

This is the accepted manuscript version of the contribution published as:

Wei, M., Harnisch, F., Vogt, C., Ahlheim, J., Neu, T.R., Richnow, H.H. (2015):
Harvesting electricity from benzene and ammonium-contaminated groundwater using a microbial fuel cell with an aerated cathode
RSC Advances **5** (7), 5321 - 5330

The publisher's version is available at:

<http://dx.doi.org/10.1039/c4ra12144a>

RSC Advances



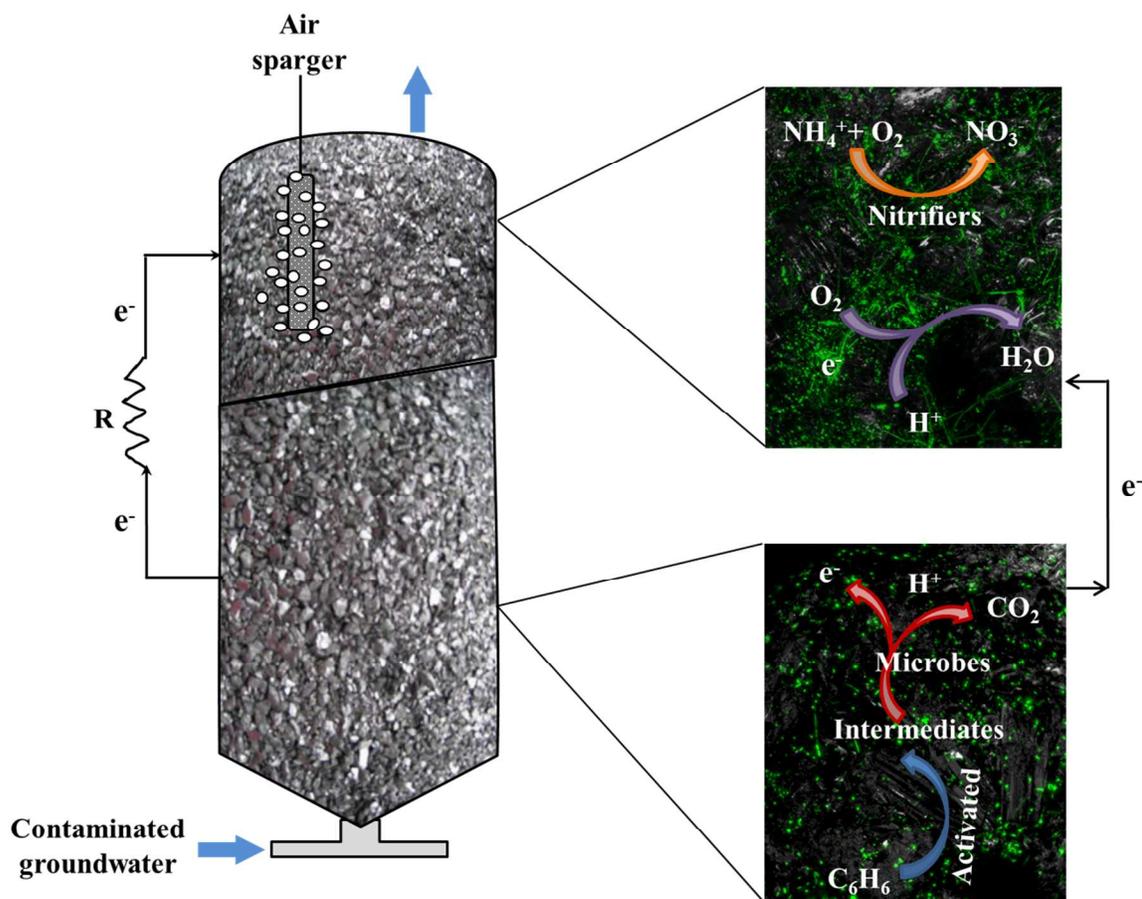
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A microbial fuel cell (MFC) was successfully applied for the treatment of benzene and ammonium co-contaminated groundwater.



1 Harvesting electricity from benzene and ammonium-contaminated
2 groundwater using a microbial fuel cell with an aerated cathode

3
4 Manman Wei,^{ab} Falk Harnisch,^c Carsten Vogt,^{*a} Jörg Ahlheim,^d Thomas R. Neu,^e Hans H.
5 Richnow^a

6
7 ^a Department of Isotope Biogeochemistry, Helmholtz Centre for Environmental Research -
8 UFZ, Permoserstraße 15, 04318 Leipzig, Germany

9 ^b Faculty of Natural Sciences, Hohenheim University, 70593 Stuttgart, Germany

10 ^c Department of Environmental Microbiology, Helmholtz Centre for Environmental
11 Research - UFZ, Permoserstraße 15, 04318 Leipzig, Germany

12 ^d Department of Groundwater Remediation, Helmholtz Centre for Environmental Research
13 - UFZ, Permoserstraße 15, 04318 Leipzig, Germany

14 ^e Department of River Ecology, Helmholtz Centre for Environmental Research - UFZ,
15 Brueckstrasse 3A, 39114, Magdeburg, Germany

16
17 * Author for correspondence:

18 Telephone: +49-341-235-1357; fax: +49-341-235-450822; email: carsten.vogt@ufz.de

19 **Abstract**

20 Groundwater contaminated with benzene and ammonium was continuously treated using a
21 microbial fuel cell (MFC) with an aerated cathode and a control without aeration at the
22 cathode. Benzene (~15 mg/L) was completely removed in the MFC of which 80%
23 disappeared already at the anoxic anode. Ammonium (~20 mg/L) was oxidized to nitrate at
24 the cathode, which was not directly linked to electricity generation. The maximum power
25 density was 316 mW/m³ NAC at a current density of 0.99 A/m³ normalized by the net anodic
26 compartment (NAC). Coulombic and energy efficiencies of 14% and 4% were obtained based
27 on the anodic benzene degradation. The control reactor failed to generate electricity, and can
28 be regarded as a mesocosm in which granular graphite was colonized by benzene degraders
29 with a lower benzene removal efficiency compared to the MFC. The dominance of
30 phylotypes affiliated to the *Chlorobiaceae*, *Rhodocyclaceae* and *Comamonadaceae* was
31 revealed by 16S rRNA illumina sequencing in the control and the MFC anode, presumably
32 associated with benzene degradation. Ammonium oxidation at the cathode of the MFC was
33 mainly carried out by phylotypes belonging to the *Nitrosomonadales* and *Nitrospirales*.
34 Compound specific isotope analysis (CSIA) indicated that benzene degradation was initially
35 activated by monohydroxylation with molecular oxygen. The intermediates of benzene
36 degradation pathway were subsequently oxidized accompanied by transferring electrons to
37 the anode, leading to current production. This study provided valuable insights into the
38 application of MFCs to treat groundwater contaminated with petroleum hydrocarbons (e.g.
39 benzene) and ammonium.

40

41 **Key words:** benzene degradation; ammonium oxidation; microbial fuel cell; electricity
42 recovery; compound-specific stable isotope analysis

43

44 **1. Introduction**

45 Remediation by microbial fuel cells (MFCs) is a promising technology for wastewater
46 treatment due to the combination of effective pollutants removal and electricity generation.¹

47 Recently, the practical application of MFCs has been widely developed to treat various
48 contaminated waters including e.g., municipal wastewater,² animal wastewater,^{3, 4} and
49 industrial wastewater.^{5, 6} In a few cases, the effective removal of petroleum constituents

50 from contaminated sediments or soil coupled to electricity generation have been also
51 achieved using MFC technology.^{7, 8} Many industrial activities such as oil refining and

52 chemical industry produce wastewater containing ammonium, sulfide and petroleum
53 hydrocarbons (e.g. BTEX, benzene, toluene, ethylbenzene, and xylene).^{9, 10} For example,

54 the Leuna site (Saxony-Anhalt, Germany) has been a center of chemical industry for about
55 100 years, leading to groundwater contaminated mainly by benzene and ammonium.¹¹

56 These pollutants have been reported to cause severe environmental and public health
57 damage.¹² A variety of remediation technologies, such as constructed wetlands,^{13, 14} soil

58 filter systems,⁹ and aerated treatment ponds with biofilm promoting mats,¹⁵ have been used
59 to treat groundwater contaminated with benzene and ammonium from the Leuna site.

60 However, oxygen was considered to be one of the limiting factors for efficient ammonium
61 removal in these remediation systems.^{13, 14, 16} Heterotrophic bacteria were reported to

62 potentially compete for oxygen and inorganic nitrogen with nitrifiers, possibly resulting in

63 low nitrification rates under oxygen-limited conditions.^{17, 18} In addition, the presence of
64 BTEX (benzene, toluene, ethylbenzene, and xylene isomers) and their metabolic
65 intermediates (e.g. phenol) were described to inhibit the nitrification process.¹⁹⁻²¹ The
66 inhibitory effect of benzene on the nitrification process, seemingly caused by the
67 competition between benzene degraders and nitrifiers, is an obstacle to simultaneously
68 remove benzene and ammonium using conventional treatment technologies.

