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**Preparation of MnO₂ coated fibers for gamma spectrometric measurements - A
comparison of four practical approaches**

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Abstract: The analysis of natural radium-in-water activity concentrations is for two reasons of general interest: (1) radium in natural waters may pose a problem with regard to radiation protection and (2) radium isotopes in natural waters can be used as environmental tracers in hydrological studies. A state-of-the-art method for radium extraction from (generally large) water sample volumes is radium adsorption onto MnO₂ coated acrylic fibers. In our study we comparatively evaluated four methodical approaches for post-extraction preparation of the fiber to allow gamma spectrometric measurements. The methods included (1) straightforward measurement of the loose fiber, (2) compressing the fiber after mixing it with an adhesive, (3) combustion of the fiber and embedding the ash in candlewax, and (4) leaching of the fiber and embedding the resulting precipitate in candlewax. The aim of the study was to compare the advantages and disadvantages of the four preparation approaches with respect to their individual practicability. Even though the methodical fiber preparation approaches have been suggested in the literature before (as cited in this paper), results of their direct practical comparison have not been presented yet. Our study revealed that balancing practical sample preparation effort against data reproducibility suggests a measurement of the compressed fiber applying an adhesive to be the preferable approach.

Preparation of MnO₂ coated fibers for gamma spectrometric measurements - A comparison of four practical approaches

1 Introduction

The omnipresence of radium in natural waters can be seen from two ambivalent perspectives. On the one hand, elevated radium concentrations in waters that are used for drinking or irrigation can pose a serious health risk. Related problems have e.g. been reported for fossil groundwater originating from deep sandstone aquifers on the Arabian Peninsula and in Northern Africa (Schubert et al., 2011; Vengosh et al., 2009). Furthermore, radium in “produced” waters (brines) of the oil and gas industry may be of related concern (e.g. Vengosh et al., 2017; Rowan et al., 2011). On the other hand, the presence of radium in natural waters can also be considered beneficial. In a number of studies it was demonstrated that the four naturally occurring radium isotopes (²²⁴Ra ($t_{1/2} = 3.7$ d); ²²³Ra ($t_{1/2} = 11.4$ d); ²²⁸Ra ($t_{1/2} = 5.7$ y); ²²⁶Ra ($t_{1/2} = 1600$ y)) can be used as tracers or indicators in a wide range of hydrological applications such as water migration investigations or seawater/groundwater interaction studies (e.g. Moore and Sarmiento, 2008; Moore 2003). In both cases, i.e. radium as a potential threat and radium as an environmental tracer, straightforward and reliable analysis of radium in natural waters is of key interest.

The water volume that is required for sound radium analysis depends on the radium activity concentration of the water and on the applied detection technique. For instance, if ²²⁶Ra is the isotope of interest, detection by means of ICP-MS requires even at low activity concentrations (as they are observed e.g. in seawater (²²⁶Ra ~ 1.3 mBq/l)) only rather small water volumes of about 200 - 300ml (Bourquin et al., 2011; Varga, 2008). If, however, all four natural radium isotopes are to be detected by means of gamma spectrometry much larger water volumes are necessary, thus requiring a pre-concentration of radium prior to analysis.

25 Pre-concentration of radium by means of enforced evaporation is only feasible for water
26 sample volumes of up to several tens of liters. Such rather small sample volumes can also be
27 treated by radium extraction from the water applying special adsorbents such as specific
28 resins (Maxwell, 2006), MnO₂-coated polyamide discs (Eikenberg et al., 2001) or
29 3M EMPORE™ Radium Rad disks (Joannon and Pin, 2001).

