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1	Simulation of Dual Carbon–Bromine Stable Isotope Fractionation during 1,2-
2	Dibromoethane Degradation
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24 Abstract

We performed a model-based investigation to simultaneously predict the evolution of concentration, 25 as well as stable carbon and bromine isotope fractionation during 1,2-dibromoethane (EDB, 26 ethylene dibromide) transformation in a closed system. The modelling approach considers bond-27 cleavage mechanisms during different reactions and allows evaluating dual carbon-bromine isotopic 28 signals for chemical and biotic reactions, including aerobic and anaerobic biological transformation, 29 30 dibromoelimination by Zn(0) and alkaline hydrolysis. The proposed model allowed us to accurately simulate the evolution of concentrations and isotope data observed in a previous laboratory study 31 and to successfully identify different reaction pathways. Furthermore, we illustrated the model 32 capabilities in degradation scenarios involving complex reaction systems. Specifically, we 33 examined (i) the case of sequential multistep transformation of EDB and the isotopic evolution of 34 the parent compound, the intermediate and the reaction product, and (ii) the case of parallel 35 competing abiotic pathways of EDB transformation in alkaline solution. 36

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38 Keywords: degradation; organic contaminants; isotope modelling; compound-specific isotope
39 analysis; stable carbon and bromine isotopes

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43 1. Introduction

In the last few decades, 1,2-dibromoethane (EDB, ethylene dibromide) has been frequently detected 44 in drinking water and natural aquatic systems, due to its extensive application as an agricultural 45 46 fumigant as well as a lead scavenger in gasoline [1,2]. EDB is a widespread pollutant and several studies have investigated its degradation under different environmental conditions [3–5]. However, 47 the environmental fate of EDB is difficult to understand and to quantitatively assess since this 48 49 chemical can undergo different transformation processes and its concentration distribution in aquatic systems also depends on physical processes such as mass-transfer, dilution and sorption [6-50 8]. Therefore, the application of compound specific isotope analysis (CSIA) is beneficial to 51 52 investigate the environmental fate of EDB. CSIA techniques have been developed and applied to a wide variety of organic pollutants [9–12], for which the determination of the change of stable 53 isotope signals could be used to identify and quantify specific transformation processes. Carbon is 54 the most common element for CSIA applications in contaminant hydrology; however, recent 55 developments on analytical techniques for chlorine and bromine CSIA allowed increasing 56 applications of dual-element isotope analysis for organohalides [13-18]. Thus, different reaction 57 pathways of halogenated organic pollutants could be characterized and understood using dual-58 element CSIA [19-23]. In a very recent laboratory study, Kuntze et al. [24] applied dual carbon-59 60 bromine CSIA to investigate different reaction mechanisms during EDB degradation.

In this work we propose an isotope modelling approach for dual carbon-bromine isotope fractionation based on the reaction mechanisms and the experimental data from the study of Kuntze et al. [24]. Isotope models are valuable tools to provide quantitative interpretation of isotopic data obtained during different transformation processes as well as in complex environmental systems where both physical and transformation processes influence the observed isotopic signals [25–27]. So far, isotope models have been developed and applied for multi-element isotopic prediction of

various organic contaminants, including chlorinated hydrocarbons [28-31], BTEX compounds 67 [32,33] and organic pesticides [34,35]. However, such a modelling framework is still lacking for 68 brominated organic compounds. This modelling case study illustrates an integrated carbon-bromine 69 isotope modelling approach to simultaneously predict the evolution of concentration, as well as 70 71 carbon and bromine isotopic signals. Our work focuses on chemical and biotic transformations of 72 EDB with the specific goals to: (i) describe a mechanism-based integrated modelling approach to simulate carbon and bromine isotope fractionation; (ii) validate the model with the isotopic data 73 74 observed during EDB degradation reactions; (iii) illustrate the capabilities of the model based on scenarios of complex EDB degradation pathways, including multistep reactions and parallel 75 degradation pathways, and considering the evolution of dual C and Br isotope signals not only of 76 EDB but also of its degradation intermediates and products. 77

78

79 2. Modelling approach

80 2.1. Degradation pathways and reaction mechanisms

We focus on EDB degradation through two important degradation pathways, dibromoelimination 81 and nucleophilic substitution (S_N2). The two degradation pathways can occur both chemically and 82 biotically. Dibromoelimination occurs during reduction of EDB with Zn(0) in aqueous solution, as 83 well as during biotic transformation by Sulfurospirillum multivorans. A stepwise nucleophilic 84 substitution may take place in aqueous alkaline solution and also occurs during biotic 85 86 transformation by Ancylobacter aquaticus [24]. The two pathways involve different bond-cleavage mechanisms. Dibromoelimination is assumed to result in simultaneous cleavage of two C-Br bonds, 87 while $S_N 2$ reaction follows a stepwise cleavage of one C-Br bond [5,24]. 88

90 2.2. Pathway-specific reaction rates and isotope fractionation

91 In order to simulate carbon and bromine isotopic evolution of EDB via different reactions, we track 92 dual element isotopologues. The relative abundances of such isotopologues can be computed 93 considering the occurrence of both stable carbon and bromine isotopes:

(1) 94 where *A* is the relative abundance of the j^{th} EDB isotopologue containing a^{13} C out of a total of two 95 carbon atoms and b^{81} Br out of a total of two bromine atoms. *X* and *Y* are the abundance of heavy 96 carbon and bromine isotopes, respectively.