69 It is already reported that the MFC technology has a practical potential for removing
70 benzene at the anode. Zhang et al.²² observed benzene degradation in contaminated marine
71 sediments by providing a graphite electrode as an electron acceptor, demonstrating the
72 potential of electrode-based systems for degradation of aromatic hydrocarbons in anoxic
73 environments. Luo et al.²³ operated a packing-type MFC and found that 600 mg/L benzene
74 was completely degraded within 24 h with simultaneous power generation when 1000
75 mg/L glucose was provided as the co-substrate. Simultaneous benzene biodegradation and
76 electricity production with potassium ferricyanide as electron acceptor in a MFC was also
77 reported by Wu et al.²⁴. Due to the fast reaction kinetics, the aromatic ring of benzene was
78 likely activated and cleaved by mono- and/or dioxygenases^{12, 25} in the studies of Luo et al.²³
79 and Wu et al.²⁴, indicating aerobic or microaerobic conditions; it has been reported
80 previously that benzene can be effectively degraded under oxygen-limited conditions.^{26, 27}
81 Nevertheless, the data indicate that benzene can be biodegraded under anoxic or
82 oxygen-limited conditions in a MFC system, although the mentioned studies were
83 performed under simulated and simplified conditions in the laboratory.

84 Ammonium removal in MFCs has been observed to occur at the cathode. He et al.²⁸

85 reported that ammonium can be removed mainly by partial nitrification with nitrite
86 production in a rotating-cathode MFC, although the process showed only a low coulombic
87 efficiency ($CE=0.34\%$). Subsequently, ammonium removal by simultaneous nitrification
88 and denitrification was achieved in a MFC coupled with a nitrifying bioreactor²⁹ or by
89 introducing additional oxygen into the cathode.³⁰⁻³³ Notably, a denitrifying liter-scale MFC
90 has been successfully used to enhance the total nitrogen removal in a municipal wastewater
91 treatment facility.³⁴

92 Therefore, MFCs are a good choice to sequentially remove benzene and ammonium due to
93 the presence of separated anodic and cathodic compartments. Considering the complexity
94 of contaminated groundwater, the feasibility of MFCs for contaminant removal is usually
95 limited by high internal resistance, pH buffering, inhibition effect between co-contaminants
96 and low efficiency of mixed culture biofilms on an electrode.³⁵ The practical application of
97 a MFC for the treatment of benzene and ammonium-contaminated groundwater has not
98 been studied yet. Hence, the objective of this study was to investigate whether a MFC can
99 be used to remediate real groundwater contaminated with benzene and ammonium while
100 simultaneously recovering energy. For that purpose, a MFC with an aerated cathode and a
101 control without aeration were compared for the performance of benzene and ammonium
102 removal as well as electricity generation. Additionally, the effect of hydraulic retention time
103 (HRT) on the performance of the MFC was investigated. To understand the electrochemical
104 processes occurring in the MFC, benzene and ammonium spiking as well as oxygen
105 interruption experiments were performed in batch mode. Additionally, the degradation
106 pathways and key players were elucidated by compound specific isotope analysis (CSIA)

107 and illumina sequencing.

108 **2. Experimental**

109 **2.1. Reactor setup**

110 Two reactors, MFC and control, were constructed as previously described by Rakoczy et al.
111 ³⁶ with some modifications (Fig. 1). Every reactor consisted of two cylindrical glass
112 compartments having each a diameter of 6 cm which were separated by a Nafion-117
113 cation exchange membrane (CEM) (QuinTech, Göppingen, Germany). The cathode
114 compartment (10 cm height, 330 mL total volume) was located on the top of the anode
115 compartment (22 cm height, 680 mL total volume). Both compartments were completely
116 filled with granular graphite of 1-6 mm diameter (Edelgraphit GmbH, Bonn, Germany)
117 which served as electrodes. After filling with granular graphite, the anodic and cathodic
118 compartment had a net liquid volume of 320 mL and 160 mL, respectively. A ring of
119 stainless steel (304 SS, 2 cm length, 6 cm diameter) was used as electron collector for
120 granular graphite at each compartment and interfaced to the external resistor. A loop
121 between the anode and cathode compartment was added (Fig. 1), allowing the anodic
122 effluent flowing directly into the cathodic compartment and thus eliminating proton mass
123 transfer loss and membrane pH gradients. An air sparger linked to an air pump (EHEIM
124 100, Stuttgart, Germany) was installed in the cathodic compartment of the MFC to provide
125 aeration. Continuous recirculation was generated using peristaltic pumps (Ismatec REGLO
126 Analog MS-2/6, Wertheim, Germany) at a rate of 60 mL/min in both compartments in
127 order to maintain well-mixed conditions. The reactors were completely covered with
128 aluminum foil to avoid light exposure, hence inhibiting growth of phototrophic organisms.

129 In the control reactor, pseudo-anode and pseudo-cathode were defined as two separate
130 compartments (corresponding to the anode and cathode compartments of the MFC), in
131 which no electrochemical reactions occurred.

132 **2.2. Start-up and reactor operation**

133 Both reactors were inoculated and continuously fed with contaminated groundwater from
134 Leuna; the composition is listed in Table 1. The groundwater was periodically taken from a
135 nearby groundwater well and stored in a 50 L tank which was kept at 0.5 bar N₂ pressure to
136 maintain anoxic conditions and was also connected to a cooling system to keep the
137 temperature at 10-12 °C.

138 The reactors were firstly operated in continuous treatment mode (day 0-134), using 1,000
139 Ω external resistance and a flow rate of 0.3 mL/min, resulting in a HRT of 27 h. These
140 parameters were only changed for polarization measurements (see 2.4.). Between day 134
141 and 160, the MFC was operated at the flow rates of 0.1, 0.5, 0.7, and 1.0 mL/min,
142 corresponding to HRTs of 80, 16, 12, and 8 hours, in order to investigate the effect of
143 varying HRTs. Once changing to a new flow rate, the MFC was running for one week to
144 reach a stable current output before further measurements. After 160 days, benzene and
145 ammonium spiking experiments were performed in the MFC. Briefly, 15, 30, and 50 mg/L
146 benzene were injected into the anode which was operated in a fed-batch mode, while the
147 cathode was running in a continuous flow mode. Subsequently, 20, 50 and 100 mg/L
148 ammonium were injected to the cathode in a fed-batch mode, while the anode was operated
149 in a continuous mode. Finally, through switching on or off the air pump, oxygen
150 interruption was performed twice in the continuous flow mode in order to prove whether

151 oxygen was the electron acceptor.

152 **2.3. Chemical analysis**

153 Benzene was analyzed using a gas chromatograph equipped with a flame ionization
154 detector (Varian CP-3800 GC, Palo Alto, CA) described elsewhere.³⁷ NH_4^+ -N, NO_2^- -N and
155 NO_3^- -N were analyzed colorimetrically as described before;³⁸ the detection limit was 10
156 μM for each compound. The pH was monitored by a pH meter (Knick, Berlin, Germany).
157 Dissolved oxygen (DO) was measured using an optical trace sensor system (PreSens sensor
158 spot PSt6 and FIBOX-3 minisensor oxygen meter, Regensburg, Germany) described in
159 more detail by Balck, et al.³⁹ The redox potential (Eh) was measured with a pH/mV/Temp
160 meter (Jenco Electronics 6230N, San Diego, USA). Samples for Fe^{2+} and total Fe
161 measurements were acidified to pH 2 directly after the sampling and analyzed
162 photometrically according to the guideline DIN 38405 D11. PO_4^{3-} , Cl^- , and SO_4^{2-} were
163 measured using the ion chromatograph (Dionex DX500, Idstein, Germany) following the
164 guideline EN ISO 10304-2, DIN 38405-19. Total organic carbon (TOC), inorganic carbon
165 (IC), 5-day biological oxygen demand (BOD), and chemical oxygen demand (COD) were
166 analyzed according to previously described methods.¹⁵

167 **2.4. Electrochemical measurements and calculations**

168 The Voltage (V) across a resistor (R) was recorded at 20 min intervals using a multimeter
169 (Metrix MTX 3282, Paris, France). Current (I) was calculated by Ohm's law ($I = V/R$) and
170 power (P) was calculated as $P = V \times I$. Current and power density were normalized by the net
171 anode compartment volume (NAC). The coulombic efficiency (CE) was calculated as the
172 ratio of the number of electrons recovered as charge versus the number of released electrons

173 by substrate removal. The energy efficiency (η) was defined as the ratio of power produced
174 by the cell to the heat of combustion of organic substrate, and was calculated as previously
175 described.⁴⁰ Polarization and power density curves were generated by varying the resistor
176 from 56,000 to 100 Ω (forward). Backward polarizations were also recorded by varying
177 resistance from 100 to 56,000 Ω ; the hysteresis was comparably low (Fig. S4). The reactor
178 was initially disconnected with the external resistance and was running under open circuit for
179 five hours to produce a stable open circuit potential (OCP). Data for each resistor adjustment
180 were recorded in intervals of 30 min or longer until the voltage change was less than 2 mV in
181 1 min. Individual anode and cathode potentials were measured using an Ag/AgCl reference
182 electrode (Sensortechnik SE11, Meinsberg, Germany) and assumed to be +0.197 V against
183 the standard hydrogen electrode (SHE).