30 The most appropriate extraction method for much larger sample volumes (up to several
31 hundred liters) is adsorption onto MnO₂ coated acrylic fiber (e.g. Moore and Reid, 1973;
32 Moore and Arnold 1996; Bourquin et al., 2008). This extraction approach requires pumping
33 of the water sample with a flow rate of about 1 l/min through ~20 g of MnO₂ coated fiber that
34 is placed in a column. A suitable state-of-the-art column is readily designed for subsequent
35 analysis of the short lived ²²³Ra (half-life 11.4 days) and ²²⁴Ra (half-life 3.7 days) by means of
36 Radium Delayed Coincidence Counting (“RaDeCC”; Moore and Arnold, 1996). It is common
37 practice that after the RaDeCC analysis the long lived ²²⁶Ra (half-life 1600 years) and ²²⁸Ra
38 (half-life 5.75 years) are determined by means of gamma spectrometry. This requires the
39 adaptation of the MnO₂ coated fibers to the specific geometry of the gamma detector used, i.e.
40 further preparation of the fiber is necessary.

41 Conventional fiber preparation methods are (i) quantitative combustion of the fiber (Charette
42 et al., 2001, 2007) or (ii) leaching of the fiber to extract the radium for subsequent recovery as
43 Ra/BaSO₄ precipitate (e.g. Moore, 1987; Moore et al., 1985; Moore and Santschi, 1986).

44 Although both approaches show good results they have the disadvantage of being rather time-
45 consuming and laborious. Moreover, the GEOTRACES inter-comparison study for radium
46 isotopes showed rather poor agreement of the participating laboratories in the ²²⁶Ra
47 determinations, which may be also due to problems of adequately calibrating and preparing
48 fibers for gamma spectrometric measurements (Charette et al., 2012).

49 In our study we comparatively evaluated four methods to prepare MnO₂ coated acrylic fiber
50 for ²²⁶Ra and ²²⁸Ra measurement by means of gamma spectrometry. The four methods
51 include: (1) placing the loose fiber into a defined gamma detection geometry without any
52 further processing, (2) molding of the fiber into a defined pellet applying an adhesive,
53 (3) combustion of the fiber and embedding the ash in candlewax, and (4) leaching of the fiber
54 and embedding the subsequently produced Ra/BaSO₄ precipitate in candlewax. Aim of the
55 study was to compare the advantages and disadvantages of the four procedures based on
56 practical considerations including (i) the extent of the required practical preparation efforts,
57 (ii) the reproducibility of the gamma measurement results and (iii) the magnitude of self-
58 absorption resulting from the sample matrix.

59

60 2 Material and methods

61 From a certified ²²⁶Ra stock solution (provided by the national German metrology institute
62 Physikalisch-Technische Bundesanstalt) with a concentration uncertainty of 1.4 % twelve
63 1 liter aqueous sub-solutions with defined radium concentrations were produced (*cf.* Tab. 1).
64 Distilled H₂O was used for the preparation of the sub-solutions. The twelve sub-solutions
65 (with a pH around 7) were each filled in a 1 liter Marinelli beaker equipped with an inlet and
66 an outlet port. For the production of twelve defined fiber specimens each beaker was
67 connected in a closed loop to a peristaltic water pump and a column that contained 15 g of
68 MnO₂ coated acrylic fiber (dry weight). Each sub-solution was pumped through its individual
69 closed-circuit system at a pump rate of 2 l/min and a temperature of 20°C. In order to allow
70 complete adsorption of the dissolved radium onto the fibers, each sub-solution was circulated
71 for one hour, i.e. each sub-solution was pumped 120 times through its fiber column. For
72 verification of quantitative radium adsorption onto the fibers each of the twelve residual sub-
73 solutions was subsequently analyzed for remaining traces of dissolved radium by means of

74 gamma spectrometry. If remaining radium was detected the quantity was taken into account
75 for definition of the nominal radium activity of the respective fiber specimen. We assume any
76 adsorption of radium on tubings, pump and beaker to be negligible.

77 The procedure resulted in twelve fiber specimens with known, i.e. nominal ^{226}Ra activities.
78 Four of the specimens contained a comparably low and eight of the specimens a comparably
79 high activity (*cf.* Tab. 1). After drying the twelve fiber specimens at room temperature, they
80 were grouped in four sets with three specimens each, two of relatively high activity and one
81 of relatively low activity. The four sets of specimens were treated in abovementioned four
82 different ways of sample preparation (see below for details).