97 Position specific fractionation factors for the j^{th} EDB isotopologue can be calculated according to 98 the corresponding apparent kinetic isotope effect (AKIE) derived from the observed bulk 99 enrichment factors:

> (2) (3)

100 where α_{rp} is the fractionating factor at reactive position, ε is bulk enrichment factor, and z is the 101 number of carbon or bromine atoms at reactive positions. In this work we calculated α_{rp} based on 102 the AKIE values reported in Kuntze et al. (2016); however, α_{rp} could be also derived by fitting the 103 proposed model to the raw isotope data.

We track the concentration change of each isotopologue considering a specific kinetic rate law. To 104 illustrate the approach, a first-order kinetic formulation is considered in the following equations; 105 however, as discussed for the application examples in Section 3, any degradation rate can be 106 implemented, including Michaelis-Menten kinetics. Since different reaction mechanisms of EDB 107 involve carbon and bromine atoms located at different isotopically-sensitive positions, the reaction 108 rates have to take into account all the fractionating atoms. Concerning the dibromoelimination 109 reaction, the two C-Br bonds are cleaved simultaneously, and thus the reaction rate for a specific 110 111 carbon-bromine isotopologue is given as:

(4) 112 where r_j is the reaction rate for the j^{th} isotopologue, k is the first-order reaction rate constant, C_j is 113 the concentration of the j^{th} isotopologue, α_{rp} is the fractionation factor as defined in Eqs. (2) and (3).

Considering S_N2 reaction of EDB, such reaction pathway involves the cleavage of one single C-Br 114 bond. In this case the reaction rate of EDB depends on the isotopic composition of the C-Br bond 115 that is cleaved, and thus the reaction rate of the individual isotopologues is defined in a bond-116 specific manner as previously proposed for the carbon-chlorine isotope modelling of chlorinated 117 ethenes [30]. Due to the fact that the two C-Br bonds of EDB are chemically equivalent for $S_N 2$ 118 reaction, the approach is based on isotopologues without the need of specifying individual 119 isotopomers. Thus, the reaction rate, r_i , for a given j^{th} EDB isotopologue is expressed as the sum of 120 the following bond-specific reaction rates: 121

- (5)
- (6)
- (7)
- (8) (9)

where k is the first-order rate constant, C_j is the concentration of the j^{th} isotopologue, and n_{rp} represents the total number of reactive carbon or bromine atoms within the isotopologue, *i* indicates the C-Br bond cleaved during the reaction and N is the total number of C-Br bonds that can be cleaved for the j^{th} EDB isotopologue. Note that an overall rate for all isotopologues can be computed from the rate of each isotopologue as , where *m* is the total number of isotopologues. The concentration change of the j^{th} isotopologue of EDB is described as:

(10)128 The total concentration of EDB can be obtained by summing the concentrations of each129 isotopologue:

130

(11)

131 where C_{tot} is the total concentration of EDB, C_j is the concentration of the j^{th} isotopologue and m is 132 the total number of EDB isotopologues.

133 The concentration of each isotopologue is used to calculate stable carbon and bromine isotope ratios134 by considering the total number of heavy and light isotopes [14] and are expressed as:

(12)

(13) 135 where R_C and R_{Br} are the carbon and bromine isotope ratios of EDB, C_j is the concentration of the j^{th} 136 isotopologue, *m* is the total number of EDB isotopologues as defined in Eq (11), *a* and *b* are the 137 number of heavy carbon and heavy bromine isotopes, as defined in Eq (1).

138 2.3. Complex Reaction Pathways

In the previous section we illustrated the modelling for single step reactions. However, the model can be applied also when degradation occurs through more complex reaction pathways involving sequential and parallel reactions. In this cases, if one aims at describing the formation and consumption of intermediates and products and the evolution of their dual-element isotopic composition, it is necessary to take into account that a given intermediate (or product) can be formed by two distinct isotopologues of the parent compound (or intermediate).

Sequential multistep reactions. We consider EDB degradation through sequential multistep reactions, specifically, through a reaction pathway involving two S_N2 type reactions. As parent compound (*P*) the two different EDB isotopologues considered are: the *j*th isotopologue and the (*j*+1)th isotopologue. The latter contains one more ⁸¹Br isotope in the molecule compared to the *j*th isotopologue. The parent compound (*P*) is sequentially degraded into the *k*th and (*k*+1)th isotopologues of the intermediate (*I*), and finally into the *i*th isotopologue of the end product (*E*), which is completely debrominated. The two-step reaction can be illustrated as:

$$\begin{array}{c} P_{j} \xrightarrow{r_{IJ}} & I_{k} \xrightarrow{r_{2,k}} & E_{j} \\ P_{j+1} & I_{k+1} \end{array}$$

153

(14)

154 Note that the different letters used as subscripts indicate that the different compounds may have a 155 different number of isotopologues.