184 **2.5. Compound-specific stable isotope analysis (CSIA)**

185 Benzene-containing influent groundwater and water from the pseudo-/anodic compartments
186 of two reactors were extracted with pentane. The carbon and hydrogen stable isotope
187 compositions were determined using a gas chromatograph-combustion-isotope ratio mass
188 spectrometer system (GC-IRMS). The detailed measurement and calculation were performed
189 as previously described.³⁶ The benzene degradation pathways were analyzed by comparing
190 measured isotope composition shifts with published isotope enrichment factors (ϵ) in a
191 two-dimensional isotope plot.^{37, 41}

192 **2.6. MiSeq illumina sequencing**

193 Total DNA was extracted from graphite granules and cation exchange membranes using the
194 FastDNA® spin Kit for soil (MP Biomedicals, Santa Ana, CA). PCR amplicons of bacterial

195 and archaeal 16S rRNA genes were generated using the *Bacteria*-universal primers 341F
196 (5'-TCCTACGGGNGGCWGCAG-3') and 785R (5'-TGACTACHVGGGTATCTAAKCC-3')
197 and *Archaea*-universal primers 340F (5'-TCCCTAYGGGGYGCASCAG-3') and 915R
198 (5'-TGTGCTCCCCCGCCAATTCCT-3'). Sequencing was performed using the illumina
199 MiSeq platform at a commercial laboratory (LGC Genomics GmbH, Berlin, Germany). Raw
200 data were processed using illumina CASAVA data analysis software and reads were
201 demultiplexed according to index sequence. Overlapping regions within paired-end reads
202 were then aligned to generate contigs. If a mismatch was discovered, the paired-end
203 sequences involved in the assembly were discarded. OUT picking and taxonomical
204 classification was performed at 97% identity level with Mothur 1.33. Sequence data were
205 deposited in the Sequence Read Archive (SRA) of NCBI database under the accession
206 number SRP044693.

207 **2.7. Confocal Laser Scanning Microscopy (CLSM)**

208 Biofilms attached to the granular graphite were examined with an upright confocal laser
209 scanning microscope equipped with a super continuum light source (Leica TCS SP5X,
210 Wetzlar, Germany). The system was controlled by the LAS AF software ver. 2.6.1. Samples
211 were mounted in a coverwell chamber with a spacer of 2 mm. Bacteria were stained by
212 nucleic acid specific fluorochrome SYBR Green. Settings for imaging were as followed: the
213 line of 494 nm was used for excitation; detection was at 489-499 nm (reflection) and 505-580
214 nm (SYBR Green). Data sets were recorded by using the 25x NA 0.95 (overview) and 63x
215 NA 0.9 (bacterial morphology) water immersible lenses.

216 **2.8. Statistical analysis**

217 Statistical analyses were performed using the SPSS 22.0 package (Chicago, IL, USA). The
218 normality and homogeneity were assessed with a Shapiro-Wilk W test and a Levene test,
219 respectively. Differences in benzene and ammonium removal efficiencies between the MFC
220 and control reactor were compared with one-way ANOVA tests. The Tukey's post-hoc test
221 was used to further evaluate the difference between the different flow rates when significant
222 differences were found. Differences were considered to be statistically significant if $p < 0.05$.
223 The linear relationship between removed pollutant loads and power generations were further
224 analyzed by a regression analysis.

225

226 **3. Results and discussion**

227 **3.1. Overall treatment performance during continuous operation**

228 **3.1.1. Overall performance of the MFC with an aerated cathode**

229 The MFC was continuously fed for around 130 days with contaminated groundwater
230 containing up to 15 mg/L benzene and 20 mg/L ammonium (Table 1), generating current and
231 achieving a constant benzene and NH_4^+ -N removal (Fig. 2 and 3). During the initial stage,
232 benzene and NH_4^+ -N removal as well as current output gradually increased, probably due to
233 the attachment and growth of ammonium- and benzene-metabolizing microorganisms from
234 contaminated groundwater. After approximately two months, benzene and NH_4^+ -N removal
235 became relatively stable, indicating that electrochemical active biofilms had been fully
236 developed, reflected also by a stable current generation (Fig. 3). In the anodic effluent of the
237 MFC, roughly 80% of benzene was removed whereas NH_4^+ -N concentrations decreased only
238 slightly (~5%). The remaining benzene (20%) was removed at the cathode, resulting in an

239 overall benzene removal efficiency of 100% in the cathodic effluent. Ammonium disappeared
240 completely in the cathodic effluent, showing finally 100% removal efficiency. In summary,
241 benzene was removed mainly (80%) in the anodic compartment and NH_4^+ -N was removed
242 solely in the cathodic compartment of the MFC. Sequential removal of benzene and
243 ammonium in the different compartments avoided putative negative effects of benzene and
244 benzene degraders on the nitrification process which have been previously reported.^{15, 16}
245 During steady stage, the MFC generated current between 200 and 250 μA and a current
246 density of 0.6~0.8 A/m^3 NAC was obtained (Fig. 3) when being operated at 1,000 Ω .
247 Comparing the results of this study with data for benzene and ammonium removal from other
248 remediation technologies, similar benzene removal rates but higher ammonium removal rates
249 were achieved in our MFC system, even only considering their best removal efficiencies
250 achieved during summer. For example, Seeger et al.¹³ reported 81%-99% benzene removal
251 and 40-50% NH_4^+ -N removal in a planted constructed wetland. A novel aerated treatment
252 pond exhibited approximately 100% benzene concentration reduction, whereas ammonium
253 concentrations decreased only slightly from around 59 mg/L at the inflow to 56 mg/L in the
254 outflow, indicating no significant NH_4^+ -N removal during continuous operation.¹⁵ Recently,
255 Rakoczy et al.³⁶ investigated the treatment of benzene and sulfide-contaminated groundwater
256 using a MFC over a period of 770 days and obtained a maximum current output of
257 approximately 250 μA in a continuous flow mode, which is comparable to the outcome of our
258 study.

259 3.1.2. Overall performance of the control reactor without aeration

260 In the control reactor, benzene removal in both pseudo-anodic and cathodic effluent gradually

261 increased during the initial microbial enrichment, and then achieved steady removal
262 efficiencies of 60% and 80%, respectively (Fig. 2). Compared to the MFC, less benzene (20%
263 lower) was removed in the control reactor ($p < 0.05$), indicating enhanced benzene degradation
264 in the anode compartment of the MFC. Ammonium was only slightly removed ($< 10\%$) in the
265 control, similar to the anode of the MFC. A low noise current ($< 5 \mu\text{A}$) was observed in the
266 control, hence current was not efficiently generated (Fig. S2). In MFCs, the electrochemical
267 potential difference of anode and cathode, depending on the redox potential of the electron
268 donor and the terminal electron acceptor, determines the possibility and extent of generated
269 current.⁴² As nearly the same anode and cathode potentials were observed in the control (data
270 not shown), no electromotive force was obtained and thus no current generated. Therefore,
271 the control reactor without aeration can be regarded as a benzene-degrading mesocosm in
272 which granular graphite was colonized by benzene degraders but not served as electron donor
273 or acceptor.

274 **3.2. Mechanisms of benzene and ammonium removal**

275 **3.2.1. Benzene removal mechanism**

276 CSIA was performed in order to identify the initial activation mechanisms of benzene
277 degradation in the MFC and the control reactor. The initial attack on thermodynamically very
278 stable benzene is critical for its degradation process. Combined carbon and hydrogen isotope
279 fractionation has been proved to be a powerful tool for the characterization of initial
280 metabolic reactions for benzene biodegradation.^{37, 41} Carbon and hydrogen isotope
281 fractionations of benzene were significantly higher in the control compared to the MFC (Fig.
282 S3). However, two dimensional plots of carbon versus hydrogen isotope fractionation were

283 similar for both MFC and control (Fig. 4), indicating that isotope fractionation was masked in
284 the MFC. The detected values matched with those indicative for benzene monooxygenation
285 to phenol catalyzed by a monooxygenase (Fig. 4). The produced phenol might be further
286 transformed into catechol by a second monooxygenation and also the other possible
287 intermediates after a possible *ortho* or *meta* ring cleavage of catechol.^{43, 44}
288 Monohydroxylation as benzene activation step was also identified in the anodic reaction of a
289 MFC for treating benzene and sulfide-contaminated groundwater.³⁶ However, we cannot
290 exclude that benzene was actually anaerobically activated and degraded by a mechanism
291 producing similar carbon and hydrogen isotope fractionation as observed for aerobic
292 monohydroxylation, as different anaerobic benzene activation mechanisms are currently
293 proposed: an anaerobic hydroxylation to phenol⁴⁵ or a carboxylation to benzoate.^{46,47} In any
294 case, the intermediates of the initial benzene activation steps were probably further oxidized
295 anaerobically and accelerate electricity generation by transferring the released electrons to the
296 anode. This is also supported by our electrochemical results (see 3.4.) and the fact besides
297 benzene, no other electron donors were provided by the contaminated groundwater.

