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Diet composition and trophic niche differentiation of *Neogobius melanostomus* along an invasion gradient in a large lowland river

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2 invasion gradient in a large lowland river

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33

34 **Abstract**

35 The Ponto-Caspian goby *Neogobius melanostomus* (Pallas 1814) is a widespread non-native
36 fish species in European rivers. It occurs in high abundance and can potentially alter the food
37 webs of invaded water bodies profoundly. However, the invasion process usually goes
38 unnoticed, and changes in food webs during the early stages of invasions are rarely described.
39 Here, we studied populations of *N. melanostomus* along an invasion gradient in the Elbe River
40 (Germany) and tested the effects of time since invasion on the diets and the associated trophic
41 dynamics in two dominant habitats, i.e., rip rap and sand. Results showed that the abundance
42 and biomass of *N. melanostomus* was lowest at the most recently invaded sites. The trophic
43 niche space quantified by stable isotopes ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$) showed that populations at earlier
44 invaded sites exhibited a wider trophic niche than those at the most recently invaded sites.
45 Diptera and Crustacea were the most abundant taxa in *N. melanostomus* guts at both habitat
46 structures. At rip rap habitats, *N. melanostomus* showed a significant increasing preference for
47 Crustacea and a decreasing one for Gastropoda along the invasion gradient. At sand habitats no
48 significant relationships between the invasion gradient and electivity index for *N.*
49 *melanostomus* were detected. We demonstrated that the invasion stage and habitat in which *N.*
50 *melanostomus* occurs affect the feeding preference of this invasive fish and that these factors
51 are therefore important for tracing changes in the macroinvertebrate communities of affected
52 river systems.

53
54 **KEY WORDS:** Invasive fish, Round goby, Invasion front, Elbe River, feeding ecology, Stable
55 isotopes

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65 **Introduction**

66 The Ponto-Caspian round goby *Neogobius melanostomus* (Pallas 1814) is a benthic-feeding
67 fish species with its origins in the Ponto-Caspian region. Since the 1990s, it has successfully
68 spread and invaded rivers throughout Europe and North America (Borcherding et al., 2013;
69 Hempel & Thiel, 2013; Kalchhauser et al., 2013).

70 The round goby has a suite of typical invader traits such as a high reproduction rate, tolerance
71 of a wide range of environmental conditions, low food selectivity as well as species-typical
72 behaviours like dispersal tendencies or aggressiveness (Church et al., 2017; Foley et al., 2017;
73 Groen et al., 2012; Pothoven, 2018). These traits result in the ability to dominate even with a
74 limited food supply and lead to an impact on the prey population (Krakowiak & Pennuto, 2008;
75 Lederer et al., 2008), local food webs (Brush et al., 2012) and could ultimately even lead to the
76 local extirpation of native benthic species and habitat destruction (Cooper et al., 2009; Pettitt-
77 Wade et al., 2015). Therefore, this introduced fish species may significantly alter food webs
78 and trophic structures by changing community composition and major nutrient fluxes (Herlevi
79 et al., 2018). *N. melanostomus* prefers artificial shoreline structures commonly found in
80 navigable rivers, i.e., rip rap structures (Borcherding et al., 2011) and feeds on a wide variety
81 of macroinvertebrates. Its dominant prey throughout rivers in Europe and its home range are
82 Crustacea (mainly amphipods) and Diptera (mainly chironomids) (Błońska et al., 2015;
83 Borcherding et al., 2013; Raby et al., 2010).

84 Studies conducted in the Baltic Sea or the North American Great Lakes area have shown that
85 round goby affects macroinvertebrate communities (Kipp & Ricciardi, 2012; Lederer et al.,
86 2008) by decreasing their densities and changing taxonomic composition (Henseler et al., 2021;
87 Krakowiak & Pennuto, 2008). The consequence is that *N. melanostomus* may have a crucial
88 influence on the taxonomic composition of macroinvertebrate communities (Henseler et al.,
89 2021) and may cause shifts in their assemblage structure with possible impacts such as the
90 exploitation of prey resources and increased competition for native fish (Mikl et al., 2017). This
91 is mainly due to the ability of *N. melanostomus* to exhibit an opportunistic feeding behaviour
92 and an adaptive predation strategy that facilitates the successive establishment of new water
93 bodies and contributes to its widespread distribution (Didenko et al., 2020; Perello et al., 2015).
94 The prey selection of *N. melanostomus* depends on several factors, e.g., **(1)** the habitat structure
95 in which they occur and, therefore, the availability of habitat-specific prey (Borcherding et al.,
96 2013), **(2)** seasonal dynamics of macroinvertebrates abundance and thus varying availability
97 (Didenko et al., 2020), **(3)** inter and intra-specific interactions, e. g. strong competition for food
98 and high dietary overlaps (Borcherding et al., 2019), **(4)** mouth gaps in smaller fish may

99 influence the feeding ability (Borcherding et al., 2013; Krebs & Turingan, 2003; Raby et al.,
100 2010), (5) diel patterns as feeding can vary during day and night. The main feeding time of *N.*
101 *melanostomus* was determined directly after sunrise whereby, especially juveniles may migrate
102 into gravel and sand habitats during the night (Borcherding et al., 2013), or (6) body size, as *N.*
103 *melanostomus* revealed an ontogenetic diet shift with a switch from preying upon insects and
104 crustaceans to mainly molluscs with at a total length of 10 cm (Brandner et al., 2013b).

105 The resulting alterations in food webs (Brandner et al., 2013a; Brush et al., 2012) are expected
106 to change with time since the invasion began (Herlevi et al., 2018). As invasion stages are
107 correlated to the numerical abundance of *N. melanostomus* (Brandner et al., 2013b), knowledge
108 of their ecology at all stages of the invasion is crucial to estimate associated ecosystem impacts
109 and to quantify the potential impact on riverine food webs (Brandner et al., 2013b). However,
110 determining the current invasion stage in invaded rivers is notoriously difficult, as the required
111 field monitoring is often lacking or carried out after the invasion (Brandner et al., 2013b).
112 Consequently, the invasion process usually goes unnoticed, and changes in food webs during
113 the early stages of invasions are rarely described.

114 This knowledge about invasion dynamics and the factors that facilitate the adaption to the new
115 ecosystem is necessary to understand the interactions between the invasive species and their
116 prey and to help elucidate any associated impacts on the ecosystem (Taraborelli et al., 2010).
117 In particular, an early-stage characterization of the 'invaders' impact can help to develop
118 sufficient ecosystem conservation measures and preventative management actions (Henseler et
119 al., 2020; Kornis et al., 2013).