The concentration of the isotopologues of the parent compound (P), the intermediate (I) and the end product (E) are described:

(15)

- (16)
- (17)

where $r_{l,j}$, $r_{l,j+1}$, $r_{2,k}$ and $r_{2,k+1}$ are the isotopolgue-specific reaction rates for the parent compound and for the intermediate, respectively. The kinetic formulation for such reaction rates $r_{l,j}$ and , $r_{l,j+1}$ (parent compound), as well as $r_{2,k}$ and $r_{2,k+1}$ (intermediate) are based on Eqs. 5-9. The carbon and bromine isotope ratios for the parent compound, intermediate and end product can be calculated according to Eqs. 12-13.

<u>Parallel reactions</u>. Competition between different reaction pathways of EDB degradation has been
observed in several experimental studies [5,24]. We consider the case of EDB transformation
through two competing reaction pathways that yield two different products:

$$P_{j} \xrightarrow{r_{A,j}} E_{A,k}$$

$$P_{j+1} \xrightarrow{r_{B,j+1}} E_{B,i}$$

$$(18)$$

166

167 The concentration of the j^{th} isotopologues of the parent compound (P_j) and the k^{th} and i^{th} of the two 168 end products $(E_{A,k} \text{ and } E_{B,i})$ are given as:

(20) (21)

169 where $r_{A,j}$, $r_{A,j+1}$, $r_{B,j}$ and $r_{B,j+1}$ are the reaction rates for the individual reaction pathway of the j^{th} and 170 $(j+1)^{\text{th}}$ isotopologue of the parent compound (*P*).

171 2.4. Model implementation

The governing equations describing the simultaneous evolution of the concentrations, as well as the 172 carbon and bromine isotope ratios are implemented in MATLAB®. The system of ordinary 173 differential equations is solved numerically using the function ode15s. The experimental data and 174 175 the key isotope fractionation parameters are taken from the experimental work of Kuntze et al. (2016); the latter are summarized in Table 1. The simulation was run for a time covering the 176 duration of the experiments (i.e., 4 hours for the two cases of dibromoelimination by both Zn(0) and 177 178 S. multivorans, 350 hours for abiotic degradation in alkaline solution and 8 hours in the case of 179 biotic degradation by A. aquaticus). The EDB concentration data were used to determine the kinetic parameters of the degradation rates. First-order and Michaelis-Menten kinetics were considered for 180 181 the abiotic and biotic reaction pathways, respectively. A fitting procedure, minimizing the sum of normalized squared errors based on the function lsqnonlin, was used to obtain the values of the 182 kinetic parameters. As illustrated above, the proposed approach tracks the dual-element EDB 183 isotopologues. Nine EDB isotopologues were considered in the simulations by taking into account 184 all possible combinations of carbon and bromine isotopes. The abundances of these isotopologues 185 186 were determined based on Eq. 1.

187 [insert Table 1 here]

188

189 **3. Results and discussion**

190 3.1. Chemical and biotic dibromoelimination reactions

The dual carbon and bromine isotope approach has been used to investigate EDB degradation by 191 dibromoelimination reactions [24]. To reproduce the experimental data observed during 192 dibromoelimination reactions, we simulated carbon and bromine isotopic evolution according to the 193 hypothesized two-electron transfer dibromoelimination mechanism. The simulation results (solid 194 lines in Fig. 1) are shown together with the reported experimental data (symbols in Fig. 1). A first-195 order kinetic (k=0.9 h⁻¹) is used to describe the concentration variation of EDB during 196 dibromoelimination by Zn(0) (Fig. 1a), where the concentration decreases down to 4.3% of the 197 initial concentration value. The model also accurately predicts the carbon and bromine isotope 198 fractionation (Fig. 1b), which are simulated based on the experimentally evaluated AKIE values 199 $(AKIE_{C}=1.0223, AKIE_{Br}=1.0042)$ [24]. The results show different extents of carbon and bromine 200 fractionation, with δ^{13} C values changing from -26.3‰ to -3.8‰ and δ^{81} Br varying from 0.5‰ to 201 5.4‰. For biotic dibromoelimination, a Michaelis-Menten kinetics, with maximum degradation rate 202 k_{max} =0.2289 mmol·L⁻¹·h⁻¹ and half-saturation constant K_s =0.0166 mmol·L⁻¹, is used in our model to 203 reproduce the observed concentration data during biotic dibromoelimination by S. multivorans (Fig. 204 1c). The carbon and bromine AKIE values of 1.0107 and 1.0046 are used in the model to describe 205 206 carbon and bromine isotope effects. The fractionation was introduced in the maximum degradation rate and led to an increase of 12.7‰ for ¹³C and of 6.2‰ for ⁸¹Br isotopes during degradation of 207 about 95% of the initial EDB concentration. 208

209 [insert Figure 1 here]

Linear dual carbon-bromine isotopic trends with different slopes are obtained for chemical (slope of 5.3) and biotic (slope of 2.4) dibromoelimination reactions and the model accurately captures the two different dual-isotope trends. The excellent agreement between experimental and modelling results demonstrates the capability of the proposed mechanistic model to simultaneously capture theevolution of both concentration and carbon-bromine stable isotopes.