298 3.2.2. Ammonium removal mechanism

299 At the cathode of the MFC, ammonium was mainly oxidized to nitrate by nitrification, as
300 indicated by an increase of NO_3^- -N (Fig. S1). In the control and the anode of the MFC, nitrite
301 and nitrate concentrations were extremely low (data not shown), showing that ammonium
302 removal by nitrification activity was negligible; the oxygen concentrations were obviously
303 too low for nitrification, as previously observed in an aerated treatment pond with biofilm
304 promoting mats at the Leuna field site.¹⁶ The small ammonium losses in the control reactor

305 and the anode compartment of the MFC might be due to physical-chemical processes, e.g.
306 adsorption or volatilization.^{4, 48}

307 **3.3. Effect of the flow rate on the performance of the MFC**

308 To determine the effect of HRTs, different flow rates were used to study benzene and $\text{NH}_4^+\text{-N}$
309 removal performance as well as power generation in the MFC. As shown in Fig. 5A, benzene
310 removal at the anode decreased from 80% at a flow rate of 0.3 mL/min (27 h HRT) to 40% at
311 a flow rate of 1.0 mL/min (8 h HRT). At a flow rate of 0.1 mL/min (80 h HRT), benzene was
312 completely removed already at the anode. However, 100% benzene removal efficiency was
313 always obtained in the final cathodic effluent at the five studied HRTs. At the lower flow rate
314 (0.1 and 0.3 mL/min), ammonium was completely removed (Fig. 5A). At flow rates higher
315 than 0.3 mL/min, the removal efficiencies decreased gradually to around 70% at a flow rate
316 of 1.0 mL/min, implying that nitrification was limited at the shorter HRT.

317 The maximum power density of 316 mW/m^3 NAC was achieved at a current of 0.99 A/m^3 ,
318 when the MFC was operated at the flow rate of 0.3 mL/min (Fig. 5B). As shown by the
319 polarization and power density curves, lower performances were obtained at higher or lower
320 flow rates. The electrode potentials as a function of current density were also examined at
321 different flow rates (Fig. 5C). As expected from the removal data, it was observed that the
322 cathodic potentials were not affected by the different flow rates. The anodic OCP values
323 shown at zero current density were also similar under the different flow rates, demonstrating
324 an excellent stability of the system. The electrode polarization showed very different profiles
325 for the anodic potentials at the different flow rates. Therefore, the difference of power density
326 at the different flow rates was a result of the polarization behavior of the anodic potentials,

327 indicating that anodic benzene oxidation was rate-limiting and thus determined electricity
328 generation. In order to confirm this finding, subsequent benzene injection experiments were
329 performed and also approved that benzene served as the main anodic electron donor in the
330 MFC (Fig. 6A).

331 Coulombic efficiencies of $28 \pm 4.7\%$, $14 \pm 3.4\%$, $10 \pm 1.2\%$, $8 \pm 1.4\%$ and $7 \pm 0.9\%$ were
332 obtained at the increased flow rates of 0.1, 0.3, 0.5, 0.7, and 1.0 mL/min, respectively;
333 analogously, energy efficiencies of $8 \pm 1.3\%$, $4 \pm 0.9\%$, $3 \pm 0.3\%$, $2 \pm 0.3\%$ and $1.2 \pm 0.2\%$
334 were achieved. The lower coulombic and energy efficiencies at higher flow rates are
335 indicative for incomplete benzene degradation, eventually resulting in a lower number of
336 electrons transferred to the anode. The data also indicate that a substantial number of
337 electrons were generally not transported to the anode, probably caused by the use of
338 penetrating oxygen or alternative substances as electron acceptors (e.g. carbonate leading to
339 methanogenesis or organic or inorganic metabolites upon fermentation processes). Notably,
340 the MFC with aerated cathode was more efficient with regard to benzene degradation than the
341 ferricyanide-based MFC described recently by Wu et al.²⁴, which produced a power density
342 of 2.1 mW/m^2 and showed a coulombic efficiency of 3.3%, at slightly lower benzene loadings
343 and slightly higher benzene degradation rates. In subsequent experiment, a flow rate of 0.3
344 mL/min was used due to the maximum power density and high pollutant removal
345 performance achieved at this date.

346 **3.4. Detection of electrochemical reaction in the MFC**

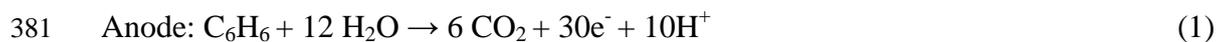
347 Although the complete degradation of benzene can theoretically release 30 electrons,²⁴
348 current generation strongly depends on whether electrons from benzene oxidation can be

349 efficiently transferred to the anode. In our study, when 15 mg/L benzene was injected into the
350 anode, the current rapidly increased from around 240 to 350 μA within 3 h; subsequently, the
351 current gradually decreased corresponding to decreasing benzene concentrations (Fig. 6A).
352 Injecting higher benzene concentrations of 30 and 50 mg/L generated higher current maxima
353 of 415 and 450 μA , respectively. A linear correlation was obtained between the current and
354 the concentration of benzene at the anode (Fig. 6D), demonstrating that benzene oxidation
355 was the main anodic reaction, and that the released electrons were efficiently delivered to the
356 anode. In contrast, increasing ammonium concentrations by injection of 20, 50, 100 mg/L
357 $\text{NH}_4^+\text{-N}$ did not affect the current output (Fig. 6B), indicating that cathodic ammonium
358 oxidation did not limit electricity generation. Subsequently, the effects of varying oxygen
359 concentrations at the cathode were investigated (Fig. 6C). When the supply of oxygen at the
360 cathode was stopped, the DO concentration gradually decreased from about 6 to 1 mg/L, and
361 the current also decreased from about 300 to 60 μA . The current generation immediately
362 increased when oxygen was again supplied and the DO concentration increased. Hence, the
363 DO concentration was coupled to current generation, suggesting that oxygen was the main
364 terminal electron acceptor at the cathode.

365 As $\text{NH}_4^+\text{-N}$ can release maximally eight electrons via oxidation to nitrate, ammonium can
366 theoretically serve as an anodic electron donor.²⁸ However, ammonium as a direct anodic fuel
367 for electricity generation has not been demonstrated experimentally yet. In our study,
368 ammonium was completely oxidized to nitrate at the cathode by nitrifying microorganisms
369 (Fig. S1). Our results demonstrate that the released electrons were not involved in
370 electrochemical reactions; probably, the electrons were directly taken by nitrifiers, explaining

371 why ammonium consumption was not directly connected to electricity generation in the MFC.
372 You et al.⁴⁹ showed that additional protons produced from ammonium oxidation can reduce
373 the ohmic resistance and maintain the pH balance in the absence of a phosphate buffer, which
374 can contribute to the electricity generation process. Recently, it was also reported that the
375 oxygen reduction relied on the nitrification activity at the biocathode, at least to some
376 extent.⁵⁰ In this study, the contaminated groundwater from the Leuna site probably had a
377 good buffer capacity, so that biological nitrification was not directly linked with current
378 generation.

379 Based on these results, the anodic and cathodic reactions occurred in the MFC are described
380 by the following equations, with the assumption of complete oxidation of benzene.



384 Coulombic and energy efficiencies at the different flow rate were calculated according to the
385 electrochemical reactions shown in equations 1-3.