120 In the Elbe River, *N. melanostomus* was detected for the first time in 2008 in the tidal reach
121 (Hempel & Thiel, 2013) in Germany and 2015 in the upper Elbe River in the Czech Republic
122 (Buřič et al., 2015; Roche et al., 2015). A previous study successfully traced the dispersal
123 patterns and population structure of *N. melanostomus* in the Elbe River (Nogueira Tavares et
124 al., 2020). There, we could demonstrate that *N. melanostomus* had not invaded the entire river
125 course, as sites in the middle course showed a very low *N. melanostomus* abundance and had
126 low environmental DNA (eDNA) concentrations (Nogueira Tavares et al., 2020). Based on
127 these findings, the ongoing invasion process in the Elbe River was described as continuously
128 increasing from upstream to downstream. This study took advantage of the unique opportunity
129 to follow the invasion process and studied differences in diet composition and corresponding
130 trophic dynamics along the invasion gradient. We hypothesized that macroinvertebrate
131 abundance are influenced negatively by *N. melanostomus* abundance along the invasion
132 gradient. We expected earlier invaded sites to have higher numbers of *N. melanostomus*

133 individuals and therefore, lower numbers of macroinvertebrates than the recently invaded sites.
 134 Further, we expected gut contents of *N. melanostomus* caught in sand habitats to indicate
 135 different prey items than those caught in rip rap habitats.

136 Moreover, we hypothesised that more established populations at the longer invaded sites would
 137 face stronger intraspecific competition, leading to a broader trophic niche.

138

139 **Materials and methods**

140

141 **Study area and invasion gradient**

142 The study area was located at the Elbe River, from river km (rkm) 66 downstream to rkm 268,
 143 covering a free-flowing river section in Germany. We took samples from six sites placed
 144 approximately every 60 rkm along the river course. Based on a previous study (Nogueira
 145 Tavares et al., 2020), the river course between rkm 66 and 217.4 can be characterised as being
 146 invaded earlier (i.e., before 2018). In contrast, rkm 259.1 and 268 can be characterised as the
 147 most recently invaded sites at sampling times (i.e., after 2019, Fig. 1).

148



149

150 **Fig. 1.** Invasion history of *N. melanostomus* in the Elbe River and its main tributaries (blue lines). The
 151 invasion gradient is marked with an arrow, indicating the invasion progress and the corresponding year
 152 of round goby detection.

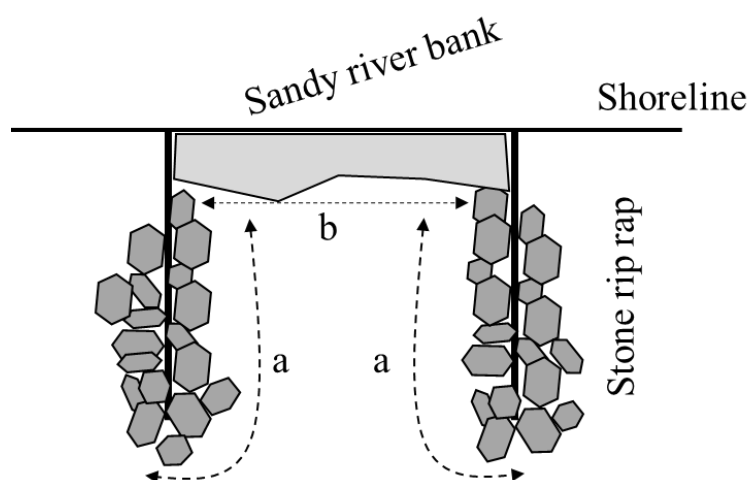
153

154 **Sampling and sample analysis**

155 **Fish**

156 Fish sampling was carried out in May 2019 by point abundance electrofishing (electrofishing
 157 generator DEKA 7000, DEKA Gerätebau Marsberg, Germany) in the morning during daylight
 158 hours. Sampling was carried out along 400 m near-shore stretches consisting of two groynes
 159 and two habitat structures: sand and rip rap (Fig. 2). The rocks of the rip rap varied from 5 to
 160 50 cm in diameter. Similarly, structured sampling sites were chosen to minimize habitat effects
 161 on *N. melanostomus* consumption patterns. Caught fishes were identified to the species level,
 162 counted, and total body length was measured to the nearest 1 mm. Gobies were additionally
 163 weighed to the nearest 0.01 g and frozen at -20°C. Up to 30 individuals per rip rap and sandy
 164 habitat from each site were randomly subsampled for gut content and stable isotope analyses in
 165 the laboratory.

166



167

168 **Fig. 2** Scheme of analysed groyne structure, divided into a rip rap (a) and a sandy river bank (b).

169

170 **Macroinvertebrates**

171 To characterise potential food resources of *N. melanostomus*, macroinvertebrates were
 172 collected along with the fish sampling from rip rap and sandy groyne fields (Fig. 2) with four
 173 replicates each. Stones were sampled by brushing off macroinvertebrates. To collect
 174 macroinvertebrates from sandy groyne fields, kick sampling was applied. Therefore handheld
 175 kick nets (0.25 mm mesh size) were placed vertically to the water bottom. The sampled area
 176 was calculated by measuring each boulder's length, height, and width under the assumption of

177 a cuboid shape (sampled area 0.063 m²) following (Brabender et al., 2016). Samples were
178 preserved in 70% ethanol. Macroinvertebrates were sorted, counted and identified to the species
179 or genus level, except for Oligochaeta (order level) and Diptera (family level), excluding
180 Chironomidae (species or genus level).

181

182 **Gut content analysis**

183 To determine the feeding activity and prey selection of *N. melanostomus*, the entire
184 gastrointestinal tract was removed and preserved in 70% ethanol. Each gut was weighed both
185 full and empty to the nearest 0.1 mg. Gut contents were preserved in 70% ethanol and later
186 sorted under a dissecting microscope. The prey items were identified to the lowest possible
187 taxonomic level and counted. Body lengths of all individuals or a random subsample (if more
188 than ten individuals of one taxon were counted) were measured to the nearest 0.1 mm using an
189 object micrometre. Shell length was measured for Bivalvia, shell height in Gastropoda, head
190 capsule width in Trichoptera, carapax width in *Eriocheir sinensis* and length and width in
191 Oligochaeta. The dry body mass of each prey individual was calculated using length–dry mass
192 regressions (Baumgärtner & Rothhaupt, 2003; Benke et al., 1999; Burgherr & Meyer, 1997;
193 Hellmann et al., 2015; Mährlein et al., 2016; Meyer, 1989; Normant et al., 2000). As shells are
194 not digested but expelled from the gut, shell-free dry mass was calculated for molluscs.