215 3.2. Chemical and biotic nucleophilic substitution (S_N 2) reactions

Nucleophilic substitution $(S_N 2)$ reaction is another important degradation mechanism for EDB. A stepwise scenario is followed by both biotic and chemical $S_N 2$ reactions, where the cleavage of one carbon-bromine bond of EDB is hypothesized as the isotopically sensitive step. We provide a model-based interpretation of the experimental data provided in the study of Kuntze et al. [24], who observed carbon and bromine isotope fractionation of EDB during chemical degradation in aqueous alkaline solution as well as during biotic $S_N 2$ reaction by *Ancylobacter aquaticus*.

222 [insert Figure 2 here]

We use a first-order kinetics (k=0.0125 h⁻¹) to describe the concentration change during the 223 chemical S_N2 reaction (solid line in Fig. 2a). The corresponding carbon isotope ratio varies from 224 -11.6‰ to 93.8‰, and the bromine isotope fractionation occurs in a range between 0.4‰ and 225 226 4.3‰. The biotic S_N2 reaction of EDB is described by a Michaelis-Menten kinetics with maximum degradation rate ($k_{max}=0.4329$ mmol·L⁻¹·h⁻¹) and half-saturation constant ($K_s=0.0288$ mmol·L⁻¹) 227 228 evaluated based on the observed concentration data (Fig. 2c). The simulation of carbon and bromine isotope signals is based on the reported AKIE values ($AKIE_{c}=1.062$ and $AKIE_{Br}=1.002$ for the 229 chemical S_N2 reaction; $AKIE_c=1.014$ and $AKIE_{Br}=1.0012$ for the biotic S_N2 reaction). The 230 simulations of the biotic and chemical S_N2 transformations of EDB were able to capture the 231 different fractionation of the two reaction pathways observed in the experiments. Specifically, the 232 biotic transformation resulted in a smaller extent of both carbon and bromine isotope fractionation 233 (17.9‰ for $\delta^{13}C$ and 1.8‰ for $\delta^{81}Br)$ whereas the chemical $S_{\rm N}2$ reaction resulted in stronger 234 fractionation (105.4‰ for δ^{13} C and 3.9‰ for δ^{81} Br). 235

Fig. 3 summarizes the dual element isotope plots for the four cases of chemical and biotic EDB 237 degradation through dibromoelimination and S_N2 type nucleophilic substitution. The four different 238 reactions are adequately characterized in the dual carbon-bromine isotope plot. In all the cases the 239 240 simulation outcomes closely reproduce the experimental results. Note that these outcomes are no 241 linear fits of the experimental data, but represent mechanistic descriptions of EDB degradation 242 through different reaction pathways according to the approach outlined in Section 2. For all considered cases of EDB degradation the normalized root mean squared error was calculated as a 243 244 quantitative measure of the goodness-of-fit. Such metric was computed for both carbon and bromine isotope data and yielded values in a range of 0.036-0.158 for carbon and 0.068-0.16 for 245 bromine. The successful comparison of the simulation results with the experimental data highlights 246 the capability of the proposed approach to quantitatively describe different mechanisms of EDB 247

248 degradation.

249 [insert Figure 3 here]

250 3.3. Scenario modelling

251 Based on the validated model presented above, we also investigated scenarios involving complex 252 EDB reaction pathways, such as sequential multistep reactions (Scenario 1 in Fig. 4) and parallel reactions (Scenario 2 in Fig. 4). In the examples illustrated in the previous section and in most 253 254 experimental studies, the CSIA approach has been mainly focusing on the parent compound. However, stable isotope analysis of reaction products can also be very informative about the 255 underlying reaction steps characterizing different reaction mechanisms [22,31]. To explore the 256 potential of carbon and bromine CSIA of EDB degradation products, we simulate the evolution of 257 the concentration and the isotopic signals of the parent compound, the intermediates and the end 258

products for the two proposed reaction scenarios illustrated in Fig. 4: 1) multistep $S_N 2$ nucleophilic substitution and 2) simultaneous occurrence of the $S_N 2$ reaction and dehydrobromination.

