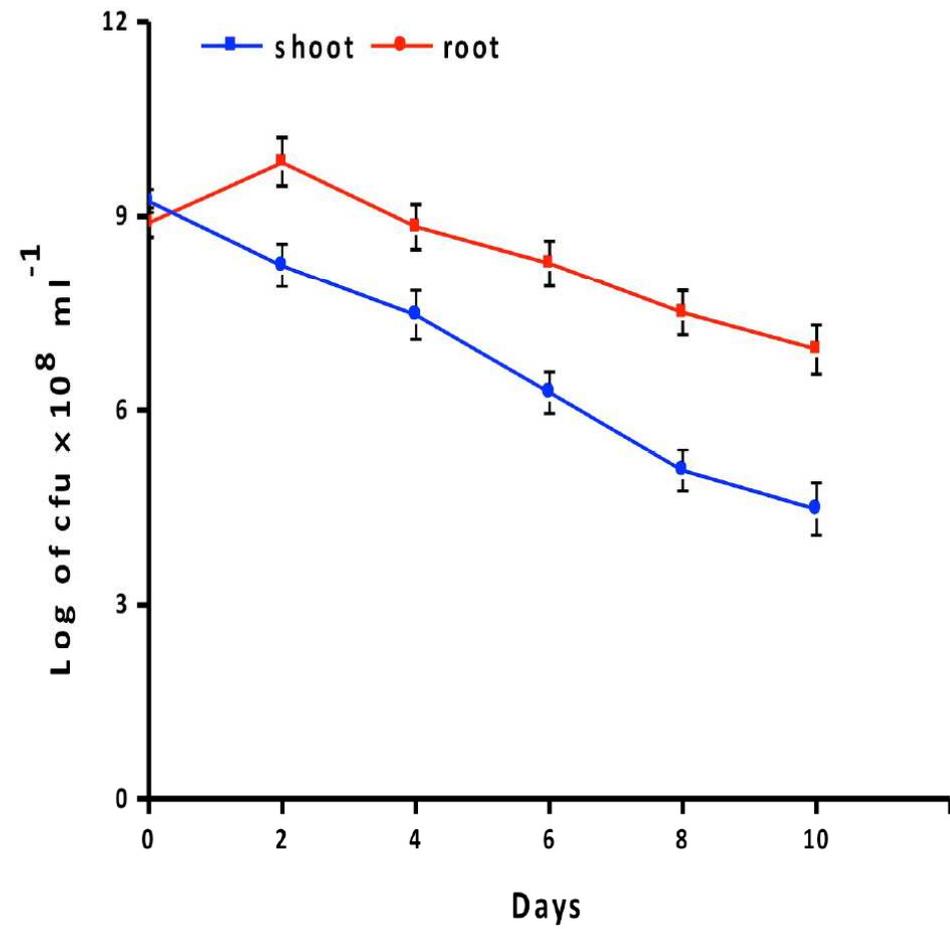
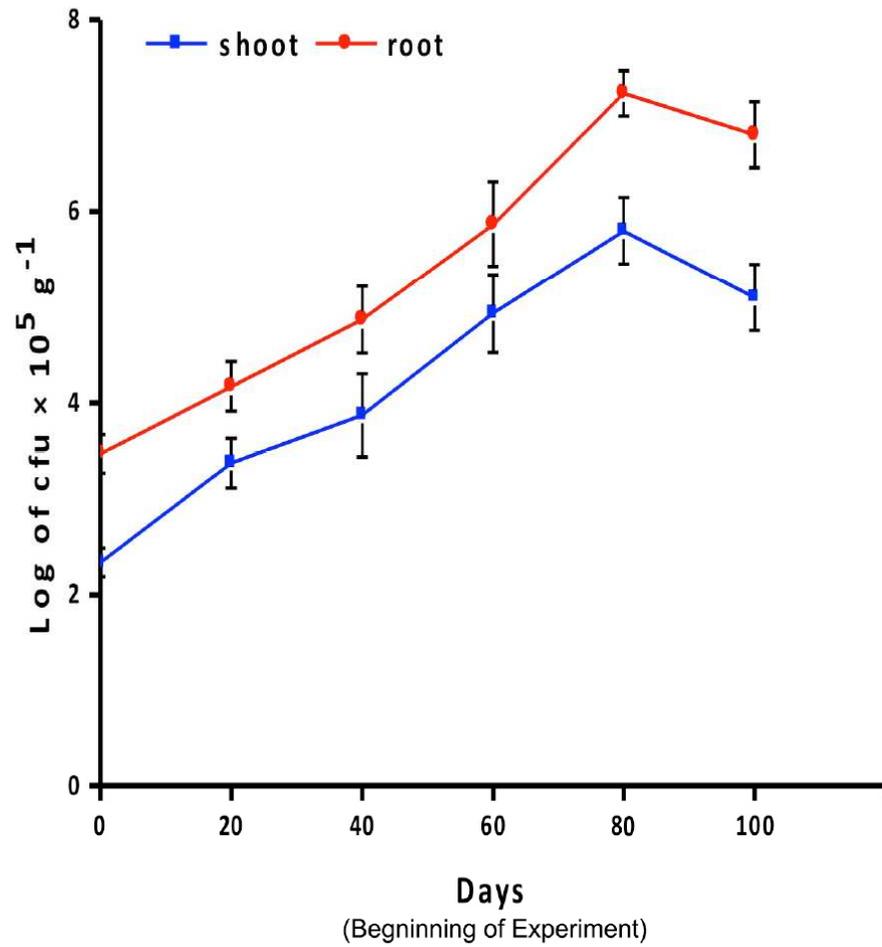
Fig. 1. Development and implementation of floating treatment wetlands (FTWs) macrocosms for the remediation of wastewater at Interloop Limited Khurrianwala, Faisalabad. Wastewater equalization tank (A), floating mat for the plantation of *Phragmites australis* seedlings (B), nursery of *P. australis* in the vicinity of NIBGE, Faisalabad (C), and growth of *P. australis* in FTWs macrocosms at the end of experimentation (D).