195 Average dry masses were calculated for zooplankton prey, mostly Cladocera, cyclopoid
196 copepods and copepodites. For Bryozoa, no length–dry mass relationship was available, and an
197 average individual dry mass of 0.001 mg was assumed based on the biovolume of a typical
198 zooid. The dry mass for all ingested individuals was calculated and summed up, and dry mass
199 proportions in the diet were calculated for each fish. The dry mass proportions of the non-
200 countable food sources detritus and filamentous algae were estimated visually and included in
201 the diet calculation.

202

203 **Stable isotope analyses**

204 We conducted stable isotope analyses to determine the trophic niche space of *N. melanostomus*.
205 Samples for stable isotopes were obtained by dissecting a small part of the dorsal muscle. The
206 tissues were oven-dried for 48 hours at 40 °C and grounded with an oscillating mill (MM400,
207 Retsch, Germany) to a fine homogeneous powder. Samples were weighed into tin cups (5 x 9
208 mm, IVA Analysentechnik GmbH & Co. KG, Germany). Carbon and nitrogen elemental
209 concentration and $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ ratios were determined on a Thermo-Finnigan Flash 2000
210 elemental analyser connected to a Delta V Advantage mass spectrometer (Thermo Fisher

211 Scientific, Waltham, Massachusetts). Stable isotope data were expressed as the relative
 212 difference between the ratios of samples and international standards (PeeDee Belemnite for
 213 $\delta^{13}\text{C}$, atmospheric N for $\delta^{15}\text{N}$)

214 $\delta (\text{‰}) = (\text{R}_{\text{sample}} / \text{R}_{\text{standard}}) * 1000$, and $\text{R} = {}^{13}\text{C}/{}^{12}\text{C}$ or ${}^{15}\text{N}/{}^{14}\text{N}$, respectively.

215

216 **Statistical analysis**

217 The change of round goby abundance and biomass along the invasion gradient was tested with
 218 linear models of abundance and biomass and river km in Sigma Plot (v 13, Systat Software
 219 GmbH, Düsseldorf, Germany). We conducted separate models for the sand and rip rap habitats
 220 as differences between both habitats have been described previously (Borcherding et al., 2013).
 221 We visualised differences in prey communities along the invasion gradient by conducting non-
 222 metric multidimensional scaling on 4th root transformed macroinvertebrate abundances (Primer
 223 v7, PRIMER-E Ltd., Plymouth, UK). The analysis was supplemented by linear models of total
 224 macroinvertebrate abundance, taxa richness and abundance of macroinvertebrate major groups
 225 against river km. We also tested if macroinvertebrate abundance was related to goby abundance
 226 at rip rap and sand habitats by conducting Pearson correlation analysis in Sigma Plot.

227

228 To show the role of prey availability and goby dietary preferences, we calculated the gut
 229 fullness index (GFI, in %) to describe the feeding activity and estimated ingested food
 230 (Borcherding et al., 2013; Didenko et al., 2020; Všeticková et al., 2015):

231

$$232 \text{GFI} = (\text{gut content weight}) / (\text{fish weight}) * 100 \quad \text{eq. 1}$$

233

234 where gut content weight is the sum of the wet weights of individual prey items. We quantified
 235 the diversity of ingested prey items as the Shannon Wiener Index calculated from prey
 236 composition (online Resource, Figure A.1).

237 The relative abundance of macroinvertebrates in *N. melanostomus* guts was compared to the
 238 relative abundance sampled in the Elbe River by calculating the electivity-index (E)
 239 (Lechowicz, 1982; Stauffer et al., 2016; Wiegler et al., 2018):

240

$$241 \text{E} = (\text{r}_i - \text{p}_i) / (\text{r}_i + \text{p}_i) \quad \text{eq. 2}$$

242

243 where r_i is the relative abundance of prey item i (percentage of total gut contents) in the gut and
 244 p_i is the relative abundance of the same item found in the environment.

245 Values range from -1 to $+1$. Negative values indicate the rejection or inaccessibility of the prey,
 246 zero = random feeding, and positive values indicate an active selection (Piria et al., 2016).

247
 248 We used linear models to test for relationships of proportion of major macroinvertebrate groups
 249 in guts and electivity-index for macroinvertebrate major groups with river km in R (R Core
 250 Team, 2020). Prior to linear modelling, we tested for normality and constant variance.

251
 252 Trophic niche width was calculated as the standard ellipse area corrected for small sample sizes
 253 (SEAc) using the SIBER package (Jackson et al., 2011) in R. Sample sizes were highly uneven
 254 for *N. melanostomus* caught in sand habitats (Table 1). To preclude that larger ellipse areas are
 255 simply the result of larger sample sizes, we randomly selected with replacement five isotope
 256 values (lowest sample size at rkm 66) out of the initial sample and repeated this procedure 100
 257 times for rkm 124.3, 165 and 217. From the resulting 100 vectors for each rkm, we calculated
 258 the mean and 95% highest posterior density (HPD) intervals of SEAc for sand habitats. The
 259 HPD interval is the Bayesian counterpart to the frequentist's confidence intervals for posterior
 260 probability distributions (Marin & Robert, 2008).

261 We graphically analysed the relationship between trophic niche width and prey diversity but
 262 refrained from linear model analysis. Only four out of the six sites had sufficient goby
 263 abundances (Table 1) to calculate trophic niche width.

264 **Results**

265 **Abundance of *N. melanostomus* along the invasion gradient**

266
 267 A total of 318 individuals of *N. melanostomus* were caught at the six study sites, and round
 268 goby were overall more abundant in rip rap than in sand habitats (Table 1). The two most
 269 recently invaded sites (rkm 259.1 and 268) had the lowest number of individuals ($< 1\%$ of total
 270 catch) and biomass (Table 1). However, abundance and biomass did not significantly change
 271 along the invasion gradient in rip rap and sand habitats (all models $P > 0.05$).

