

1 **Fish early life-stage toxicity and environmental relevance: What does**
2 **high-dose toxicity testing tell us?**

3
4 **Abstract**

5 The early-life stage (ELS) toxicity syndrome in fish is well described and has
6 been reported in hundreds of toxicity studies. It is generally characterized by a
7 reduced heart rate, yolk sac and pericardial edemas, and various morphological
8 abnormalities, the most common being spinal curvature. For many of those
9 studies it appears that the ELS toxicity syndrome is the result of non-specific
10 (baseline) toxicity that occurs at aqueous and whole-body concentrations that
11 are just below lethal concentrations. Baseline toxicity is essentially a non-
12 specific response that results from chemicals accumulating in and disturbing
13 function of biological membranes that leads to lethality and sublethal effects at
14 relatively high doses. The commonality of this acute ELS toxicity syndrome
15 among highly diverse organic and inorganic chemicals is remarkable. It is
16 important to identify baseline toxicity because it is considered minimal toxicity
17 that acts in all tissues and cells, and it has the potential to impair all cellular
18 functions. This means if an effect is observed around baseline-toxic
19 concentrations, it is likely that other cellular functions are also affected, i.e., the
20 effect is not specific. The fish ELS toxicity syndrome can also be the result of
21 specific effects involving receptor interactions; therefore, we emphasize the
22 importance of distinguishing between specific and non-specific toxicity
23 responses to provide the most relevant data for environmental risk assessment.

24
25
26
27

28 Introduction

29 There are hundreds, and possibly a few thousand studies that describe toxic
30 effects for fish at high-dose exposure concentrations. Many of these studies are
31 conducted with fish embryos, commonly zebrafish (*Danio rerio*), which are easy
32 to raise and maintain in a laboratory setting. Zebrafish provide a constant
33 supply of eggs and embryos that can be acquired in large numbers and
34 subjected to rapid toxicity tests in shallow, low-volume plate wells, which can
35 exhibit highly variable concentrations (Truong et al. 2011; Knobel et al. 2012).

36

37 A common theme for these high-dose studies with early life-stage (ELS)
38 zebrafish (embryos and larvae) concerns developmental effects, such as
39 pericardial and yolk sac edemas, bradycardia, spinal and facial deformities, and
40 mortality. These effects have been considered as a syndrome and in most cases
41 are correlated and occur together at relatively high doses (Ducharme et al.
42 2013; Jarque et al. 2020). As noted by others, effects on the heart and
43 circulation can result in downstream effects causing edemas and morphological
44 abnormalities due to disruption in peripheral circulation (Incardona et al. 2004).
45 There are many compounds that can cause teratogenic effects in fish; however,
46 the ELS toxicity syndrome discussed here refers to the combination of a reduced
47 heart rate, edemas, and morphological deformities at high exposure doses that
48 likely result from non-specific cell toxicity. In these cases, this syndrome of
49 effects is caused by baseline toxicity, which occurs at constant critical tissue
50 concentrations. Specific effects occur at lower concentrations than baseline
51 toxicity. Classification of specific mechanisms or modes of toxic action in fish
52 have been addressed in several studies (Scholz et al. 2018; Zhu et al. 2018;
53 Wilhelmi et al. 2024) and will not be discussed here.

54 Baseline toxicity has been described and addressed by several authors (McCarty
55 1993, Escher et al. 2002; Meador et al. 2011; Klüver et al. 2016; Escher et al.
56 2020b; Lee et al. 2021). Baseline toxicity is a result of accumulation of organic
57 compounds in tissues to critical levels triggering a non-specific membrane effect

58 that destabilizes cell function. As it is a physical effect in the membranes, it is
59 not dependent on the structure of the chemical but on the molar volume and
60 can be approximated by a constant membrane concentration. Its universality is
61 striking and extremely difficult to refute. It is usually best considered in terms of
62 whole-body (internal) or membrane concentrations, which mitigates the high
63 variability of external exposure concentrations due to organismal toxicokinetics
64 and physicochemical properties (Escher and Hermens 2004; Meador et al.
65 2008). Toxicity metrics based on water exposure concentrations can vary by
66 orders of magnitude among species and organic compounds, especially as a
67 function of their hydrophobicity, whereas toxicity based on whole-body
68 concentrations is far less variable. Most studies evaluating the baseline toxic
69 response conclude that mortality for a high percentage of species and organic
70 chemicals occurs at a mean concentration of approximately 2.7 mmol/kg_{whole-body}
71 wet weight (95%CI = 0.5 – 15 mmol/kg_{whole-body wet weight}), with sublethal effects
72 occurring at concentrations between 1 and 10 fold lower (McCarty et al. 2013;
73 Scholz et al. 2018; Kluver et al. 2016). This relatively constant and high-dose
74 tissue concentration among fish species and compounds has been directly
75 demonstrated and modeled for hundreds of organic compounds. For many
76 organic compounds with molecular weights commonly in the range of 250
77 Daltons, baseline mortality in fish occurs at concentrations greater than 125
78 mg/kg_{whole-body wet weight} and sublethal effects above 13 – 25 mg/kg_{whole-body wet weight}
79 (assuming a 5-10 fold lower response level). These predictions are variable due
80 to compound characteristics, species, time for the response to develop, duration
81 of exposure, compound molecular weight, health of the organism, organismal
82 lipid content, and other relevant factors. Our goal here is to highlight the
83 misidentification of baseline toxicity and provide guidance on how to tell the
84 difference between baseline toxicity and adverse responses occurring at lower
85 exposure doses due to specific (receptor-mediated) action or reactive toxicity.