261 [insert Figure 4 here]

262 The S_N2 reaction involves the stepwise cleavage of two C-Br bonds, kinetic isotope effects for C-Br cleavage ($KIE_{C}=1.042$; $KIE_{Br}=1.002$) were calculated in the previous experimental study based on 263 the Streitwieser limit [24]. Since isotope fractionation of the intermediate has not been 264 265 experimentally determined (yet), the theoretical KIEs values are used as model input parameters to differentiate the reaction rates of the different carbon-bromine isotopologues of both parent and 266 intermediate compounds. The simulation results for sequential multistep EDB degradation 267 (Scenario 1) are shown in Fig. 5. The degradation of the parent compound EDB (blue solid line in 268 Fig. 5a) results in the formation of the intermediate, bromoethylene glycol (red dotted line, k=0.5 h⁻ 269 ¹), which is further transformed to ethylene glycol (green dash-dotted line, k=2.5 h⁻¹) that 270 accumulates as the end product. The temporal carbon and bromine isotope trends are reported in 271 Fig. 5b and 5c and show a linear increase of δ^{13} C and δ^{81} Br values for EDB. However, the 272 increasing trends of carbon and bromine isotope ratios become nonlinear for the intermediate, 273 bromoethylene glycol. This is due to the fact that bromoethylene glycol (red dotted line) further 274 degrades and preferentially transfers ¹²C isotopes to the end product and meanwhile preferentially 275 releases ⁷⁹Br during its transformation. As a result, the δ^{13} C values of the end product, ethylene 276 glycol (green dash-dotted line), continuously increase and approach the original carbon isotope 277 278 signature of EDB. In the dual carbon-bromine isotope plot (Fig. 5d) EDB and bromoethylene glycol have different trends. EDB shows a linear increase with a slope of 20.2, whereas a nonlinear curve, 279 with a slope varying from 11 to 21.5, describes the trend of bromoethylene glycol. This nonlinear 280 behaviour is due to the simultaneous formation and consumption of bromoethylene glycol, which 281 occur at different rates and involve different extents of carbon and bromine isotope fractionation 282

during the course of the degradation reaction. For multistep reactions with formation and further degradation of intermediates, a mechanistic modelling approach is helpful since it allows the simultaneous interpretation of isotope fractionation for both precursors and reaction products.

286 [insert Figure 5 here]

In Scenario 2, degradation of EDB in alkaline solution is considered. In this scenario two competing 287 reaction pathways, i.e., nucleophilic substitution (S_N2) and dehydrobromination, occur 288 289 simultaneously. Concerning dehydrobromination, this reaction pathway involves the simultaneous cleavage of a carbon-bromine bond and a carbon-hydrogen bond. The theoretical carbon and 290 291 bromine isotopic effects during cleavage of C-H ($KIE_c=1.021$) and C-Br ($KIE_c=1.042$; KIE_{Br} =1.002) bonds [24,36] are considered to calculate the fractionation factors at reactive positions 292 used in the isotopologue-specific rate expression (Eq. 4). We applied the model to simulate 293 concentrations and isotope ratios for such a parallel reaction system. We assume that the two 294 reactions follow a first-order kinetic with rate constants of 0.5 h⁻¹ and 0.03 h⁻¹ for S_N2 reaction and 295 dehydrobromination, respectively. These values were selected according to the relative contribution 296 of 93% (sequential S_N2 reaction) and 7% (dehydrobromination) observed in the experimental study 297 of Kuntze et al. [24]. The evolution of concentration, carbon and bromine isotopic signals is 298 simultaneously simulated for EDB, the intermediate and the end products. As shown in Fig. 6a, the 299 300 two competing degradation reactions cause a decrease of the EDB concentration (blue solid line). The intermediate, bromoethylene glycol (red dotted line), is formed and further degrades into 301 ethylene glycol (green dash-dotted line) by nucleophilic substitution (S_N2). In parallel, 302 dehydrobromination causes the formation of vinyl bromide (black solid line). Fig. 6b illustrates the 303 δ^{13} C trends for the species involved in the two reaction pathways: the parent compound shows a 304 linear behaviour, whereas the intermediate and the end products show nonlinear curves with 305 decreasing slope. Bromine stable isotope ratios are shown in Fig. 6c. Stable bromine isotope 306

fractionation occurs at different extents for the two brominated degradation products: δ^{81} Br is 307 enriched by 7.6‰ for bromoethylene glycol (S_N2 reaction) and by 1‰ for vinyl bromide 308 (dehydrobromination), because the former is an intermediate which undergoes further 309 debromination, whereas vinyl bromide represents a final product in this scenario. In the dual-310 isotope plot (Fig. 6d) a linear trend is obtained for the parent compound EDB (slope: 21.2), as well 311 as for vinyl bromide from the dehydrobromination reaction (slope: 20.3). A nonlinear dual-isotope 312 trend is obtained for the S_N2 reaction intermediate, bromoethylene glycol, with a slope varying from 313 11.1 to 22.9. The dual isotope trend of EDB (blue line in Fig. 6d) appears very similar with the one 314 obtained during 100% S_N2 reaction (lower dashed line). This is due to the fact that, in the 315 considered scenario, the S_N2 reaction is the dominant pathway (about 93% contribution) during 316 317 EDB degradation in alkaline solution. The shaded grey area between the dotted lines indicates the possible range for the investigated scenario: from 100% contribution of dehydrobromination (upper 318 bound) to 100% contribution of the S_N2 reaction (lower bound). The dual element isotope 319 signatures of the reaction products from the different pathways have a distinct behaviour and are 320 different from the trend of the parent compound. The simulation results indicate that in practice it 321 322 might be difficult to accurately quantify the contribution of each concurrent reaction pathway exclusively based on the observed EDB carbon and bromine isotope data. However, these 323 simulations also demonstrate that CSIA of reaction intermediates and end products can bring new 324 possibilities to elucidate the underlying reaction steps and to accurately quantify the contributions 325 326 of individual reaction pathways.