272
 273 **Table 1** Overview of total abundance (ind./ 100m²) and biomass (g/ 100m²) of *N. melanostomus* and the
 274 number of individuals subjected to gut content and stable isotope analysis per sampling site and habitat
 275 structure.
 276
 277

	Elbe km					
	66	124.3	165	217.4	259.1	268

Rip rap	Total abundance	12.5	30.5	51.5	35	0.5	1
	Total biomass	42.6	244.25	334.8	165.65	0.75	2.5
	No. individuals	23	27	28	25	1	2
Sand	Total abundance	2.5	9.5	6	10	0	0
	Total biomass	2.65	28.2	25.4	50.95	0	0
	No. individuals	5	13	12	15	0	0

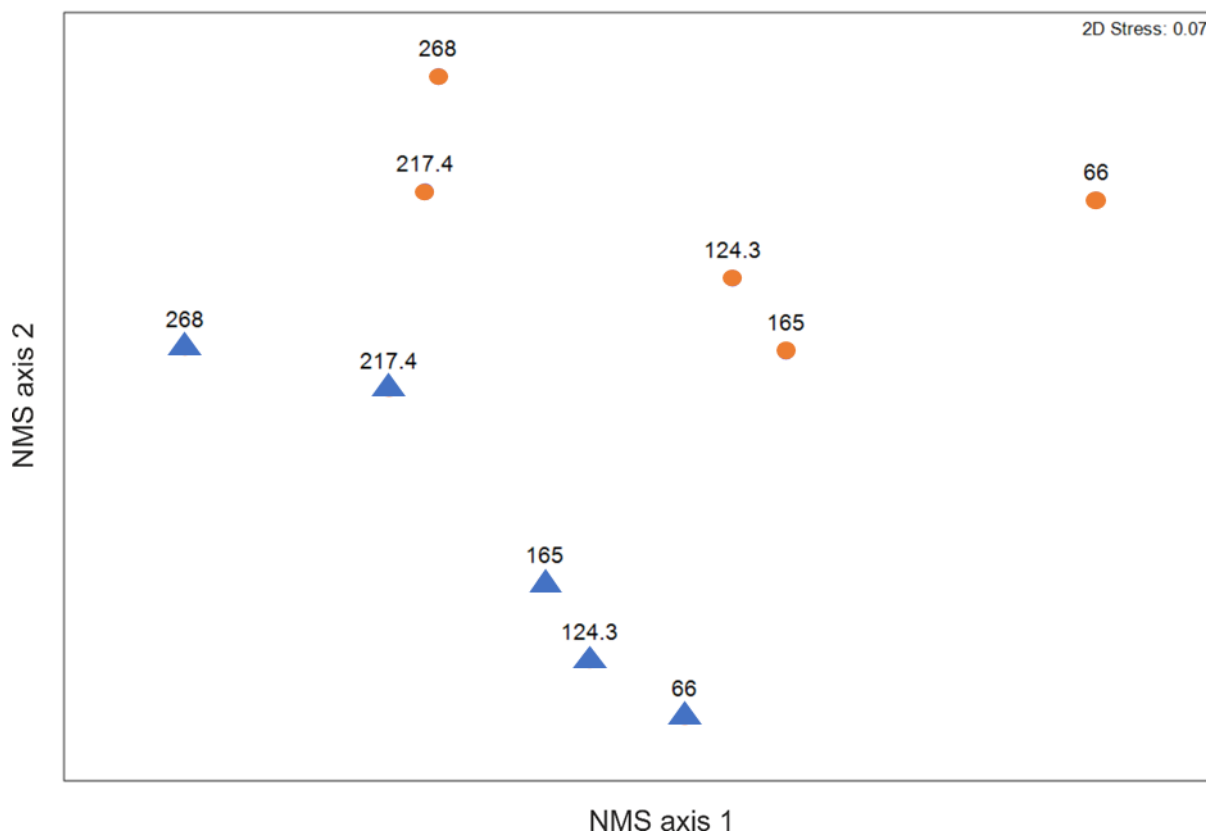
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279 **Macroinvertebrate composition in river and guts**

280 A total of 121,065 macroinvertebrates belonging to 10 major taxonomic groups were recorded
 281 (online Resource, Figure A.2). Macroinvertebrate community composition differed between
 282 habitat structures and along the invasion gradient. The downstream communities (at rkm 217.4
 283 and 268) were more dissimilar between the habitats than the upstream communities, as
 284 indicated by the larger distance in the NMS ordination (Fig. 3).

285 We found a significant relationship between rkm and abundances of Crustacea (mostly non-
 286 native *Dikerogammarus villosus*) (Linear model, $R^2 = 0.82$, $P = 0.035$) and Bivalvia (Linear
 287 model, $R^2 = 0.79$, $P = 0.018$) in rip rap habitats that increases from upstream to downstream
 288 (online Resource, Figure A.2). Conversely, total macroinvertebrate abundance and taxa
 289 richness was not significantly related with river km for sand and rip rap habitat (all linear
 290 models $P > 0.05$).

291

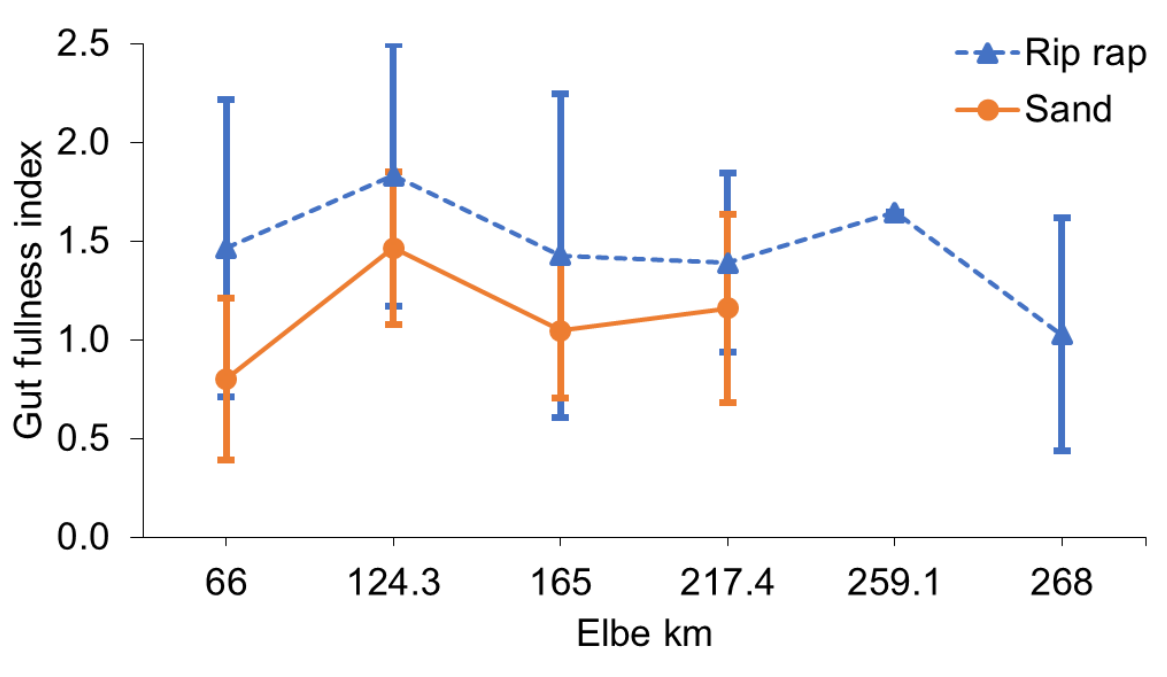


292
 293 **Fig. 3.** Non-metric multidimensional scaling ordination of macroinvertebrate communities collected
 294 from sand (circles) and rip rap habitats (triangles) at the Elbe River. Numbers refer to river km and
 295 depict the invasion gradient of *N. melanostomus*. Due to low sample sizes, rkm 259.1 is not shown.