83 Due to the four different physical characteristics of the specimens resulting from the four
84 ways of fiber treatment a different magnitude of self-absorption was to be expected for each
85 of the four sets, thus requiring individual gamma efficiency calibrations of the four different
86 matrixes. To achieve this, the specimen with the highest nominal activity from each set was
87 defined as “standard” for this particular set. The remaining two specimens were considered as
88 “samples” of the set (Fig. 1, Tab. 1). Thereby it could be guaranteed that the samples and the
89 standard were handled in the same geometric setup and have the same physical and chemical
90 characteristics (e.g. density) and hence the same self-absorption. The activity concentrations
91 of the two samples of each set were then determined by relating the detected gamma counts to
92 the corresponding counts detected in the standard of the same set applying the rule of
93 proportion.

94

95 3 Experimental

96 In order to produce the four sets of fiber specimens the fibers were taken from the individual
97 columns, dried and prepared as given in detail below and schematically illustrated in Fig. 1.

98 The preparation resulted in the four specimen types shown in Fig. 2.

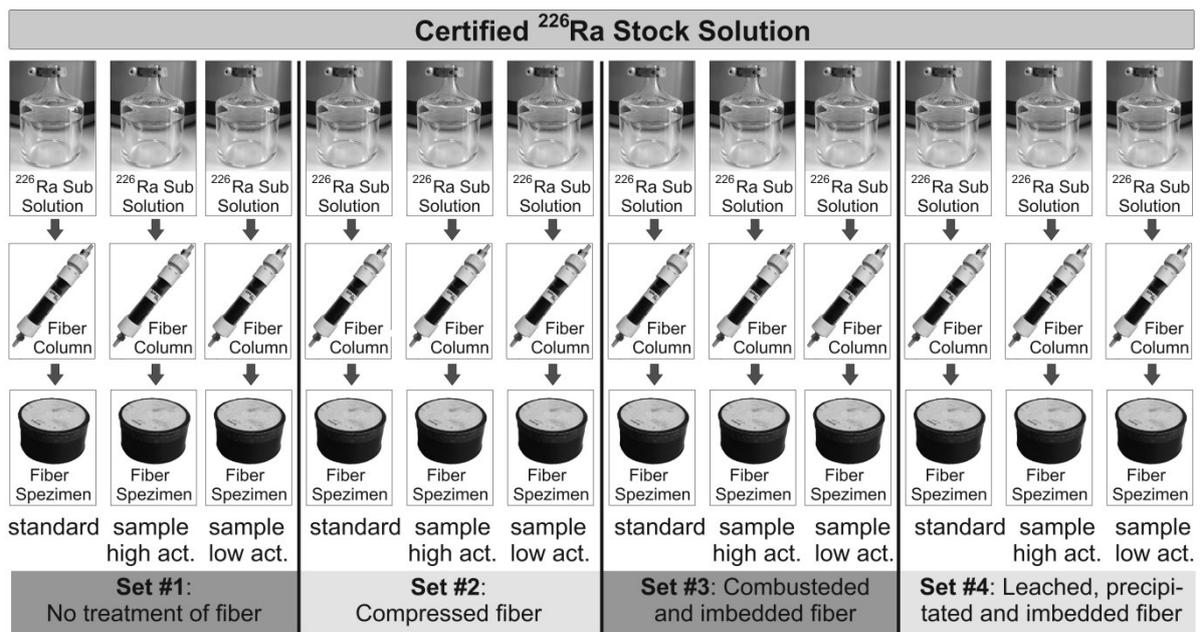
99 For “set #1” the fiber was transferred into cylindrical radon tight HDPE capsules, 7 cm in
100 diameter and 3.5 cm in height. The fiber was not treated in any way but just placed in the
101 HDPE capsules so that after closing and sealing the lids tightly, the fiber filled the capsules
102 homogeneously.