386 3.5. Analysis of microbial communities in the MFC and the control

387 3.5.1 Bacterial community analysis

388 Confocal laser scanning microscopy images demonstrated that significant biofilms were
389 formed around the granular graphite in both MFC and control (Fig. S5). The compositions of
390 bacterial communities colonizing the pseudo-/anode, CEM and pseudo-/cathode are shown in
391 Figure 7A. The anode of the MFC and the control reactor were mainly colonized by bacterial
392 phylotypes belonging to the *Chlorobiales*, *Rhodocyclales*, and *Burkholderiales*. The family

393 *Chlorobiaceae* in the *Chlorobiales* accounted for ~15% and 41% of the pseudo-/anodic
394 bacterial communities in the control and MFC, respectively. Although *Chlorobiales* are
395 known as anaerobic photoautotrophs, several recent studies suggest that they are able to
396 perform anaerobic dark respiration by the breakdown of organic substrates when sulfide is
397 not used as electron donor.^{51, 52} *Chlorobiales* were also identified as the dominant phylotypes
398 in benzene degrading enrichment cultures under nitrate-reducing conditions^{41, 53} indicating
399 that *Chlorobiales* can participate in degradation of benzene or its intermediate metabolites.
400 Large percentages of *Rhodocyclaceae* of 43% and 21% (belonging to *Rhodocyclales*) were
401 observed at the pseudo-/anode of the control and MFC respectively, indicating as well a role
402 of these phylotypes upon benzene degradation; correspondingly, phylotypes of this family
403 were shown to be associated with anaerobic benzene degradation either by DNA-stable
404 isotope probing with ¹³C-labelled benzene^{53, 54} or by phylogenetic analysis of
405 benzene-degrading enrichment cultures.⁵⁵ The *Comamonadaceae* within the order of
406 *Burkholderiales*, accounting for 5% and 6% of bacterial communities at the pseudo-/anode of
407 the control and MFC, were also shown to be involved in benzene degradation.^{53, 56} Overall,
408 the dominance of these potential benzene degraders implies that those were actually related to
409 benzene degradation in both MFC and control. The whole control reactor, including
410 pseudo-anode, CEM and pseudo-cathode, were colonized by similar bacteria with slightly
411 different abundances, again supporting the view that it is a homogenous mesocosm.
412 Different from the bacterial community at the anode, the dominant bacteria orders at the
413 cathode of the MFC were *Nitrospirales* (18%), *Burkholderiales* (15%), *Rhodocyclales* (7%),
414 *Nitrosomonadales* (4.5%), and *Rhizobiales* (4%). Obviously, the dominant families of

415 *Comamonadaceae* and *Rhodocyclaceae* affiliated to *Burkholderiales* and *Rhodocyclales* were
416 able to degrade the residual benzene from the anode.⁵³ Phylotypes affiliated to
417 *Nitrosomonadales* and *Nitrospirales* belong to the genera of *Nitrosovibrio* and *Nitrospira*,
418 which presented the dominant ammonia-oxidizing and nitrite-oxidizing bacteria
419 respectively.^{57, 58} They were also identified as dominant nitrifying bacteria in wastewater
420 treatment plants and MFC systems for nitrogen removal.^{31, 59} The data suggests that
421 ammonium was firstly oxidized to nitrite by *Nitrosovibrio*, then nitrite was further oxidized to
422 nitrate by *Nitrospira*, leading to a high rate of nitrification at the cathode of the MFC.

423 3.5.2 Archaeal community analysis

424 The compositions of archaeal communities were shown in Fig. 7B. The archaeal communities
425 were dominated by phylotypes belonging to the acetoclastic *Methanosarcinales* (64-76%),
426 and hydrogenotrophic *Methanomicrobiales* (1.5-8%) in the three parts of control and the
427 anode of the MFC (Fig. 7B). Methanogens can compete for substrates with electrochemically
428 active microorganisms and hence reduce electron recovery.⁶⁰ It was however shown that
429 electrochemically active microorganisms can outcompete acetoclastic methanogens for
430 organic substrates in MFCs, eliminating electron consumption by methanogens.^{60, 61} As the
431 acetoclastic *Methanosarcinales* were the predominant archaea at the MFC anode, electron
432 loss by methanogenesis was probably rather minor. Notably, *Methanosarcinales* and
433 *Methanomicrobiales* have been also identified as syntrophic methanogens in anaerobic
434 benzene-degrading enrichment cultures.^{62, 63}

435 Different from the archaeal communities in the control and the anode of the MFC, the CEM
436 and cathode of the MFC was dominated by unclassified *Euyarchaeota* (46%), unclassified

437 archaea (39%) and *Methanosarcinales* (12%). Possibly, archaea from anoxic groundwater
438 were not able to grow at the aerobic cathode of the MFC due to the completely different
439 physicochemical conditions.

440 Due to the complexity of the anode biofilm, it was very difficult to elucidate concrete
441 electrochemical mechanisms at the anode, e.g. identifying the microorganisms actually
442 transferring electrons to the anode. However, various dominant bacterial phylotypes were
443 identified in the MFC, suggesting that the electrons were transferred to the anode after the
444 initial activation reaction by monohydroxylation rather by a network of microorganisms,
445 using different metabolites of benzene degradation as substrates, than by a single organism.
446 These processes might be syntrophic; such syntrophic processes have been described to
447 govern anaerobic benzene degradation in both benzene-degrading enrichment cultures and in
448 situ benzene-degrading columns.⁶³

449 **4. Conclusions**

450 This study demonstrated the principal feasibility of treating benzene and ammonium
451 contaminated groundwater by a MFC equipped with an aerated cathode. Benzene was
452 initially activated by enzymatic monohydroxylation at the oxygen-limited anode; the further
453 anaerobic oxidation of the intermediate metabolites released electrons, which were
454 transferred to the anode and eventually captured by oxygen, driving oxygen reduction and
455 accelerating electricity production. Nitrification took place at the aerated cathode of the MFC
456 and was catalyzed by nitrifiers; the process was not directly linked to electricity generation.
457 Although it is promising that nitrification occurred in high rate at the cathode of the MFC, the
458 accumulated nitrate still needs to be removed by an additional treatment reactor. Further

459 research is of particular interest in order i) to promote simultaneous benzene and ammonium
460 removal using an anoxic cathode MFC, e.g. by addition of nitrite to accelerate anaerobic
461 ammonium oxidation, avoiding the need of aeration, or the set-up of an additional
462 denitrification reactor, or ii) to develop and test a scaling-up application under field
463 conditions.

464

465 **Acknowledgement**

466 This work was financially supported by the Helmholtz Centre for Environmental
467 Research-UFZ in the scope of the SAFIRA II Research Project (Revitalization of
468 Contaminated Land and Groundwater at Megasites, subproject “Compartment
469 Transfer-CoTra”) and within the Research Programme “Terrestrial Environment”.

470 Falk Harnisch acknowledges the support by the BMBF (Research Award “Next generation
471 biotechnological processes - Biotechnology 2020+”) and the Helmholtz-Association
472 (Young Investigators Group).

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593

594 **Figure and table legends**

595 Table 1: Physico-chemical properties of contaminated groundwater used in this study

596

597 Figure 1: Schematic view (A) and picture (B) of the MFC and the control used in this
598 study.

599

600 Figure 2: Benzene and $\text{NH}_4^+\text{-N}$ removal in the MFC (A) and the control (B) during
601 continuous treatment of contaminated groundwater.

602 *represents significant higher removal efficiency ($p < 0.05$) in the MFC compared to the
603 control at the same sampling time.

604

605 Figure 3: Current generation in the MFC during continuous treatment of contaminated
606 groundwater.

607

608 Figure 4: Two-dimensional isotope plot of $\Delta\delta^{13}\text{C}$ versus $\Delta\delta^2\text{H}$ values of benzene measured
609 at the anode of the MFC (red symbol) and pseudo-anode of the control (black symbol)
610 during continuous treatment. Values of $\Delta\delta$ were calculated by subtracting the measured
611 isotopic value from the initial isotopic value determined at the influent.

612

613 Figure 5: Effect of flow rate on benzene and ammonium removal (A), power generation (B),
614 and electrode potentials (C) in the MFC. B: polarization curve (solid symbol) and power
615 density curve (open symbol); C: anode potential (solid symbol) and cathode potential (open

616 symbol). Different letters (a, b, c, d) placed above the bars indicate a significant difference
617 in removal efficiencies at the different flow rates.

618

619 Figure 6: Effect of benzene and ammonium additions, and oxygen interruption on current
620 generation in the MFC. (A): benzene injection of 15 mg/L (a1), 30 mg/L (a2), and 50 mg/L
621 (a3); (B): NH_4^+ -N injection of 20 mg N/L (b1), 50 mg N/L (b2) and 100 mg N/L (b3); (C):
622 Interruption of O_2 supply at the cathode; (D): correlation between current generation and
623 benzene concentration at the anode.

624

625 Figure 7: Phylogenetic distribution of 16S rRNA genes based on the order of bacteria (A)
626 and archaea (B) detected in the MFC and control. Bacterial orders with a read abundance <1%
627 in all of the six samples were pooled in 'others'. MFCA: MFC anode; MFCM: MFC CEM;
628 MFCC: MFC cathode; ConA: Control pseudo-anode; ConM: Control CEM; ConC: Control
629 pseudo-cathode.