296
 297 Within the rip rap habitats, total macroinvertebrate abundance decreased with increasing goby
 298 abundance (Pearson correlation $r = 0.67$; $P = 0.049$, $n = 6$). Within sand habitats, total
 299 macroinvertebrate abundance was not significantly correlated to goby abundance ($r = 0.02$, $P =$
 300 0.777 , $n = 6$).

301
 302 We examined the gut contents of 151 individuals of *N. melanostomus* (Table 1). The most
 303 abundant prey at both habitat structures were Crustacea (mostly *Dikerogammarus villosus*) and
 304 Diptera (mostly Chironomidae Gen. sp.) (online Resource, Figure A.1). Gut fullness was
 305 affected by habitat structure, and scores were overall lower for *N. melanostomus* caught at sand
 306 habitats. The lowest gut fullness was detected at rkm 268 (number of examined guts = 2) for
 307 rip rap habitats and at rkm 66 (number of examined guts = 5) for sand habitats (Fig. 4). Gut
 308 fullness was highest for both habitats at rkm124.3. There were no significant relationships with
 309 river km neither for sand (linear model, $R^2 = 0.43$, $P = 0.160$) nor the rip rap habitat ($R^2 = 0.19$,
 310 $P = 0.387$) (Fig. 4).

311

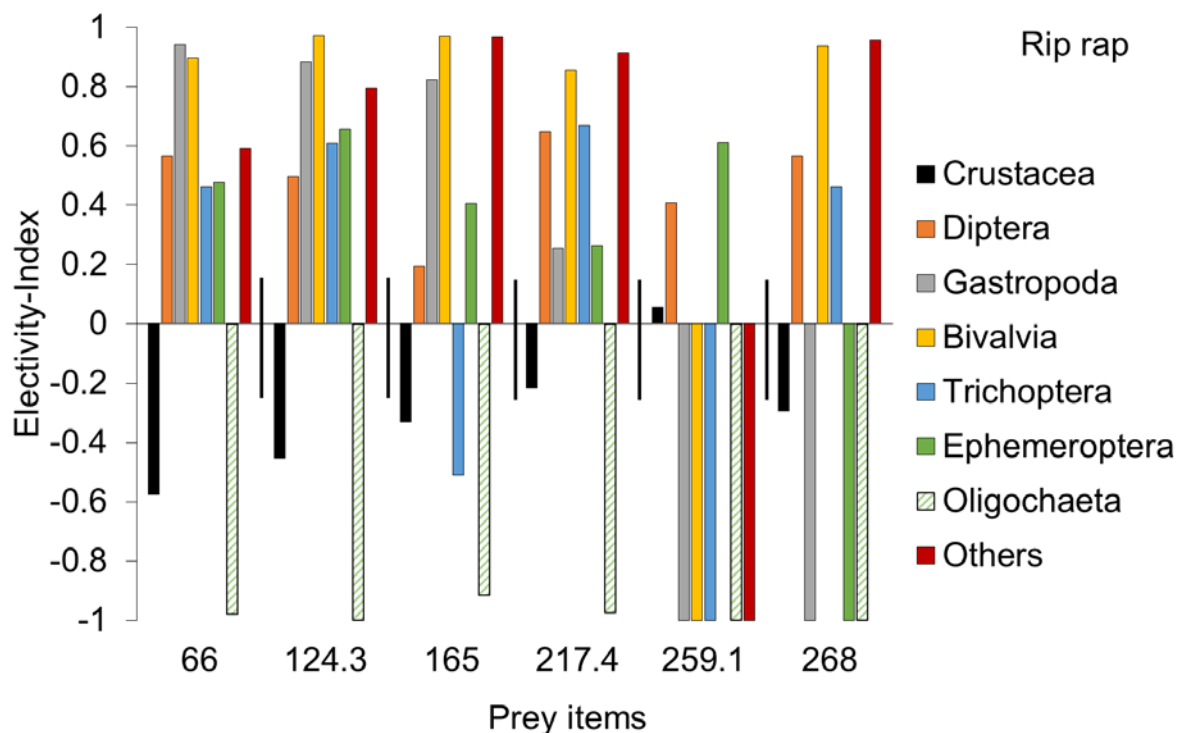


312
 313 **Fig. 4** Mean (\pm SD) gut fullness index (GFI) of *N. melanostomus* caught on sand and rip rap habitats
 314 along the invasion gradient. No data were available for sand habitats at rkm 259.1 and 268 as no gobies
 315 were present.