86

87 **Analysis**

88 The commonality of studies reporting this ELS toxicity syndrome of effects for a
89 wide diversity of organic compounds leads to a simple hypothesis that these
90 effects are the result of a non-specific response by the organism exposed to
91 doses high enough to result in critical tissue concentrations. In most cases, this
92 response can be attributed to baseline toxicity, which is also known as
93 "narcosis". The underlying syndrome of effects is disturbance of cellular
94 membrane structure and functioning (van Wezel 1995), leading to cytotoxicity,
95 which manifests in whole-organism toxicity tests as well as *in vitro* cellular
96 assays (Escher et al. 2020b). This observation is a good example of Occam's
97 razor, that prescribes a simple explanation over one that is more complex based
98 on speculation about toxicological function. Given the commonality of this
99 response, the working hypothesis is that most of these high-dose studies are
100 reporting on a non-specific response that occurs at doses slightly below those
101 causing mortality. In many cases the results are provocative showing severe
102 effects and a variety of physiological, morphological, and gene expression
103 alterations, which is not surprising for organisms that are near death.
104 Differential expression of cardiac or developmental genes are not proof of a
105 specific effect causing the ELS syndrome because such alterations are expected
106 given the near lethal, high-dose exposure. A common theme among many of
107 these studies is oxidative stress, which is the hallmark of cytotoxicity
108 (Chakraborty et al. 2023; Simmons et al. 2009). With *in vitro* toxicology, this
109 observation has been named "cytotoxicity burst" (Fay et al. 2018; Escher et al.
110 2020b).

111

112 It is no coincidence that many of the compounds causing the ELS toxicity
113 syndrome fall along the regression line defining aqueous exposure,
114 bioaccumulation, and toxicity as a function of hydrophobic partitioning
115 (Konemann 1981; Kluver et al. 2016; Scholz et al. 2018; Meador 2021). This
116 partitioning can be expressed by the octanol-water partition constant K_{ow} for
117 neutral chemicals (Figure 1), which forms a quantitative structure-activity

118 relationship (QSAR). For ionizable compounds, the K_{ow} can be simply replaced
119 with the lipid membrane-water distribution ratio (D_{lipw}) without substantially
120 changing the equations (Kluver et al. 2019; Escher et al. 2020a). The key point
121 here is that compounds along this regression line bioaccumulate to critical levels
122 triggering the baseline toxicity response. For many compounds this occurs at
123 relatively high aqueous concentrations (“high dose”); however, for hydrophobic
124 compounds this can occur at lower external aqueous concentrations.

125

126 While the vast majority of studies reporting biological effects consistent with the
127 fish ELS toxicity syndrome are due to baseline toxicity, this syndrome can also
128 be observed at lower exposure doses and is not the result of baseline toxicity.
129 The concentration for organic compounds causing toxic responses by receptor-
130 specific interaction is generally well below values predicted with the baseline
131 toxicity QSAR. Many of these are due to specific action on mitochondrial
132 processes that produce very similar phenotypic responses to those non-specific
133 responses (Meador, 2021). As hypothesized by Meador (2021), the high-dose
134 baseline toxicity response also manifests in mitochondrial membrane disruption
135 leading to altered calcium handling, which affects the developing heart and
136 downstream effects such as edema and morphological abnormalities. It has also
137 been shown that the specific effect of uncoupling of oxidative phosphorylation,
138 which is caused by a protonophoric shuttle mechanisms can transition into
139 baseline toxicity depending on the membrane concentration of toxicant needed
140 to trigger the observed response (Escher and Schwarzenbach 2002). Examples
141 of chemicals causing the ELS toxicity syndrome by specific action include
142 cyanide, tralopyril, tributyltin, and retinoic acid (Meador 2021) (Figure 1).