630

631 Wei et al., Table 1

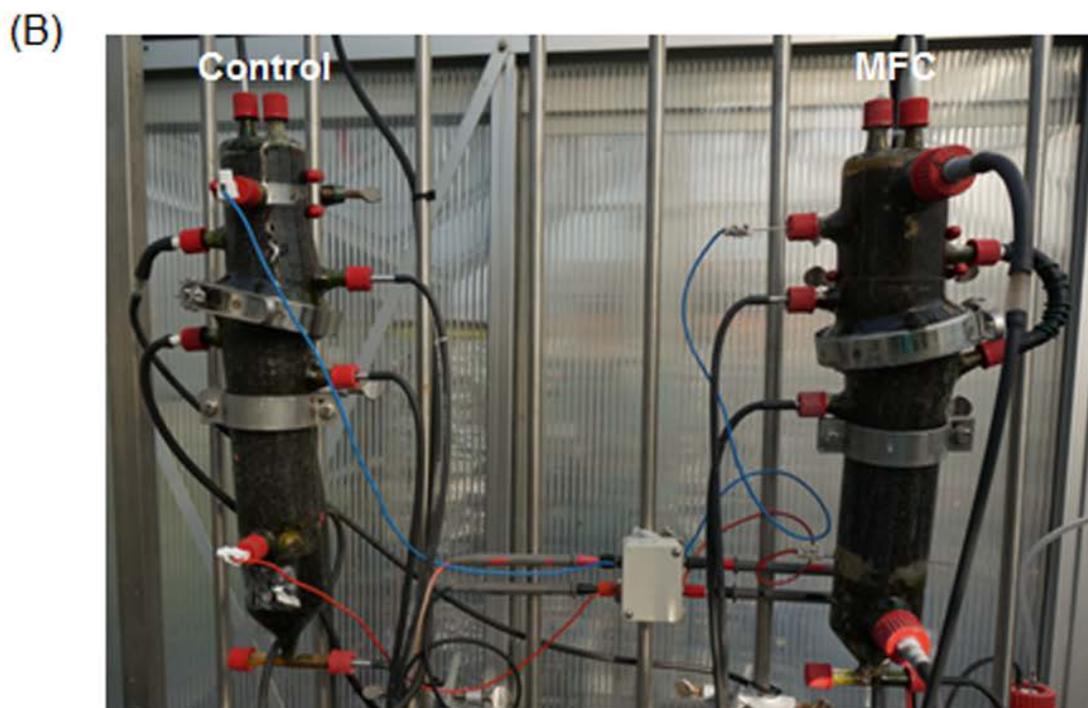
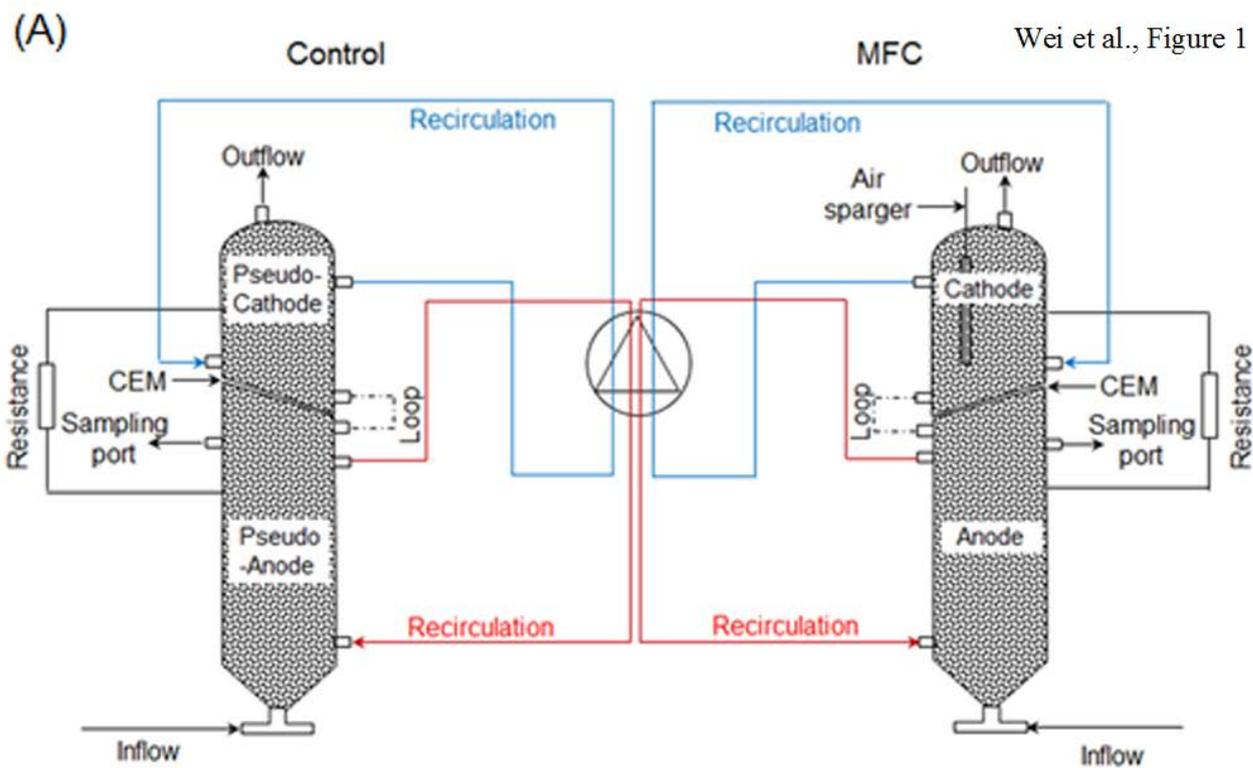
632

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Parameters	Means \pm Stdv
NH ₄ ⁺ (mg/L)	21.3 \pm 1.6
NO ₂ ⁻ (mg/L)	<0.02
NO ₃ ⁻ (mg/L)	<0.1
SO ₄ ²⁻ (mg/L)	2.4 \pm 1.5
Total Fe (mg/L)	5.6 \pm 0.6
Fe ²⁺ (mg/L)	5.4 \pm 0.8
PO ₄ ³⁻ (mg/L)	2.3 \pm 0.1
Cl ⁻ (mg/L)	95.2 \pm 5.6
Benzene (mg/L)	15.2 \pm 0.6
MTBE (mg/L)	1.1 \pm 0.1
COD (mg/L)	83.8 \pm 2.6
BOD ₅ (mg/L)	36.8 \pm 4.7
TOC (mg/L)	29.0 \pm 7.2
IC (mg/L)	275.3 \pm 27.6
DO (mg/L)	0.05 \pm 0.01
Eh (mV)	-167 \pm 23
pH	7.5 \pm 0.3