316
 317 **Prey selectivity**

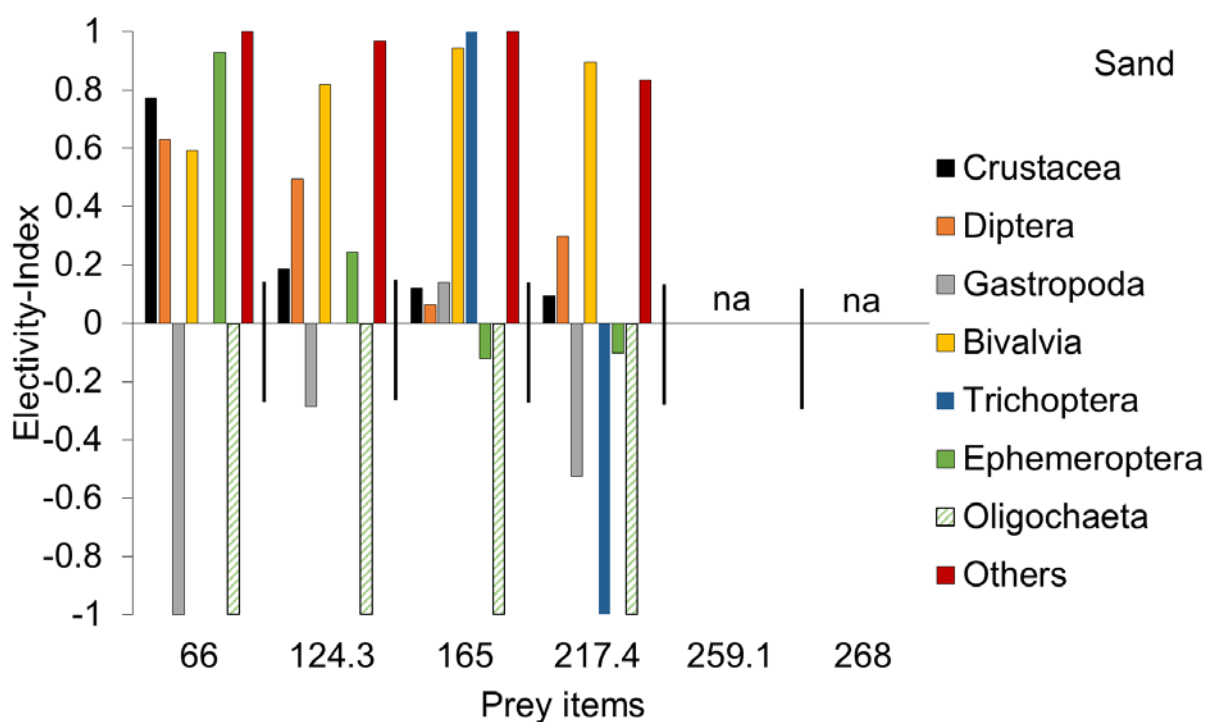
318 The analysis of the electivity index of *N. melanostomus* indicated differences in feeding
 319 preferences of *N. melanostomus* along the invasion gradient and between habitat structures (Fig.
 320 5). At rip rap habitats, preference was significant for two taxa along the invasion gradient.
 321 Linear models between rkm and the electivity index were significant for Crustacea ($R^2 = 0.70$,
 322 $P = 0.037$) and Gastropoda ($R^2 = 0.80$, $P = 0.016$) at rip rap habitats, indicating that feeding
 323 preferences of *N. melanostomus* for both taxa changed along the invasion gradient (Fig. 5).
 324 Whereas the electivity index indicated mainly negative values for Crustacea, with the exception
 325 of rkm 259.1, and positive values for Gastropoda, with an exception at rkm 259.1 and 268
 326 indicating negative ones. At the more recently invaded sites *N. melanostomus* from the rip rap
 327 habitat had a negative electivity index for Gastropoda, Bivalvia, Trichoptera and Oligochaeta
 328 at rkm 259.1 ($n = 1$) and for Crustacea, Gastropoda, Ephemeroptera and Oligochaeta at rkm 268
 329 ($n = 2$) (Fig. 5). There were no significant relationships between invasion history and electivity
 330 index for *N. melanostomus* caught in the sand habitat ($P < 0.05$ for all linear models). There was
 331 a tendency for an active selection of Crustacea and avoidance of Gastropoda (Fig. 5).

332



333

334



335

336 **Fig. 5** Prey preferences of *N. melanostomus* quantified by the electivity-index along the invasion
 337 gradient for rip rap and sand habitats. Positive values indicate a preference, and negative values indicate
 338 avoidance of the respective macroinvertebrate major group. "na" indicates sites with no gut available.

339

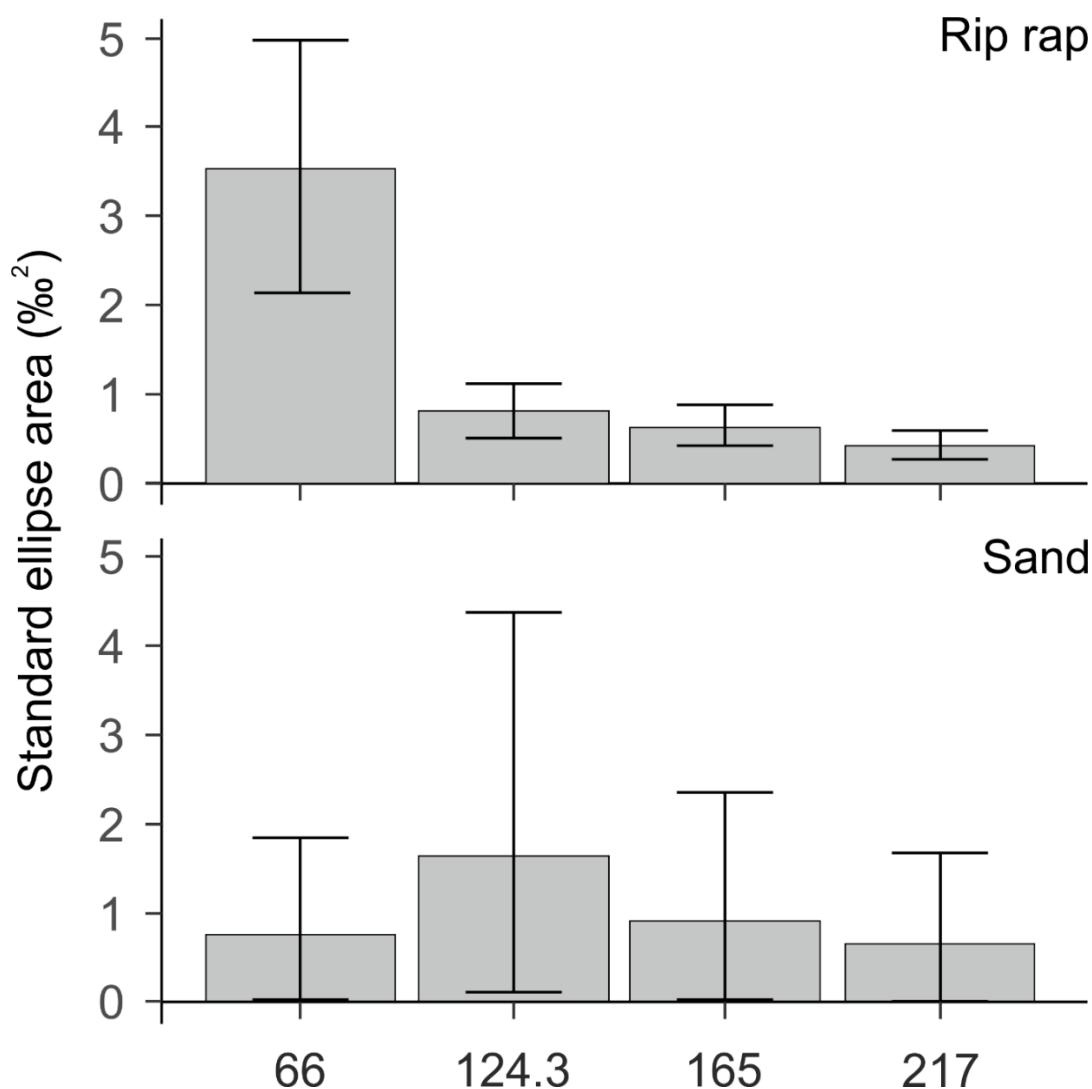
340 **Trophic niche size of *N. melanostomus* along the invasion gradient**

341 In general, $\delta^{15}\text{N}$ values ranged between 15‰ and 17‰, indicating that *N. melanostomus* fed at
 342 similar trophic positions regardless of invasion stage or habitat structure (online Resource,
 343 Figure A.3). A decreasing trophic niche size was detected along the invasion gradient for *N.*
 344 *melanostomus* caught in the rip rap habitats. With increasing invasion progress, the trophic
 345 niche width decreased and most recently invaded sites had smaller niches than those invaded
 346 earlier (Fig. 6).