327 [insert Figure 6 here]

328 4. Conclusion

Dual carbon-bromine isotope investigation significantly improves the understanding of various 329 reaction mechanisms of brominated organic compounds. Recent studies have focused on the 330 development of analytical techniques for bromine CSIA as well as experimental investigation of 331 degradation mechanisms of different brominated organic pollutants. In this study, we have proposed 332 333 an integrated modelling approach allowing the simultaneous prediction of concentrations and dualelement isotope ratios. Our investigation focused on carbon and bromine isotope fractionation 334 during 1.2-dibromoethane (EDB, ethylene dibromide) transformation through different reaction 335 pathways. The proposed modelling approach tracks dual-element isotopologues. The method 336 allowed us reproducing the carbon and bromine isotopic signal observed in experimental studies of 337 different chemical and biotic EDB reaction pathways. The approach is based on bond-specific 338 339 reaction rates and can be readily extended to cases in which different contaminants and/or reaction pathways will require tracking also isotopomers (i.e., molecules with the same number of each 340 isotopic atom but differing in their position). Furthermore, we exemplified the model capabilities 341 with two scenarios involving complex reaction systems with sequential and parallel reactions, 342 respectively. In the considered case of multistep nucleophilic substitution (S_N2) reaction, 343 344 concentration and isotopic ratios of the parent compound, the intermediate and the end product were 345 predicted based on the validated model. Different carbon and bromine isotopic behaviours of the parent compound and intermediate were obtained. In the scenario modelling of EDB degradation 346 via two concurrent reactions, our simulation results showed that stable isotope analysis of the 347 348 reaction products is beneficial, since it allows quantifying further degradation of the intermediate product in multistep reactions, as well as providing a more accurate evaluation of the individual 349 350 contributions of different concurrent pathways to the overall reaction. This is particularly beneficial 351 when one of the reaction pathways is dominant and, therefore the sole analysis of the dual isotope trend of the parent compound would not be conclusive in identifying which degradation reactions 352

are responsible for the contaminant degradation. The proposed model was applied to the specific case of EDB degradation, however it provides a framework that can be extended to other brominated compounds that may undergo degradation through different reaction pathways. The first-principle based formulation of the approach will also facilitate future model-based applications in complex environmental systems, in which both transformation and mass transfer processes may affect the observed isotope signals.

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Table 1. Reaction mechanisms, bulk enrichment factors (ε_{bulk}) and fractionation factors at reactive position (α_{rp}) for the different EDB degradation reactions.

	Reaction	Mechanism	Ebulk		α _{rp}	
			С	Br	С	Br
		dibromoeliminatio				
	Zn (0)	n	-10.9 ± 1.1	-2.1 ± 0.3	$0.9891 {\pm} 0.0011$	0.9979±0.0003
		dibromoeliminatio				
	S. multivorans	n	-5.3 ± 0.5	-2.3 ± 0.2	0.9947 ± 0.0005	0.9977 ± 0.0002
	Alkaline solution	abiotic S _N 2	-29.2±2.6	-1.0 ± 0.1	$0.9416 {\pm} 0.0052$	0.9980 ± 0.0002
	A. aquaticus	biotic S _N 2	-6.9±0.4	-0.6±0.1	$0.9862 {\pm} 0.0008$	$0.9988 {\pm} 0.0002$
383						
204						
384						
385						
386						
500						





Figure 1. Concentration change and dual carbon-bromine isotope fractionation during dibromoelimination reaction by Zn(0) (Panels (a) and (b)) and biotic reaction with *S. multivorans* (Panels (c) and (d)). The symbols represent the experimental data reported in Kuntze et al. [24], and the solid lines are the simulation results.





Figure 2. Carbon and bromine isotope fractionation during EDB chemical transformation in alkaline solution and biotic reaction by *Ancylobacter aquaticus*. Panel (a) and (c): the symbols represent the observed concentration profiles reported in Kuntze et al. [24], and the lines are the simulation results. Panel (b) and (d): the symbols are carbon-bromine isotopic data and the solid lines are the simulation results.



Figure 3. Carbon and bromine isotope fractionation for different EDB degradation reactions. The symbols represent the experimental data reported in Kuntze et al.[24], and the solid lines represent the results of the simulations corresponding to 99% degradation of the initial EDB concentration.