634 Note: Contaminated groundwater was collected during September-December 2013 from the Leuna site.

635



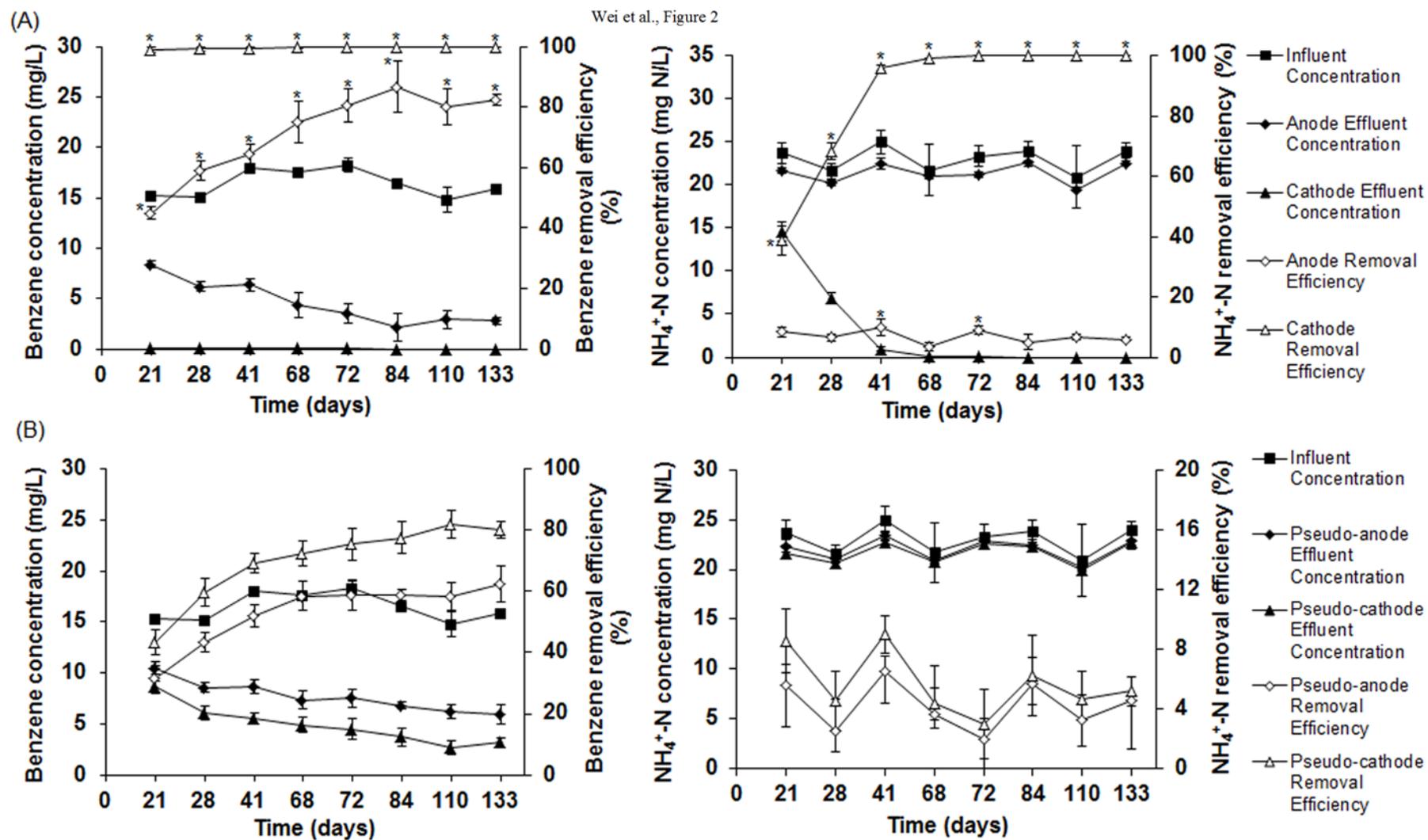
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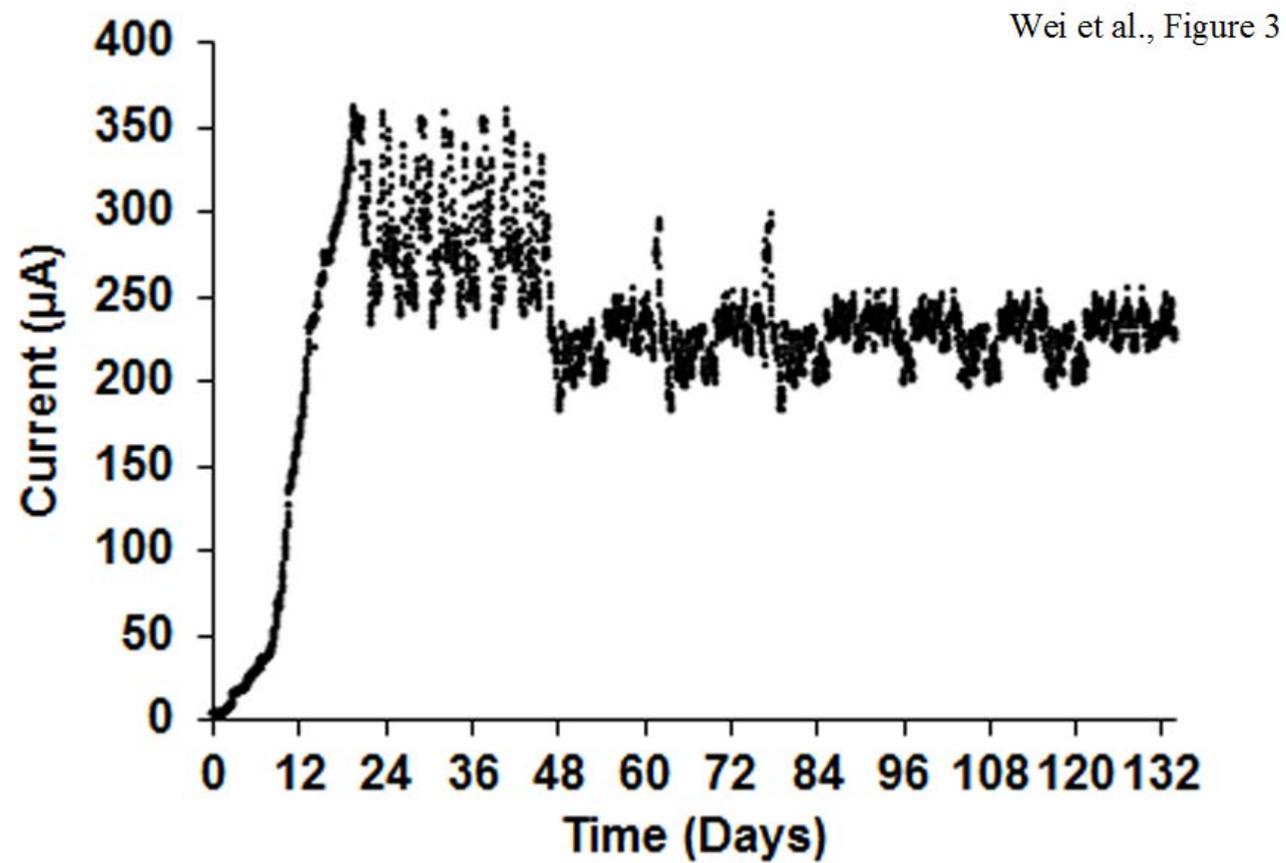
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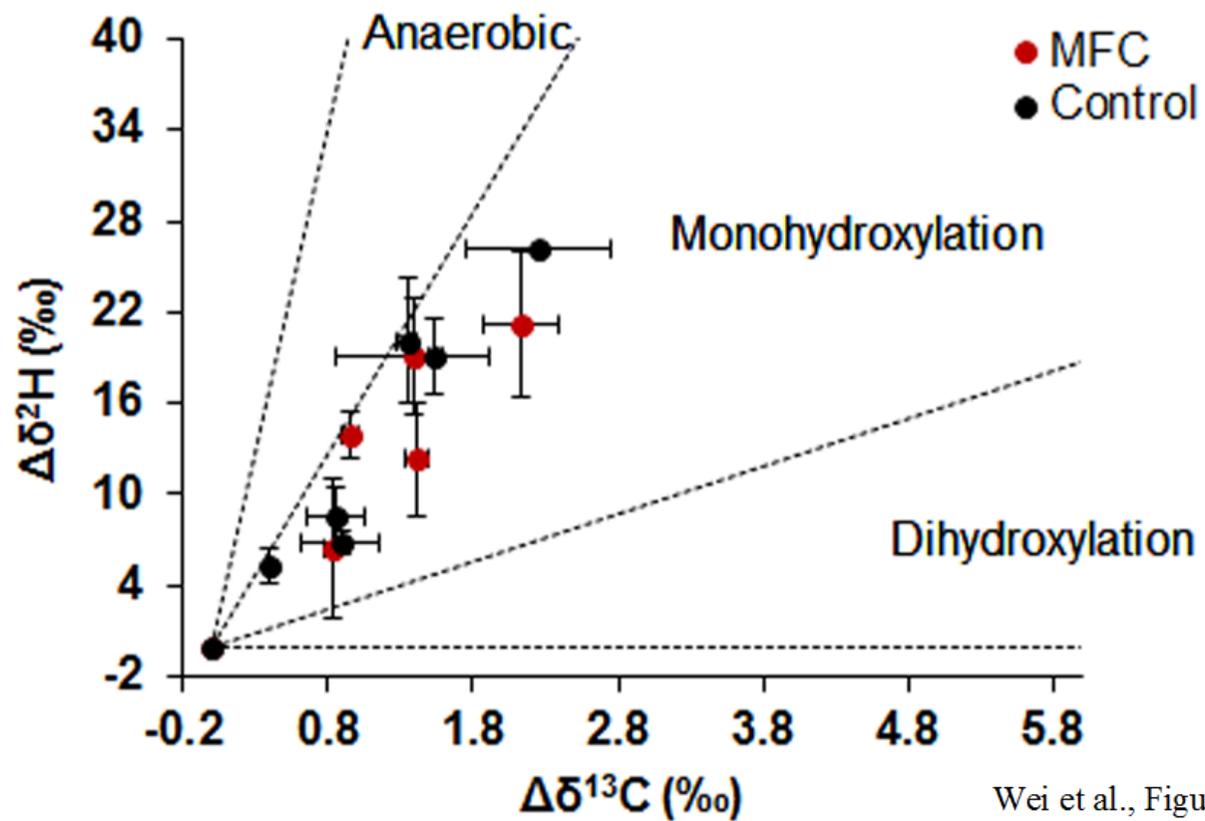
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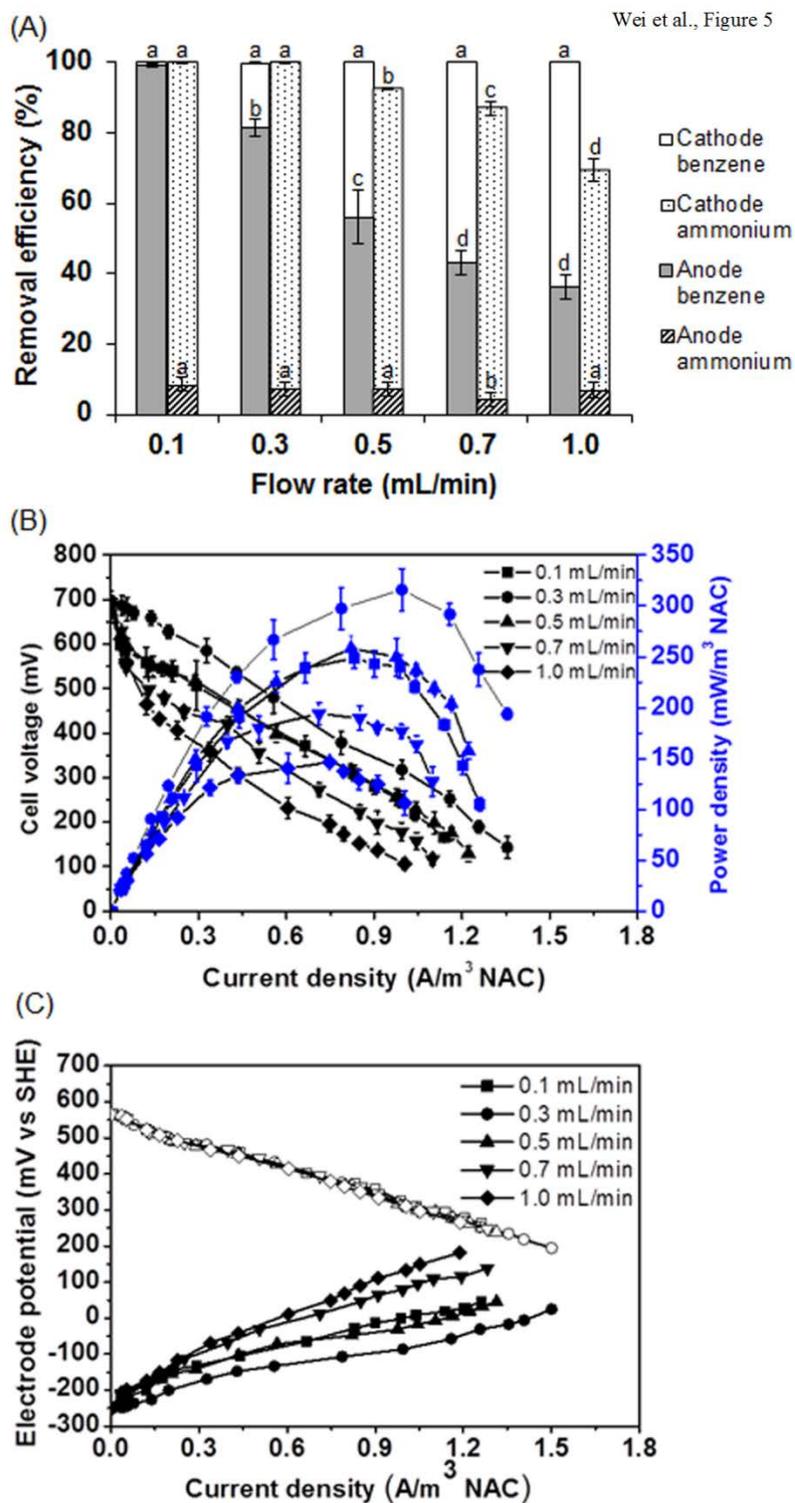
Wei et al., Figure 4