347 For *N. melanostomus* caught in sand habitats, trophic niche size showed a unimodal pattern
 348 along the invasion gradient, and the largest trophic niche was found at rkm 124.3 (Fig. 6).

349 The size of the trophic niche decreased with increasing prey diversity in the guts in the rip rap
 350 but did not show a relationship in the sand habitats (Fig. 7).

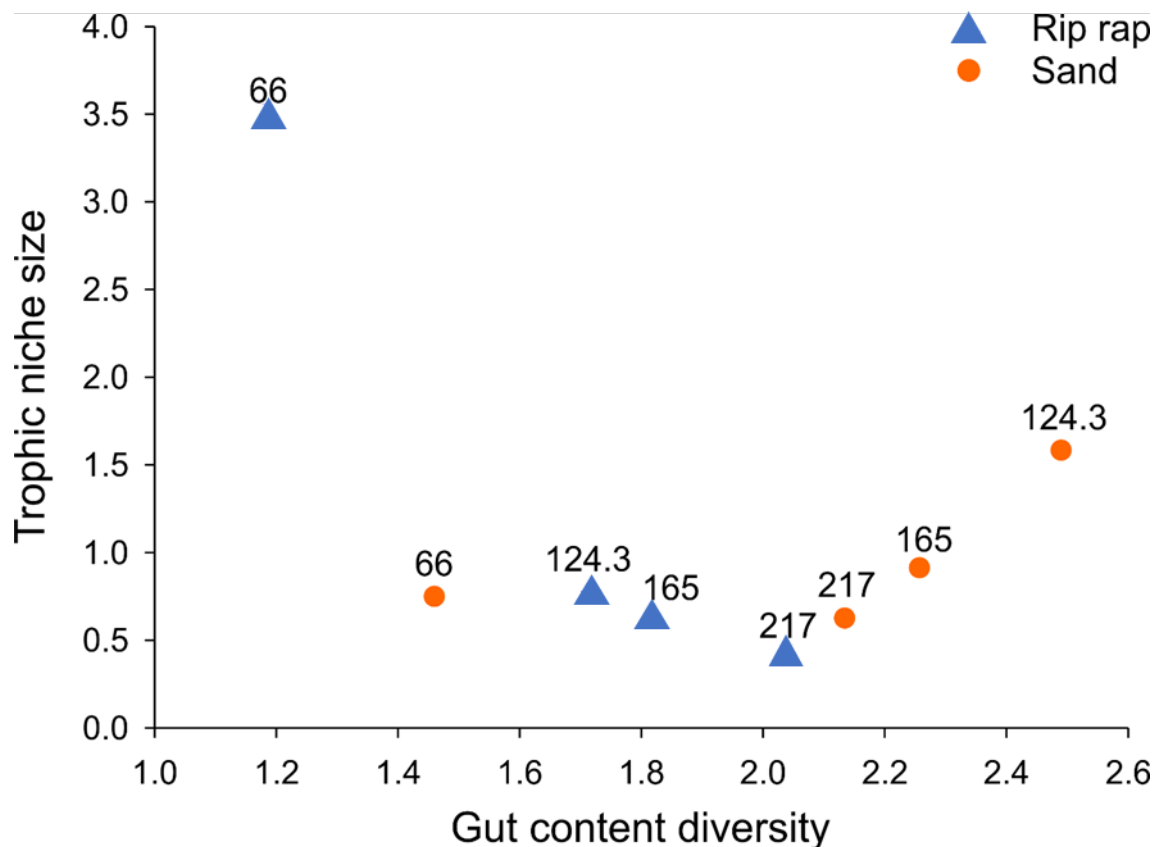
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352

353 **Fig. 6** Size of the trophic niche of *N. melanostomus* along the invasion gradient for rip rap and sand
 354 habitats expressed as the mean (\pm 95% highest posterior density intervals) standard ellipse area corrected
 355 for small sample sizes (SEAc).

356



357
 358 **Fig. 7.** Relationship between Shannon-Wiener diversity index of prey in guts and size of the trophic
 359 niche expressed as the mean of the standard ellipse area corrected for small sample sizes. Due to low
 360 sample sizes, rkm 259.1 and 268 are not shown.

361
 362 **Discussion**

363
 364 To predict future impacts for riverine ecosystems to be invaded by *N. melanostomus*,
 365 knowledge about the invasion process, habitat use and the factors that may influence the feeding
 366 behaviour and consequently trophic relationships of the invaded ecosystem remain essential.
 367 Here we were able to relate *N. melanostomus* feeding behaviour and trophic relationships to its
 368 respective invasion stage based on the known invasion pathways of *N. melanostomus* in the
 369 Elbe River (Nogueira Tavares et al., 2020).

370
 371 **Prey preference along the invasion gradient**

372
 373 Our results show differences in *N. melanostomus* gut contents depending on habitat structure
 374 and invasion stage. Hence, they agree with Raby et al. (2010), who demonstrated that the type
 375 and quantity of prey consumed by round goby could vary along its invasion pathway with
 376 different consumption at the area near their original introduction and those occupying the edges

377 of range expansion. As expected, round gobies were more abundant in rip rap than in sand
378 habitats, which goes in line with previous studies indicating that the rip rap structure is more
379 frequently selected by round goby (Ray & Corkum, 2001). Further, we observed differences in
380 gut fullness among the different habitat structures. The overall lower gut fullness at sand
381 habitats compared to rip rap habitats could be the result of rip rap structures generally providing
382 a higher species richness and abundance of invertebrates (Brunke et al., 2002; Ray & Corkum,
383 2001) and therefore a higher resource availability.