143

144 Additionally, the ELS toxicity syndrome has been demonstrated for many
145 chemicals at baseline toxicity concentrations and for different toxicity responses
146 at far lower exposure concentrations. A good example of this can be seen in
147 Meador (2021) for ethinylestradiol (EE2), which causes the ELS toxicity

148 syndrome at baseline concentrations and specific receptor-mediated effects at
149 concentrations approximately 5 to 6 orders of magnitude lower. Similar
150 examples can be seen in Meador (2021) for atrazine, carbamazepine, bisphenol
151 S, and several other chemicals.

152

153 Of course, membrane disruption is not the only mechanism that is consistent
154 with the ELS toxicity syndrome. In general, if these symptoms occur at lower
155 membrane or whole-body concentrations than observed for baseline toxicity,
156 they must be specific. Other low-dose effects directly affecting the heart or
157 related physiological pathways may result in the syndrome of responses similar
158 to those caused by baseline toxicity. One example is activation of the aryl
159 hydrocarbon receptor (AhR) (Shankar and Villeneuve 2023) that can result in
160 the ELS toxicity syndrome. A good example of these is dioxin and dioxin-like
161 compounds such as PCBs and 3-methylcloanthrene (Seok et al. 2008). Also,
162 compounds that act specifically on cardiac heart rate and rhythms can result in
163 the ELS syndrome at low exposure concentrations (D'Amico et al. 2012). The
164 problem is that there are far too many reports of high-dose exposures that are
165 likely the result of baseline toxicity, which incorrectly conclude that toxicity is
166 caused by a specific, receptor-mediated response, such as AhR activation.

167

168 Even though baseline toxicity is commonly applied to organic compounds, the
169 syndrome of effects for ELS fish can also be seen for metals, microplastics, and
170 nanoparticles. This response for non-organic compounds is also a product of
171 non-specific cytotoxicity. For example, Chen et al. (2024) reported bradycardia,
172 edemas, and morphological abnormalities in zebrafish resulting from exposure
173 to polystyrene particles. They attributed these responses to oxidative stress and
174 increased levels of reactive oxygen species (ROS). Several metals are also
175 known to cause the ELS toxicity syndrome with observations of the same suite
176 of developmental abnormalities and bradycardia. As noted in a recent review by
177 Rana (2020), elements such as arsenic, cadmium, lead, mercury, copper,

178 chromium, and nickel, in addition to nanoparticles, are known to cause
179 endoplasmic reticulum stress, which is suspected in causing the ELS toxicity
180 syndrome (Meador, 2021). The toxicity syndrome has also been reported for
181 zebrafish embryos exposed to algal extracts (González-Penagos et al. 2024).
182 This commonality of effects for organic compounds, elements, and micro/nano
183 particles is astonishing and it deserves far more research on the mechanisms
184 involved.

185
186 Given the commonality of high-dose toxic responses, the default assumption
187 should be baseline toxicity. It is researchers' responsibility to determine if their
188 high-dose results are something other than a non-specific, cytotoxic response.
189 For most toxicity results, and the fish ELS toxicity syndrome, it is important to
190 perform an initial check to determine if toxicity is a baseline toxic response or a
191 truly specific effect.

192
193 A chemical is likely to be a baseline toxicant if:

194

195 1. Whole-body concentrations fall within range of those defining baseline toxicity
196 as highlighted above (mortality > 0.5 mM and sublethal effects > 0.05 mM
197 (lower 95% confidence interval)). These concentrations can be measured or
198 modeled with aqueous exposure concentrations and partition coefficients.

199 2. Aqueous toxicity metrics for organic compounds fall on or nearby the QSAR
200 regression line defined in Figure 1 and the toxic ratio (TR_{LC50} , eq. 1) is < 10.

$$201 \quad TR_{LC50} = \frac{LC50 \text{ baseline}}{LC50 \text{ observed}} \quad (1)$$

202 3. An aqueous sublethal response concentration is within an order of magnitude
203 of the mortality response. This can be determined with an acute to chronic ratio
204 that is < 10 (Scholz et al. 2018). In most cases, the LOECs for sublethal ELS
205 toxicity syndrome responses are close to the LC50 (Figure 1).

206 4. Because the ELS toxicity syndrome can occur by specific toxicity, the aqueous
207 sublethal response should be compared to the baseline LC50 (i.e., if the ratio of
208 $LC50_{\text{baseline}}$ to EC_{observed} (ELS toxicity syndrome) is <10 , then it is baseline
209 toxicity).

210 5. Reactive oxygen species (ROS) and apoptosis markers are elevated, in
211 addition to many of the gene markers associated with sarcoplasmic/endoplasmic
212 reticulum (ER) membrane stress. If the response occurs close to predicted
213 baseline toxicity, it is likely a result of the cytotoxicity burst and not a specific
214 effect.