103 For “set #2” the fiber was compressed into pellets by means of a hydraulic press applying
104 125 kN of pressure. The mold of the press allowed the production of round and flat pellets
105 with a diameter of 7 cm and a height of 1 cm, which fitted tightly into the cylindrical HDPE
106 capsules also used for “set #1”. First tests, in which the pure fiber was compressed, revealed
107 that the fiber pellets do not hold their defined shape after pressure release, which made it
108 impossible to obtain a constant and defined detection geometry. For stabilization of the pellet
109 shape the fiber was hence mixed with powder sugar as an adhesive. The sugar was previously
110 analyzed for traces of radium using gamma spectrometry, but no radium activity was
111 detected. Other potential adhesive materials, such as talcum, milk powder and chalk had also
112 been tested; however, the powder sugar showed the most satisfying results regarding handling
113 and stability of the fiber pellets.

114 For “set #3” the fiber was slowly and carefully combusted in an open ceramic bowl at about
115 1200°C using a Bunsen burner. The process took about 30 min per specimen. Subsequently,
116 after cooling, the ash was homogeneously mixed with fine grained pure candlewax (Merck).
117 A blank of the wax was analyzed for traces of radium using gamma spectrometry with
118 negative results. The homogeneous wax/ash mixture was subsequently compressed by means
119 of the hydraulic press described above thereby producing pellets of the same dimensions as
120 “set #2” that were also placed in the abovementioned HDPE capsules.

121 For “set #4” the fiber was leached using 6 N HCl at a temperature close to the boiling point.
122 After quantitative leaching the fiber had turned its color to plain white, indicating the
123 complete release of MnO₂ and hence of the adsorbed radium. Next the dissolved radium was

124 quantitatively precipitated from the acid solution as Ba/RaSO₄ (Moore et al., 1985). The
 125 precipitate was dried and homogeneously mixed with fine grained pure candlewax as used for
 126 “set #3”. The mixture was compressed by means of the hydraulic press again producing
 127 pellets that fitted tightly in the HDPE capsules.
 128 The twelve HDPE capsules (four different sets, each with one “standard” and two “samples”,
 129 cf. Fig. 1) where finally sealed by attaching the lid onto the capsule using a flexible glue
 130 (Hylomar® M) and additionally flexible sticky tape (Cellpack Premio 233) in order to
 131 minimize radon escape. An equilibration time of four weeks was allowed for radioactive
 132 equilibrium between ²²⁶Ra, its daughter ²²²Rn and the short-lived radon progenies ²¹⁴Pb and
 133 ²¹⁴Bi.



134

135 Fig.1: Fiber sample preparation scheme



136

137 Fig. 2: Four different sample types: set #1 unprocessed fiber, set #2 compressed fiber,

138 set #3 fiber combusted and mixed with candlewax, set #4 fiber leached and Ra/BaSO₄

139 precipitate mixed with candlewax (*cf.* Fig. 1)

140

141 Radium activity measurement of all four standards and eight samples was carried out by low-

142 level gamma-spectrometry based on the distinct gamma emission energies of ²²⁶Ra

143 (186.1 keV) as well as of the short-lived radon progenies ²¹⁴Pb (295.2 keV, 351.9 keV) and

144 ²¹⁴Bi (609.3 keV, 1120.3 keV, 1238.1 keV, 1764.5 keV, 2204.1 keV) using a coaxial Low-

145 Energy HPGe detector, n-type (ORTEC), with an active volume of 39 cm³ and a 0.5 mm Be

146 window. Each measurement lasted for about 24 h. Spectra analysis was performed with the

147 software GAMMAW[®] (Dr. Westmeier GmbH, Germany).

148

149 4 Results

150 Tab. 1 displays the detection results for the 4 x 2 samples. Given for each sample (i) its

151 nominal ²²⁶Ra activity (i.e. the target value), (ii) the activity that resulted from directly

152 evaluating the 186 keV ²²⁶Ra gamma line, (iii) the activity derived from the combined

153 evaluation of the major ²¹⁴Pb and ²¹⁴Bi gamma lines, and (iv) the set specific cpm/Bq count

154 rate at 186 keV as indicator for the individual intensity of self-absorption.