384 Prey abundance is crucial when analysing *N. melanostomus* dietary progress at the different
385 invasion stages along a river course. The higher macroinvertebrate abundance at the latest
386 invaded sites compared to the longer invaded ones (online Resource, Figure A.2) can be
387 explained by lower predation pressure (Brandner et al., 2013b), whereas the reduced
388 macroinvertebrate abundance at the earlier invaded sites could be a result of an earlier sustained
389 high consumption rate by invasive round gobies. However, the lower *N. melanostomus*
390 abundance and biomass detected at sampling site rkm 66 could indicate a previous high intra-
391 and interspecific competition for food and shelter at this sampling site (McCallum et al., 2014).
392 The results of our study at the two most recently invaded sites are based on a small number of
393 analysed *N. melanostomus* individuals ($n \leq 2$). Therefore, round goby predation may not have
394 affected macroinvertebrate abundance that strong by now.

395 At sand habitats, we observed a tendency of *N. melanostomus* to feed preferentially on
396 Crustacea with the preference continuously decreasing from upstream to downstream.
397 Nevertheless, a significant change in feeding preference along the invasion gradient was only
398 observed at rip rap habitats. Here, the preference increased for Crustacea and decreased for
399 Gastropoda from upstream to downstream. Different than at sand habitats, the electivity index
400 values for Crustacea at rip rap habitats were mainly negative along the invasion gradient,
401 indicating a general avoidance of this item at rip rap habitats in opposite to the general
402 preference at sand habitats. However, specific for Crustacea at rip rap habitats, a trend of
403 increasing electivity index values from upstream to downstream were observed. Here, the
404 longer invaded sites indicate a stronger avoidance (index value close to -1), while the latest
405 invaded sites show a trend of a more random feeding (electivity index value close to 0). Such
406 effects of negative electivity values but dominant predation were also observed by Piria et al.
407 (2016) in the Sava and Kupa Rivers, who found Gastropoda being the main prey category for
408 *N. melanostomus* and also the most abundant item in the environment, even though electivity
409 index indicate avoidance of this item. Another example is the study by Polačik et al. (2009) in

410 the Danube River, who hypothesized that Mollusca are an unavoidable alternative, rather than
411 the most preferred prey.

412 Our study indicated, that the preference for Gastropoda decreases along the invasion gradient,
413 with an avoidance of these taxa (with index values of -1) at the more recent invaded sites.

414 Previous studies reported, that crustaceans are preferred prey items in goby diet (Mikl et al.,
415 2017; Vašek et al., 2014), while Gastropoda are avoided (Brandner et al., 2013a; Piria et al.,
416 2016; Vašek et al., 2014).

417 Henseler et al. (2021) described that the effects of *N. melanostomus* might be stronger for
418 dominant taxa and also Borcherdig et al. (2013) show that round gobies prey on available
419 organisms in high abundances, which fits the results of our study, as Crustacea were a
420 frequently represented macroinvertebrate taxa in the Elbe River. In particular the Ponto-Caspian
421 amphipod *Dikerogammarus villosus*, an invasive species to the Elbe River (Worischka et al.,
422 2018) provides an abundant food source for *N. melanostomus*.

423 Not represented with significant values, but worth discussing, is the trend of *N. melanostomus*
424 consumption preferences on Bivalvia along the invasion gradient in both habitats, as indicated
425 by the mainly positive electivity index values (Fig. 5). However, compared to the other
426 available taxa present at both habitats, Bivalvia were much less abundant (online Resource,
427 Figure A.2). Nevertheless, the mainly positive values correspond with the preferred diet of *N.*
428 *melanostomus* in their native ranges, where molluscs, mainly bivalves, represent the most
429 important food items (Brandner et al., 2013a). Other studies even determined *N. melanostomus*
430 as a distinct molluscivore (French & Jude, 2001; Ray & Corkum, 1997).

431 However, it remains difficult to determine why round gobies select a particular prey type, and
432 several factors like prey size, mobility, fixing to substratum, activity and digestibility may
433 influence the choice (Roşca et al., 2010). Henseler et al. (2020) further state that prey taxa might
434 depend on the study area and, most likely, on the prey items that are available in a specific
435 environment.

436

437 **Trophic niche size of *N. melanostomus* along the invasion gradient**

438

439 Our results indicate a decreasing trophic niche coinciding with an increasing prey diversity in
440 the guts of *N. melanostomus* caught at rip rap habitats (Fig. 7). This result seems
441 counterintuitive as one would expect an increasing trophic niche width with an increasing
442 diversity of ingested prey. Nevertheless, stable isotopes capture the middle-to-long-term
443 patterns of assimilation while gut content analyses provide information on short-term feeding

444 patterns of ingested prey (Brandner et al., 2013a). Hence, our results suggest that both
445 techniques should be used in conjunction to quantify trophic dynamics of invasive fish.
446 We found differences in trophic niche width depending on the invasion gradient and habitat
447 structure (Fig. 6). Smaller niche width in both habitat structures were detected for *N.*
448 *melanostomus* caught at the latest invaded river sites and were widest at the earlier invaded sites
449 and, therefore, more established populations (online Resource, Figure A.3). This is consistent
450 with other studies, e.g. (Brandner et al., 2013b; Herlevi et al., 2018; Pettitt-Wade et al., 2015)
451 that indicated a broader niche for round goby in longer-established populations compared to a
452 newly established round goby population. Brandner et al. (2013b) observed that a less
453 intraspecific competition exhibited a more homogenous diet than an established population,
454 where a pronounced ontogenetic diet shift was visible, mirrored in a broad dietary niche. We
455 assume that the invasion process of *N. melanostomus* at those sites that were invaded earlier is
456 still ongoing and that competitive interactions have still not decreased *N. melanostomus* trophic
457 niche to minimize competition (Rakauskas et al., 2020). But we expected, that the trophic niche
458 may change with time as interspecific competition increases. For the least invaded sites, we can
459 assume that *N. melanostomus* occupies previously vacant dietary niches, which has facilitated
460 further colonization (de Carvalho et al., 2019).

461 Differences in trophic niche width show that *N. melanostomus* feeding behaviour
462 change with the invasion stage and are thus dynamic. Therefore, knowledge on the invasion
463 stage is of high importance, not only for understanding further dispersal performance but also
464 for predicting upcoming impacts on the food web structure as a result of abundance and habitat
465 occupancy and upcoming trophic links within the food web (Rakauskas et al., 2020). Such
466 knowledge can help develop appropriate ecosystem conservation and preventative management
467 actions. Taken together, our results show that diet composition and prey preferences of *N.*
468 *melanostomus* differ depending on the invasion stage and habitat structure. Future studies
469 should follow the invasion process and quantify trophic dynamics as it is likely to change over
470 time with consequences for prey communities.

471

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