215

216 A chemical is likely to be specific-acting if it does not fulfill 1-4, and if:

217 6. There is a direct demonstration of receptor interaction at environmentally
218 relevant concentrations, such as AhR activation. Additional support for receptor
219 interaction could come from competitive or non-competitive receptor interaction
220 results. Further support for potential receptor interaction may be found in the
221 ECOdrug database (Verbruggen et al. 2018), Drugbank (Wishart et al. 2018) or
222 the T3DB (Wishart et al. 2015)

223 7. An adverse outcome pathway (AOP) or target-specific sequence of events has
224 been defined for the observed response at the reported non-baseline level
225 exposure concentrations. An important component of an AOP is the identification
226 of a receptor as the molecular initiating event.

227 8. Only one of ROS and ER stress markers are elevated below predicted baseline
228 toxicity concentrations, which indicates a specific stress response. If both are
229 elevated close to predicted baseline toxicity concentrations, the ROS and ER
230 stress are likely a consequence of the cytotoxicity burst (Fay et al. 2018; Escher
231 et al. 2020b) and not a specific effect.

232

233 In most cases, high-dose responses occur at concentrations that are not
234 environmentally relevant. For organic compounds, aqueous levels in the mg/L or

235 high $\mu\text{g/L}$ range in the environment are extremely rare. The baseline response is
236 such a severe and universal response for so many compounds, any species-
237 specific response occurring at baseline levels for these high environmental
238 concentrations would have deleterious ramifications for all species. The
239 observation of severe responses at such high doses does not necessarily
240 translate to an adverse response at lower doses, especially those that are non-
241 specific, cytotoxic insults. However, baseline toxicants are concentration
242 additive in mixtures. Therefore, baseline toxicity might play a role for complex
243 environmental mixtures even if all components of the mixture are not
244 individually toxic at their concentrations in the mixture.

245

246 An important application for the default assumption of baseline toxicity concerns
247 the determination of toxicity metrics for complex mixtures. In some cases, the
248 adverse response is based on a small percentage of compounds comprising the
249 total concentration. One example of this is the myriad studies on oil toxicity
250 claiming that PAHs (specifically tricyclic PAHs) are the main toxic component
251 causing the fish ELS syndrome, when the default assumption should be baseline
252 toxicity. PAHs are a minor component of oil and are nowhere near as toxic in
253 isolation as is concluded when exposing fish to the total water-soluble fraction
254 without invoking unsubstantiated synergism. In those studies, the potential
255 toxicity of thousands of compounds is ignored in favor of PAHs that can be
256 quantified with well-developed protocols and analytical standards, as opposed to
257 myriad known, predicted, and suspected compounds in the WSF that cannot be
258 easily quantified (Meador and Nahrgang 2019). As the funnel hypothesis states,
259 the more components a mixture has, the more likely toxicity of the mixture is
260 due to additive effects resulting from the baseline toxic component (Warne and
261 Hawker 1995). A recent study by Harsha et al. (2024) demonstrated the ELS
262 toxicity syndrome for a crude oil WSF without detectable PAHs. This study
263 supports the hypothesis that PAHs are not the primary cause of the fish ELS
264 toxicity syndrome due to crude oil exposure and that baseline toxicity is the

265 more reasonable conclusion. No one component or class of compounds in crude
266 oil has been shown to cause the ELS toxicity syndrome in isolation at the
267 concentrations at which they occur in the mixture. Importantly, there are
268 exceptions such as that from Tian et al. (2021) who identified the primary
269 casual toxicant (6PPD-quinone) in stormwater, which explained toxicity of the
270 mixture (stormwater) and left no doubt that the response was due to one single
271 specific-acting compound that matched the toxic response for the mixture and
272 causative agent. Such cases are spectacular, but rare and in the big scheme of
273 things the unspectacular process of concentration addition of underlying
274 baseline toxicity can overall do more harm than a few synergistically acting toxic
275 mixture components.