Scenario 1: Multistep reactions by nucleophilic substitution (S_N2)



Scenario 2: Parallel reactions in alkaline solution (nucleophilic substitution and dehydrobromination)



412 Figure 4. Schemes representing the sequential and parallel reaction pathways considered in the scenario413 modelling.





Figure 5. Concentration, carbon and bromine isotope fractionation of EDB, the intermediate (bromoethylene
glycol), the end product (ethylene glycol) and bromide during multi-step nucleophilic substitution (Scenario
1 in Fig. 4).





Figure 6. Concentration, carbon and bromine isotope fractionation of EDB during the two competing reaction pathways: nucleophilic substitution ($S_N 2$) reaction and dehydrobromination (Scenario 2 in Fig. 4). The shaded area in Panel (d) indicates EDB dual-isotope trends corresponding to different contributions of each reaction pathway considered in Scenario 2. The dotted lines on the upper and lower bounds represent dual-isotope trends of EDB degradation when one reaction pathway occurs exclusively (i.e., 100% dehydrobromination and 100% $S_N 2$ reaction, respectively).

436 References

- Falta RW, Bulsara N, Henderson JK, et al. Leaded-gasoline additives still contaminate groundwater.
 Environ. Sci. Technol. 2005;39:379A–384A.
- Wilson, John T, Banks, Kenneth, Earle, Robert, He, Yongtian, Kuder, Tomasz, Adair C. Natural
 attenuation of the lead scavengers 1,2-dibromoethane (EDB) and 1,2-dichloroethane (1,2-DCA) at
 motor fuel release sites and implications for risk management. U.S. EPA. 2008.
- Yu R, Peethambaram HS, Falta RW, et al. Kinetics of 1,2-dichloroethane and 1,2-dibromoethane
 biodegradation in anaerobic enrichment cultures. Appl. Environ. Microbiol. 2013;79:1359–1367.
- 444 [4] McKeever R, Sheppard D, Nüsslein K, et al. Biodegradation of ethylene dibromide (1,2445 dibromoethane [EDB]) in microcosms simulating in situ and biostimulated conditions. J. Hazard.
 446 Mater. 2012;209–210:92–98.
- Kuder T, Wilson JT, Philp P, et al. Carbon isotope fractionation in reactions of 1,2-dibromoethane
 with FeS and hydrogen sulfide. Environ. Sci. Technol. 2012;46:7495–7502.
- 449 [6] Bosma TNP, Middeldorp PJM, Schraa G., Zehender AJB. Mass Transfer Limitation of
 450 Biotransformation: Quantifying Bioavailability. Environ. Sci. Technol. 1997;31:248–252.
- 451 [7] Rolle M, Kitanidis PK. Effects of compound-specific dilution on transient transport and solute
 452 breakthrough: A pore-scale analysis. Adv. Water Resour. 2014;71:186–199.
- 453 [8] Schüth C, Taubald H, Bolaño N, et al. Carbon and hydrogen isotope effects during sorption of
 454 organic contaminants on carbonaceous materials. J. Contam. Hydrol. 2003;64:269–281.
- 455 [9] Nijenhuis I, Richnow HH. Stable isotope fractionation concepts for characterizing biotransformation
 456 of organohalides. Curr. Opin. Biotechnol. 2016;41:108–113.
- Elsner M, Imfeld G. Compound-specific isotope analysis (CSIA) of micropollutants in the
 environment current developments and future challenges. Curr. Opin. Biotechnol. 2016;41:60–72.
- 459 [11] Schmidt TC, Jochmann M a. Origin and Fate of Organic Compounds in Water: Characterization by
 460 Compound-Specific Stable Isotope Analysis. Annu. Rev. Anal. Chem. 2012;5:133–155.
- 461 [12] Rosell M, Gonzalez-Olmos R, Rohwerder T, et al. Critical evaluation of the 2D-CSIA scheme for
 462 distinguishing fuel oxygenate degradation reaction mechanisms. Environ. Sci. Technol.
 463 2012;46:4757–4766.
- 464 [13] Sakaguchi-Söder K, Jager J, Grund H, et al. Monitoring and evaluation of dechlorination processes
 465 using compound-specific chlorine isotope analysis. Rapid Commun. Mass Spectrom. 2007;21:3077–
 466 3084.
- 467 [14] Jin B, Laskov C, Rolle M, et al. Chlorine Isotope Analysis of Organic Contaminants Using GC–qMS:
 468 Method Optimization and Comparison of Different Evaluation Schemes. Environ. Sci. Technol.
 469 2011;45:5279–5286.
- 470 [15] Gelman F, Halicz L. High precision determination of bromine isotope ratio by GC-MC-ICPMS. Int.
 471 J. Mass Spectrom. 2010;289:167–169.
- 472 [16] Hitzfeld KL, Gehre M, Richnow HH. A novel online approach to the determination of isotopic ratios
 473 for organically bound chlorine, bromine and sulphur. Rapid Commun. Mass Spectrom.
 474 2011;25:3114–3122.