155 It has to be pointed out, that even though the two set #1 samples were filled homogeneously
156 with fiber, no homogenous radium distribution could be taken for granted within the capsules.
157 The reason for that is that, while pumping the sub-solutions through the fiber columns for
158 radium extraction, radium that might have been preferentially extracted in the upstream part
159 of the respective fiber column resulting in an unevenly distribution of radium in the fiber
160 volume. Water flow along preferential pathways in the fiber may also have caused an
161 inhomogeneous distribution of radium on the fiber. Hence the set #1 samples were measured
162 twice, once upside-up and once upside-down. The specimen that was chosen as standard for
163 set #1 showed identical results either way.

164 The activities displayed in Tab. 1 show that the measurements resulted in activities of the
165 individual samples that match the corresponding nominal values within the counting
166 uncertainties. The only exception is the set #1 sample with the lower activity. The
167 concentrations derived from the ^{226}Ra gamma line differ significantly from the target value.

168 As discussed above this disagreement can be explained with an inhomogeneous spatial
169 distribution of radium within this particular capsule resulting in a larger (or if measured
170 upside-down a smaller) distance between radiation source and detector giving rise to a lower
171 (or if measured upside-down a higher) count rate.

173 Tab. 1: Nominal activities of the samples; Activity concentrations derived from the ^{226}Ra
 174 gamma line; Activity concentrations derived from the seven major gamma lines of the short
 175 lived ^{222}Rn progenies ^{214}Pb and ^{214}Bi ; Set specific count rate [cpm/Bq] of the ^{226}Ra gamma
 176 line at 186 keV

	nominal activity [Bq]	^{226}Ra [Bq]	^{214}Pb & ^{214}Bi mean [Bq]	186 keV count rate [cpm/Bq]
Set #1: Unprocessed Fiber				
High (upside up)	310 ± 15.5	313 ± 16	325 ± 40	0.11
high (upside down)		315 ± 17	320 ± 33	
Low (upside up)	105 ± 5.3	94 ± 4	107 ± 14	
low (upside down)		115 ± 6	108 ± 25	
Set #2: Compressed Fiber				
High	325 ± 16.0	319 ± 16	337 ± 44	0.17
Low	155 ± 7.8	159 ± 7	153 ± 31	
Set #3: Combusted Fiber				
High	315 ± 15.8	315 ± 15	307 ± 30	0.12
Low	145 ± 7.2	148 ± 8	139 ± 18	
Set #4: Precipitated Ba/RaSO₄				
High	215 ± 10.8	212 ± 12	217 ± 24	0.06
Low	133 ± 7.0	138 ± 6	131 ± 20	

177

178 5 Discussion

179 The aim of the study was to evaluate the advantages and disadvantages of four different fiber
 180 preparation approaches for measuring radium adsorbed on MnO_2 -fibers by means of gamma
 181 spectrometry. That evaluation of the four approaches was done by comparing (i) the amount
 182 of their individual practical preparation efforts, (ii) their individual reproducibilities of the

183 nominal activities and (iii) the magnitude of self-absorption resulting from the sample matrix
184 indicated by the individual count rate at 186 keV [cpm/Bq].

185 The measurement of samples without any fiber treatment (set #1) requires the least practical
186 fiber preparation effort. The fiber was simply stuffed into the detection geometry (i.e. the
187 HDPE capsules) and measured after decay equilibrium between radium and short-lived radon
188 progenies had established. Still, as shown in Tab. 1 one of the two samples revealed a
189 significant discrepancy between detected values and the nominal activity when the ^{226}Ra
190 186 keV line is evaluated (even though the mean value of two results agrees well with the
191 nominal value within the uncertainties). This lower data reproducibility can be explained with
192 the above mentioned poor degree of radium distribution homogeneity within the fiber.