276

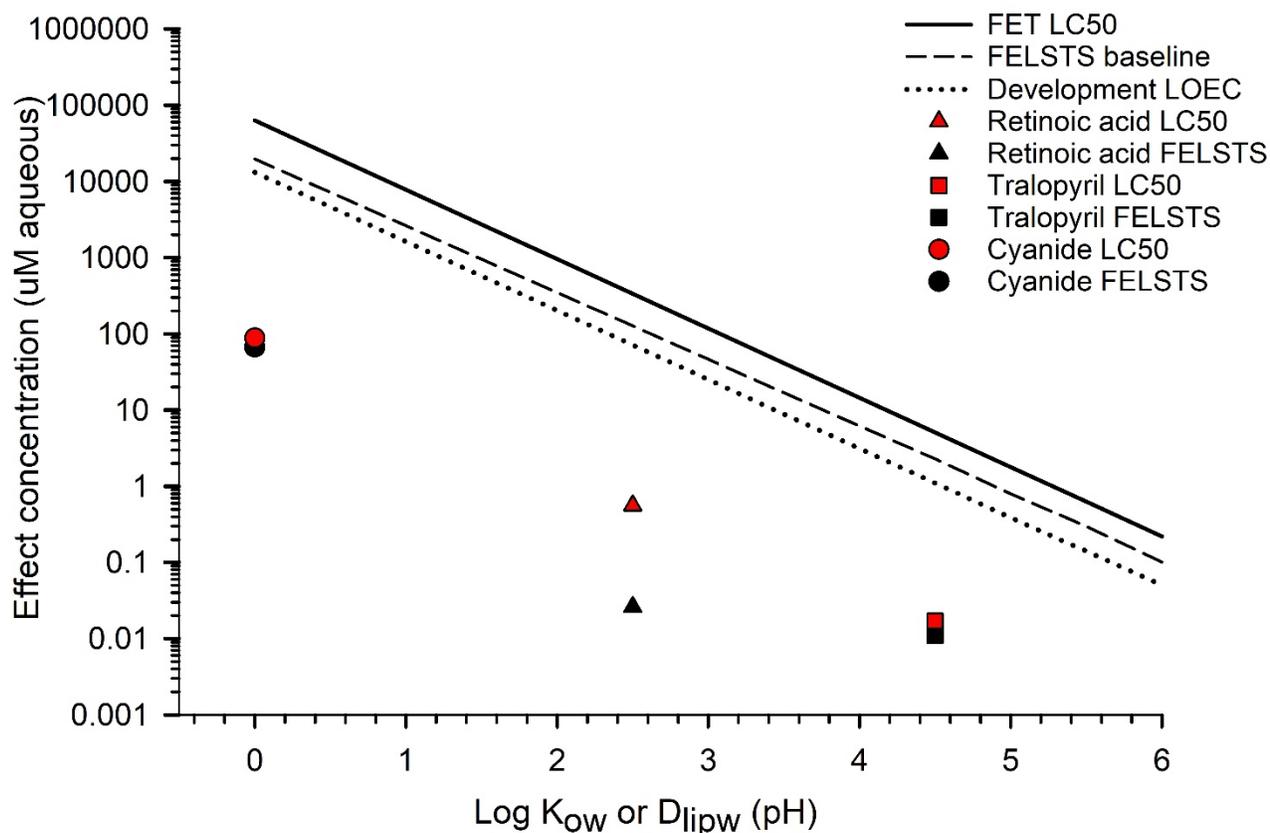
277 In aquatic toxicity studies, high-dose exposure results are useful for determining
278 mortality or near lethal toxicity but not for identifying low-dose specific toxicity,
279 which is more relevant for environmental risk assessment. There is little reason
280 to assume that biochemical targets activated by high-dose exposure will be
281 relevant when examined in terms of environmentally relevant exposure levels.
282 Also, there is no guarantee that effects from short-term, high-dose exposures
283 will be biologically significant for long-term, low dose exposures. Effects due to
284 receptor interaction at high doses may be very weak, or non-existent, at lower
285 doses and apical effects such as impaired growth, reproduction, behavior, and
286 others will likely be absent. This is also critical for baseline toxicity because
287 membrane disruption and biological effects ~~resulting from high-dose exposure~~
288 will not occur at lower doses. It must be noted that baseline toxicity for very
289 hydrophobic chemicals occurs at low external aqueous concentrations because
290 the bioaccumulation is high for hydrophobic chemicals, but the baseline toxicity
291 occurs always the highest concentrations for any given compound, any specific
292 effect occurs at lower concentrations. Additionally, in some cases environmental
293 concentrations may be very high due to intentional or unintentional releases of
294 contaminants, therefore baseline toxicity for these situations will be informative.

295 Adequate protection of aquatic populations will be accomplished with an
296 accurate evaluation for all chemicals in the environment capable of inducing
297 receptor-based sublethal effects that over the long term may impact population
298 viability. The goal should be to identify the lowest observed effect
299 concentrations for specific effects, not the highest dose that affects all bodily
300 functions regardless of the mechanism of effect. In simple terms, baseline
301 toxicity is not a useful or relevant response for environmental protection under
302 most situations.

303

304 **Conclusions**

305 It is important to realize that it is common for organic compounds to cause non-
306 specific baseline toxicity at high doses and specific, receptor-mediated toxicity
307 at low doses leading to different adverse outcomes (Meador 2021). In many
308 cases, chemicals causing abnormal morphological effects at high exposure
309 concentrations are mistakenly classified as teratogens when in fact the response
310 is nothing more than baseline toxicity. As toxicologists, we should focus our
311 research efforts on those specific-acting compounds that affect endocrine
312 systems, behavior, immune dysfunction, morphological abnormalities and other
313 important responses occurring at low and environmentally realistic
314 concentrations with the potential to affect organisms and their probability of
315 completing a successful life cycle.