**Fig. 2.** Reduction of COD (A), BOD (B), Color (C), and for Phenols, Chlorides, and Sulphates (D) in the textile wastewater by floating treatment wetlands vegetated with *P. australis* (T1), *P. australis* and bacteria (T2), and bacteria only (T3). Each value is the mean of three biological replicates. Error bars indicate standard error among three replicates.

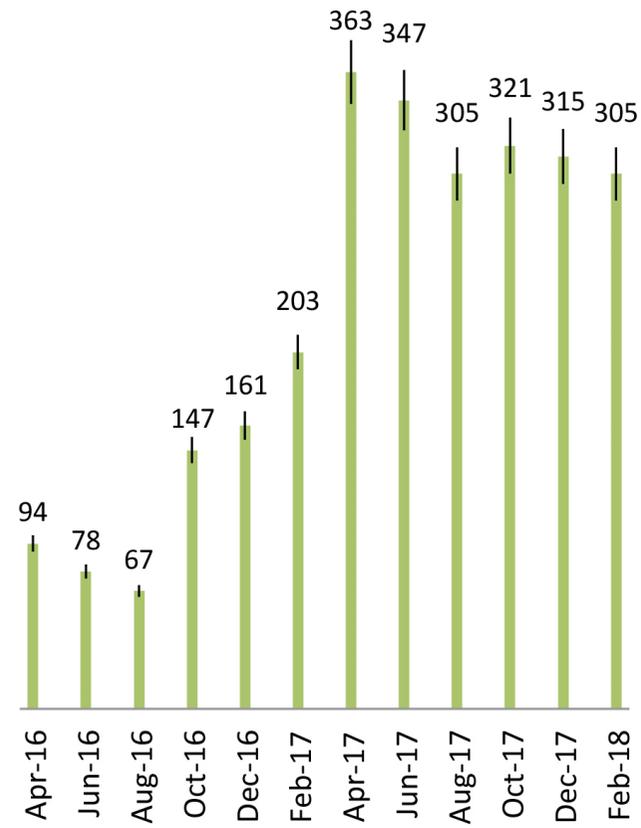


**Fig. 3:** Performance evaluation of FTW in terms of individual parameters, and overall response.



**Fig. 4.** Bacterial survival and colonization in water and in root and shoot interior of *Phragmites australis* for the macrocosms containing wastewater with vegetation and bacterial inoculation (T2).

**Biomass (g) / plant**



**Fig. 5:** Biomass of *Phragmites australis* per plant obtained after every 3-months of harvesting.

**Highlights**

Bacterially assisted FTWs macrocosms were engineered to treat real textile effluent

Combined application of plants and bacteria had a prominent effect on pollution reduction

Inoculated bacteria displayed persistence in different components of the FTWs

Treated wastewater met the National Environmental Quality Standards (NEQS) of Pakistan