193 A different picture emerges for set #1 when the ^{226}Ra activity of the samples is evaluated
194 based on the short-lived radon progenies. Both values match the nominal value well. The
195 explanation of this observation is that radon (as a gas) spreads homogeneously and
196 independently of the radium distribution within the capsule, thus resulting in equal gamma
197 activities of its short-lived progeny in spite of a potentially inhomogeneous radium
198 distribution. Hence, the potential error that arises from a poor degree of radium distribution
199 homogeneity within the fiber can be avoided by not relying on the ^{226}Ra line at 186 keV but
200 by rather using the activities of the short-lived radon progenies. For reliable results sample
201 capsules have to be sealed radon tight in order to exclude radon loss during the required
202 3 weeks equilibration time (Scholten et al. 2013). The low self-absorption due to the loose
203 packing of set #1, on the one hand, and the rather large fiber volume of the samples that
204 increases its effective distance to the detector thus decreasing the [cpm/Bq] count rate, on the
205 other hand, keeps the count rate at 186 keV at a level comparable to that of set #3 (Tab. 1).

206 The fiber preparation applying a compaction of the fiber (set #2) is hardly more laborious
207 than the measurement of the unprocessed fiber (set #1). Still, compared to set #1 the physical

208 homogeneity of the set #2 samples is significantly increased due to the lower volume of the
209 pellets, which is of general advantage for the data reproducibility. On top of that the relatively
210 low self-absorption of the pellets and the short effective distance between detector and sample
211 result in a higher [cpm/ Bq] count rate (Tab. 1). That makes this fiber preparation approach
212 suitable for samples with low activity concentrations.

213 The two fiber preparation methods that involve combustion and leaching of the fibers (set #3
214 and set #4, respectively) are much more time consuming and laborious than the two methods
215 applied for sets #1 and #2. On the other hand, a strong advantage is that the homogeneity of
216 the sample sets #3 and #4 increases significantly due to the thorough mixing of either the ash
217 or the precipitate with the candle wax. However, the addition of the candlewax increases the
218 self-absorption of the samples, thus reducing the [cpm/Bq] count rate (Tab. 1). In the case of
219 set #4 that effect is even more noticeable due to the strong self-absorption potential of the
220 precipitated Ba/RaSO₄.

221

222 6 Conclusions

223 The described advantages and disadvantages of the four different sample preparation
224 approaches are qualitatively summarized in Tab. 2. It can be stated that the approach, which is
225 represented by set #2 (compressed fiber) turned out to be the preferable one. Requiring only a
226 low sample preparation effort it leads to results that show a satisfying data reproducibility.

227 The approach represented by set #1 (unprocessed fiber) shows comparable advantages if the
228 radon progeny data is used to quantify radium activity concentrations.

230 Tab. 2: Advantages and disadvantages of the four different sample preparation approaches

	Preparation Effort	Data Reproducibility	Count Rate per Bq
Unprocessed Fiber	+	-	±
Compressed Fiber	±	+	+
Combusted Fiber	-	+	±
Precipitated Ba/RaSO ₄	-	+	-

231 + above average; ± average; - below average

232

233 References

234 Bourquin, M., van Beek, P., Reyss, J.-L., Souhaut, M., Charette, M.A., Jeandel C., 2008.
 235 Comparison of techniques for pre-concentrating radium from seawater. Mar. Chem. 109, 226-
 236 237.

237 Charette, M.A., Dulaiova, H., Gonnee, M.E., Henderson, P.B., Moore, W.S., Scholten, J.C.,
 238 Pham, K.M., 2012. GEOTRACES radium isotopes interlaboratory comparison experiment.
 239 Limnol. Oceanogr. Methods 10, 451-463.

240 Charette, M.A., Buessler, K.O., Andrews, J.E., 2001. Utility of radium isotopes for
 241 evaluating the input and transport of groundwater derived nitrogen to a Cape Cod estuary.
 242 Limnol. Oceanogr. 46 (2), 465-470.