316
 317 Figure 1: Aqueous lethal and sublethal effect concentrations (μM) against
 318 hydrophobicity expressed by the octanol-water partition coefficient $\log K_{ow}$ or the lipid
 319 membrane-water distribution ratio $\log D_{lipw}$. FET refers to the baseline toxicity QSAR for
 320 the 96 h LC50 fish embryo mortality (FET) using data from Kluver et al. (2016) ($n=99$,
 321 $r^2=0.75$). FELSTS baseline refers to the baseline toxicity QSAR for the fish early life-
 322 stage toxicity syndrome related to development, edema, and reduced heart rate
 323 endpoints after exposure of 4 – 34 days (Scholz et al. 2018 and Meador 2021) ($n=36$,
 324 $r^2=0.82$). Development LOEC refers to the QSAR for developmental abnormalities at
 325 days 28 - 34 for ELS fish ($n=14$, $r^2=0.83$) (Scholz et al. 2018) to show slightly higher
 326 toxicity for longer term results. All QSARs for $TR_{LC50} < 10$. Examples for the FELSTS for
 327 specific action ($TR_{LC50} > 10$) are shown for retinoic acid (Δ), cyanide (\bullet), and tralopyril
 328 (\square). LC50 (red) and sublethal endpoints (black). $TR_{LC50} = \frac{LC50 \text{ baseline}}{LC50 \text{ observed}}$

Citations

- Chakraborty A, Adhikary S, Bhattacharya S, Dutta S, Chatterjee S, Banerjee D, Ganguly A, and Rajak P. 2023. Pharmaceuticals and personal care products as emerging environmental contaminants: prevalence, toxicity, and remedial approaches. *Chem. Health Safe.* 30:362-388. DOI: 10.1021/acs.chas.3c00071
- Chen J, Liang Q, Zheng Y, Lei Y, Gan X, Mei H, Bai C, Wang H, Ju J, Dong Q, Song Y. 2024. Polystyrene nanoplastics induced size-dependent developmental and neurobehavioral toxicities in embryonic and juvenile zebrafish. *Aquat. Toxicol.* 267:106842. doi: 10.1016/j.aquatox.2024.106842.
- D'Amico LD, Sen WL, Yang Y, and Suter W. 2012. Assessment of drug-induced cardiotoxicity in zebrafish. In: McGrath, P. (Ed.), *Zebrafish: Methods for Assessing Drug Safety and Toxicity*. John Wiley & Sons, Inc., pp. 45–54 (Chapter 4).
- Ducharme NA, Peterson LE, Benfenati E, Reif D, McCollum CW, Gustafsson J-A, Bondesson M. 2013. Meta-analysis of toxicity and teratogenicity of 133 chemicals from zebrafish developmental toxicity studies. *Repro. Toxicol.* 41:98-108. doi: 10.1016/j.reprotox.2013.06.070
- Escher BI and Hermens JLM. 2004. Internal exposure: linking bioavailability to effects. *Environ. Sci. Technol.* 455A-462A. doi: 10.1021/es0406740
- Escher BI, Schwarzenbach RP. 2002. Mechanistic studies on baseline toxicity and uncoupling of organic compounds as a basis for modeling effective membrane concentrations in aquatic organisms. *Aquat. Sci.* 64:20-35. DOI: 10.1007/s00027-002-8052-2.
- Escher B, Abagyan R, M E, Klüver N, Redman A, Zarfl C, Parkerton T. 2020a. Recommendations for improving methods and models for aquatic hazard assessment of ionizable organic chemicals. *Environ. Toxicol. Chem.* 39:269-286. DOI: 10.1002/etc.4602.
- Escher BI, Henneberger L, König M, Schlichting R, Fischer FC. Cytotoxicity Burst? Differentiating Specific from Nonspecific Effects in Tox21 *in Vitro* Reporter Gene Assays. 2020b. *Environ. Health Perspect.* 128:77007. doi: 10.1289/EHP6664.
- Escher, B.I., Eggen, R.I.L., Schreiber, U., Schreiber, Z., Vye, E., Wisner, B., Schwarzenbach, R.P. 2002. Baseline toxicity (narcosis) of organic chemicals determined by in vitro membrane potential measurements in energy-transducing membranes. *Environ. Sci. Technol.* 36:1971–1979. doi:10.1021/es015844c
- Fay KA, Villeneuve DL, Swintek J, Edwards SW, Nelms MD, Blackwell BR, Ankley GT. 2018. Differentiating pathway-specific from nonspecific effects