- 475 [17] Bernstein A, Shouakar-Stash O, Ebert K, et al. Compound-Specific Chlorine Isotope Analysis: A
 476 Comparison of Gas Chromatography/Isotope Ratio Mass Spectrometry and Gas
 477 Chromatography/Quadrupole Mass Spectrometry Methods in an Interlaboratory Study. Anal. Chem.
 478 2011;83:7624–7634.
- 479 [18] Audí-Miró C, Cretnik S, Torrentó C, et al. C, Cl and H compound-specific isotope analysis to assess
 480 natural versus Fe(0) barrier-induced degradation of chlorinated ethenes at a contaminated site. J.
 481 Hazard. Mater. 2015;299:747–754.
- 482 [19] Nijenhuis I, Kuntze K. Anaerobic microbial dehalogenation of organohalides-state of the art and
 483 remediation strategies. Curr. Opin. Biotechnol. 2016;38:33–38.
- 484 [20] Cretnik S, Thoreson K a., Bernstein A, et al. Reductive dechlorination of TCE by chemical model
 485 systems in comparison to dehalogenating bacteria: Insights from dual element isotope analysis
 486 (13C/12C, 37Cl/35Cl). Environ. Sci. Technol. 2013;47:6855–6863.
- 487 [21] Bernstein A, Ronen Z, Levin E, et al. Kinetic bromine isotope effect: Example from the microbial
 488 debromination of brominated phenols. Anal. Bioanal. Chem. 2013;405:2923–2929.
- 489 [22] Schmidt M, Lege S, Nijenhuis I. Comparison of 1,2-dichloroethane, dichloroethene and vinyl
 490 chloride carbon stable isotope fractionation during dechlorination by two Dehalococcoides strains.
 491 Water Res. 2014;52:146–154.
- 492 [23] Palau J, Cretnik S, Shouakar-Stash O, et al. C and Cl Isotope Fractionation of 1,2-Dichloroethane
 493 Displays Unique δ 13 C/δ 37 Cl Patterns for Pathway Identification and Reveals Surprising C–Cl
 494 Bond Involvement in Microbial Oxidation. Environ. Sci. Technol. 2014;48:9430–9437.
- 495 [24] Kuntze K, Kozell A, Richnow HH, et al. Dual Carbon–Bromine Stable Isotope Analysis Allows
 496 Distinguishing Transformation Pathways of Ethylene Dibromide. Environ. Sci. Technol.
 497 2016;50:9855–9863.
- 498 [25] Eckert D, Rolle M, Cirpka O a. Numerical simulation of isotope fractionation in steady-state
 499 bioreactive transport controlled by transverse mixing. J. Contam. Hydrol. 2012;140–141:95–106.
- Jin B, Rolle M, Li T, et al. Diffusive fractionation of BTEX and chlorinated ethenes in aqueous
 solution: Quantification of spatial isotope gradients. Environ. Sci. Technol. 2014;48:6141–6150.
- 502 [27] Van Breukelen BM, Rolle M. Transverse hydrodynamic dispersion effects on isotope signals in
 503 groundwater chlorinated solvents plumes. Environ. Sci. Technol. 2012;46:7700–7708.
- Hofstetter TB, Reddy CM, Heraty LJ, et al. Carbon and chlorine isotope effects during abiotic
 reductive dechlorination of polychlorinated ethanes. Environ. Sci. Technol. 2007;41:4662–4668.
- Funkeler D, Van Breukelen BM, Elsner M. Modeling chlorine isotope trends during sequential
 transformation of chlorinated ethenes. Environ. Sci. Technol. 2009;43:6750–6756.
- Jin B, Haderlein SB, Rolle M. Integrated carbon and chlorine isotope modeling: Applications to chlorinated aliphatic hydrocarbons dechlorination. Environ. Sci. Technol. 2013;47:1443–1451.
- [31] Kuder T, Van Breukelen BM, Vanderford M, et al. 3D-CSIA: Carbon, chlorine, and hydrogen isotope
 fractionation in transformation of TCE to ethene by a dehalococcoides culture. Environ. Sci. Technol.
 2013;47:9668–9677.
- 513 [32] Thullner M, Centler F, Richnow H-H, et al. Quantification of organic pollutant degradation in contaminated aquifers using compound specific stable isotope analysis Review of recent
 515 developments. Org. Geochem. 2012;42:1440–1460.

- 516 [33] Jin B, Rolle M. Mechanistic approach to multi-element isotope modeling of organic contaminant degradation. Chemosphere. 2014;95:131–139.
- Jin B, Rolle M. Position-specific isotope modeling of organic micropollutants transformation through
 different reaction pathways. Environ. Pollut. 2016;210:94–103.
- Jin B, Rolle M. Joint interpretation of enantiomer and stable isotope fractionation for chiral pesticides
 degradation. Water Res. 2016;105:178–186.
- 522 [36] Elsner M, Zwank L, Hunkeler D, et al. A new concept linking observable stable isotope fractionation
 523 to transformation pathways of organic pollutants. Environ. Sci. Technol. 2005;39:6896–6916.
- 524