243 Charette, M.A., Gonnee, M.E., Morris, P., Statham, P., Fones, G., Planquette, H., Salter, I.,
 244 Naveira Garabato, A., 2007. Radium as tracers of iron sources fueling a Southern Ocean
 245 phytoplankton bloom. Deep-Sea Res. II 54, 1989-1998.

246 Eikenberg, J., Tricca, A., Vezzu, G., Bajo, S., Ruethi, M., Surbeck, H., 2001. Determination
 247 of ²²⁸Ra, ²²⁶Ra and ²²⁴Ra in natural water via adsorption on MnO₂-coated discs. J. Envir.
 248 Radioac. 54, 109-131.

249 Joannon, S., Pin, C., 2001. Ultra-trace determination of ^{226}Ra in thermal waters by high
250 sensitivity quadrupole ICP-mass spectrometry following selective extraction and
251 concentration using radium-specific membrane disks. *J. Anal. At. Spectrom.* 16, 32-37.

252 Maxwell, S.L., 2006. Rapid method for ^{226}Ra and ^{228}Ra analysis in water samples. *J.*
253 *Radioanal. Nucl. Chem.* 270 (3), 651-655.

254 Moore, W.S., 1987. Radium 228 in the SouthAtlantic Bight. *J. Geophys. Res.* 92 (C5), 5177-
255 5190.

256 Moore, W.S., Key, R.M., Sarmiento, J.L., 1985. Techniques for precise mapping of ^{226}Ra and
257 ^{228}Ra in the Ocean. *J. Geophys. Res.* 90 (C4), 6983-6994.

258 Moore, W.S., Santschi, P.H., 1986. Ra-228 in the deep Indian Ocean. *Deep-Sea Res.* 33 (1),
259 107-120.

260 Moore, W.S. 2003. Sources and fluxes of submarine groundwater discharge delineated by
261 radium isotopes. *Biogeochem.* 66, 75-93.

262 Moore W.S., Sarmiento J.L., Key R.M., 2008. Submarine groundwater discharge revealed by
263 ^{228}Ra distribution in the upper Atlantic Ocean. *Nat. Geosci.* 1, 309-11.

264 Moore, W.S., Arnold, R., 1996. Measurement of ^{223}Ra and ^{224}Ra in coastal waters using a
265 delayed coincidence counter. *J. Geophys. Res.* 101, 1321-1329.

266 Moore, W.S., Reid, D.F., 1973. Extraction of Radium from Natural Waters Using
267 Manganese-Impregnated Acrylic Fibers. *J. Geophys. Res.* 78, 8880-8886.

268 Rowan, E., Engle, M., Kirby, C., Kraemer, T., 2011. Radium content of oil- and gas-field
269 produced waters in the northern Appalachian Basin (USA) - Summary and discussion of data.
270 U.S. Geological Survey Scientific Investigations Report, 31 pp.

271 Scholten, J.C., Osvath, I., Pham, M.K., 2013. ^{226}Ra measurements through gamma
272 spectrometric counting of radon progenies: How significant is the loss of radon?. *Mar. Chem.*
273 156, 146-152.

274 Schubert, M., Schüth, Ch., Michelsen, N., Rausch, R., Al-Saud, M., 2011. Investigation and
275 Treatment of Natural Radioactivity in Large-Scale Sandstone Aquifer Systems. IJWRAE
276 1(1), 25-32.

277 Vengosh, A., Kondash, A., Harkness, J., Lauer, N., Warner, N., Darrah, T.H., 2017. The
278 geochemistry of hydraulic fracturing fluids. Procedia Earth Planet. Sci. 17, 21-24.

279 Vengosh, A., Hirschfeld, D, Vinson, D., Dwyer, G., Raanan, H., Rimawi, O., Al-Zoubi, A.,
280 Akkawi, E., Marie, A., Haquin, G., Zaarur, S., Ganor, J., 2009. High Naturally Occurring
281 Radioactivity in Fossil Groundwater from the Middle East. Environ. Sci. Technol. 43 (6),
282 1769-1775.