- in high-throughput toxicity data: a foundation for prioritizing adverse outcome pathway development. *Tox. Sci.* 163:500-515. DOI: 10.1093/toxsci/kfy049.
- González-Penagos CE, Zamora-Briseño JA, Améndola-Pimenta M, Cruz-Quintana Y, Santana-Piñeros AM, Torres-García JR, Cañizares-Martínez MA, Pérez-Vega JA, Peñuela-Mendoza AC, Rodríguez-Canul R. *Sargassum* spp. 2024. Ethanol extract elicits toxic responses and malformations in zebrafish (*Danio rerio*) embryos. *Environ. Toxicol. Chem.* doi: 10.1002/etc.5840.
- Harsha ML, Salas-Ortiz Y, Cypher AD, Osborn E, Turcios Valle E, Gregg JL, Hershberger PK, Kurerov Y, King S, Goranov AI, Hatcher PG, Konefal A, Cox TE, Greer JB, Meador JP, Tarr MA, Tomco PL, Podgorski DC. Toxicity of crude oil-derived polar unresolved complex mixtures to Pacific herring embryos: Insights beyond PAHs. *Sci. Tot. Environ.* In press.
- Incardona JP, Collier TK, Scholz NL. 2004. Defects in cardiac function precede morphological abnormalities in fish embryos exposed to polycyclic aromatic hydrocarbons. *Toxicol. Appl. Pharmacol.* 196, 191–205. <https://doi.org/10.1016/j.taap.2003.11.026>.
- Jarque S, Rubio-Brotos M, Ibarra J, Ordoñez V, Dyballa S, Miñana R, Terriente J. 2020. Morphometric analysis of developing zebrafish embryos allows predicting teratogenicity modes of action in higher vertebrates. *Reprod. Toxicol.* 96:337-348. doi.org/10.1016/j.reprotox.2020.08.004
- Klüver N, Vogts C, Altenburger R, Escher BI, Scholz S. 2016. Development of a general baseline toxicity QSAR model for the fish embryo acute toxicity test. *Chemosphere*164:164-173. <https://doi.org/10.1016/j.chemosphere.2016.08.079>.
- Klüver N, Bittermann K, Escher BI. 2019. QSAR for baseline toxicity and classification of specific modes of action of ionizable chemicals in the zebrafish embryo toxicity test. *Aquat. Toxicol.* 207:110-119. DOI: 10.1016/j.aquatox.2018.12.003.
- Knöbel M, Busser FJM, Rico-Rico À, Kramer NI, Hermens JLM, Hafner C, Tanneberger K, Schirmer K, Scholz S. 2012. Predicting adult fish acute lethality with the zebrafish embryo: relevance of test duration, endpoints, compound properties, and exposure concentration analysis. *Environ. Sci. Technol.* 46: 9690–9700. <https://doi.org/10.1021/es301729q>.
- Könemann H. 1981. Quantitative Structure-Activity Relationships in fish toxicity studies. Part 1: relationship for 50 industrial pollutants. *Toxicology* 19: 209-221.
- Lee J, Braun G, Henneberger L, König M, Schlichting R, Scholz S, Escher BI. 2021. Critical membrane concentration and mass-balance model to identify

- baseline cytotoxicity of hydrophobic and ionizable organic chemicals in mammalian cell lines. *Chem. Res. Toxicol.* 34:2100-2109. doi: 10.1021/acs.chemrestox.1c00182.
- McCarty LS, Arnot JA, Mackay D. 2013. Evaluation of critical body residue data for acute narcosis in aquatic organisms. *Environ. Toxicol. Chem.* 32:2301–2314. doi: 10.1002/etc.2289
- McCarty LS, Mackay D. 1993. Enhancing ecotoxicological modeling and assessment: body residues and modes of toxic action. *Environ. Sci. Technol.* 27:1719-1728.
- Meador JP and Nahrgang J. 2019. Characterizing crude oil toxicity to early-life stage fish based on a complex mixture: Are we making unsupported assumptions? *Environ. Sci. Technol.* 53:11080-11092. DOI: 10.1021/acs.est.9b02889
- Meador JP. The fish early-life stage sublethal toxicity syndrome - A high-dose baseline toxicity response. 2021. *Environ. Pollut.* 291:118201. doi: 10.1016/j.envpol.2021.118201.
- Meador JP, Adams WJ, Escher BI, McCarty LS, McElroy AE, Sappington KG. 2011. The tissue residue approach for toxicity assessment: Findings and critical reviews from a society of environmental toxicology and chemistry pellston workshop. *Integr. Environ. Assess. Manag.* 7:2–6. doi: 10.1002/ieam.133
- Meador JP, McCarty LS, Escher BI, Adams WJ. 2008. The tissue-residue approach for toxicity assessment: concepts, issues, application, and recommendations. *J. Environ. Monit.* 10:1486-1498. <http://dx.doi.org/10.1039/b814041n>
- Rana SVS. Endoplasmic reticulum stress induced by toxic elements-a review of recent developments. 2020. *Biol. Trace Elem. Res.* 196:10-19. doi: 10.1007/s12011-019-01903-3.
- Scholz S, Schreiber R, Armitage J, Mayer P, Escher BI, Lidzba A, Léonard M, Altenburger R. 2018. Meta-analysis of fish early life stage tests - association of toxic ratios and acute-to-chronic ratios with modes of action. *Environ. Toxicol. Chem.* 37:955-969. doi: 10.1002/etc.4090.
- Seok SH, Baek MW, Lee HY, Kim DJ, Na YR, Noh KJ, Park SH, Lee HK, Lee BH, Park JH. 2008. In vivo alternative testing with zebrafish in ecotoxicology. *J Vet Sci.* 9:351-7. doi: 10.4142/jvs.2008.9.4.351.
- Simmons SO, Fan CY, Ramabhadran R. 2009. Cellular stress response pathway system as a sentinel ensemble in toxicological screening. *Tox. Sci.* 111:202-225. DOI: 10.1093/toxsci/kfp140.

- Shankar P, Villeneuve DL. AOP Report: Aryl Hydrocarbon Receptor Activation Leads to Early-Life Stage Mortality via Sox9 Repression-Induced Craniofacial and Cardiac Malformations. 2023. *Environ. Toxicol. Chem.* 42:2063-2077. doi: 10.1002/etc.5699.
- Tian Z, Zhao H, Peter KT, Gonzalez M, Wetzel J, Wu C, Hu X, Prat J, Mudrock E, Hettlinger R, Cortina AE, Biswas RG, Kock FVC, Soong R, Jenne A, Du B, Hou F, He H, Lundeen R, Gilbreath A, Sutton R, Scholz NL, Davis JW, Dodd MC, Simpson A, McIntyre JK, Kolodziej EP. 2021. A ubiquitous tire rubber-derived chemical induces acute mortality in coho salmon. *Science* 371(6525):185-189. doi: 10.1126/science.abd6951.
- Truong L, Harper SL, Tanguay RL. 2011. Evaluation of embryotoxicity using the zebrafish model. *Meth. Mol. Biol.* 691:271-279. doi: 10.1007/978-1-60761-849-2_16.
- van Wezel AP, Opperhuizen A. 1995. Narcosis due to environmental pollutants in aquatic organisms: residue-based toxicity, mechanisms, and membrane burdens. *Crit. Rev. Toxicol.* 25:255-79. doi: 10.3109/10408449509089890.
- Verbruggen B, Gunnarsson L, Kristiansson E, Österlund T, Owen SF, Snape JR, Tyler CR. 2018. ECOdrug: a database connecting drugs and conservation of their targets across species. *Nucleic Acids Res.* 46:D930–D936. <https://doi.org/10.1093/nar/gkx1024> Available from: <https://ecodrug.org/>
- Warne MSJ, Hawker DW. 1995. The number of components in a mixture determines whether synergistic and antagonistic or additive toxicity predominate - the funnel hypothesis. *Ecotox. Environ. Safe.* 31:23-28.
- Wilhelmi P, Haake V, Zickgraf FM, Giri V, Ternes P, Driemert P, Nöth J, Scholz S, Barenys M, Flick B, Birk B, Kamp H, Landsiedel R, Funk-Weyer D. 2024. Molecular signatures of angiogenesis inhibitors: a single-embryo untargeted metabolomics approach in zebrafish. *Arch. Toxicol.* 98:943-956. doi: 10.1007/s00204-023-03655-5.
- Wishart D, Arndt D, Pon A, Sajed T, Guo AC, Djoumbou Y, Knox C, Wilson M, Liang Y, Grant J, Liu Y, Goldansaz SA, Rappaport SM. 2015. T3DB: the toxic exposome database. *Nucleic Acids Res.* 43:D928-34. doi: 10.1093/nar/gku1004.
- Wishart DS, Feunang YD, Guo AC, Lo EJ, Marcu A, Grant JR, Sajed T, Johnson D, Li C, Sayeeda Z, Assempour N, Iynkkaran I, Liu Y, Maciejewski A, Gale N, Wilson A, Chin L, Cummings R, Le D, Pon A, Knox C, Wilson M. 2018. DrugBank 5.0: a major update to the DrugBank database for 2018. *Nucleic Acids Res.* 46:D1074-D1082. doi: 10.1093/nar/gkx1037.

Zhu D, Li TT, Zheng SS, Yan LC, Wang Y, Fan LY, Li C, Zhao YH. 2018.
Comparison of modes of action between fish and zebrafish embryo toxicity
for baseline, less inert, reactive and specifically-acting compounds.
Chemosphere 213:414-422. doi: 10.1016/j.chemosphere.2018.09.072.