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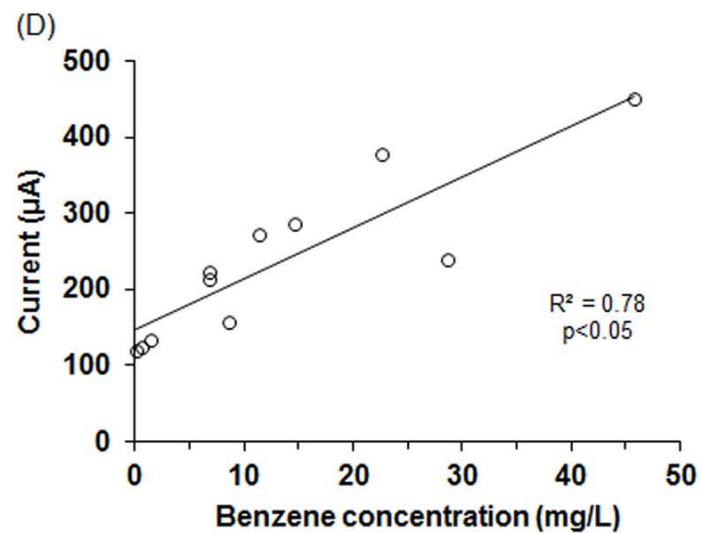
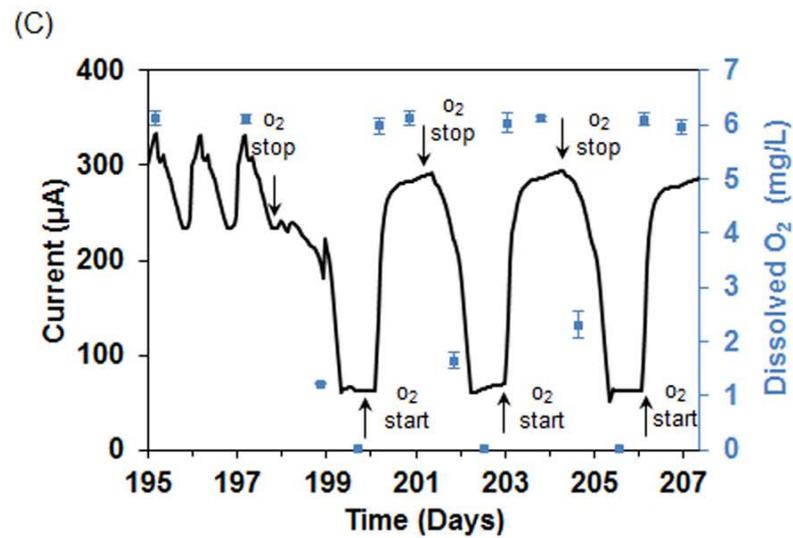
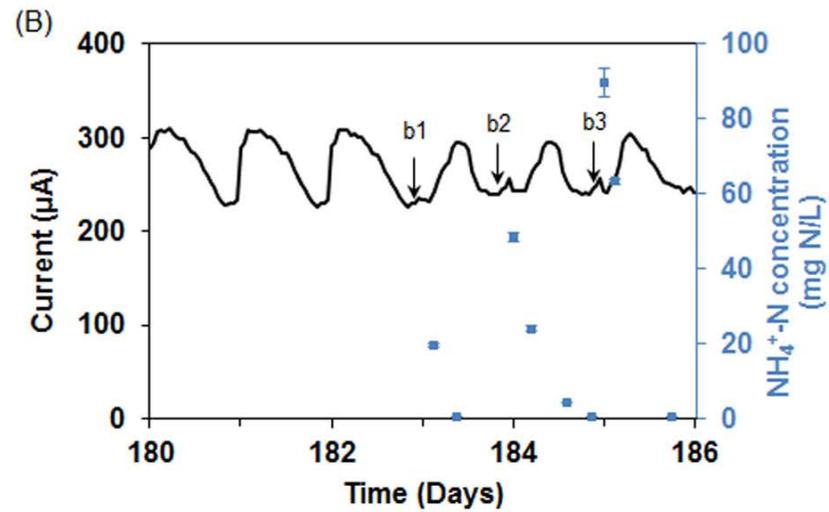
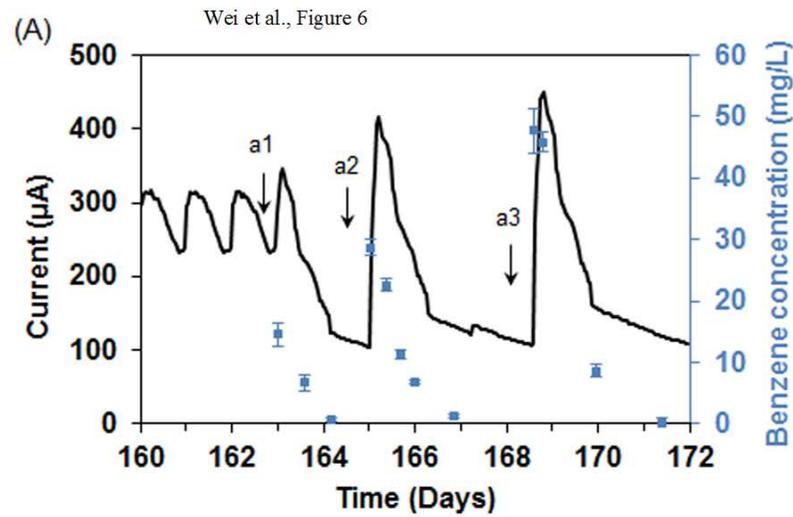
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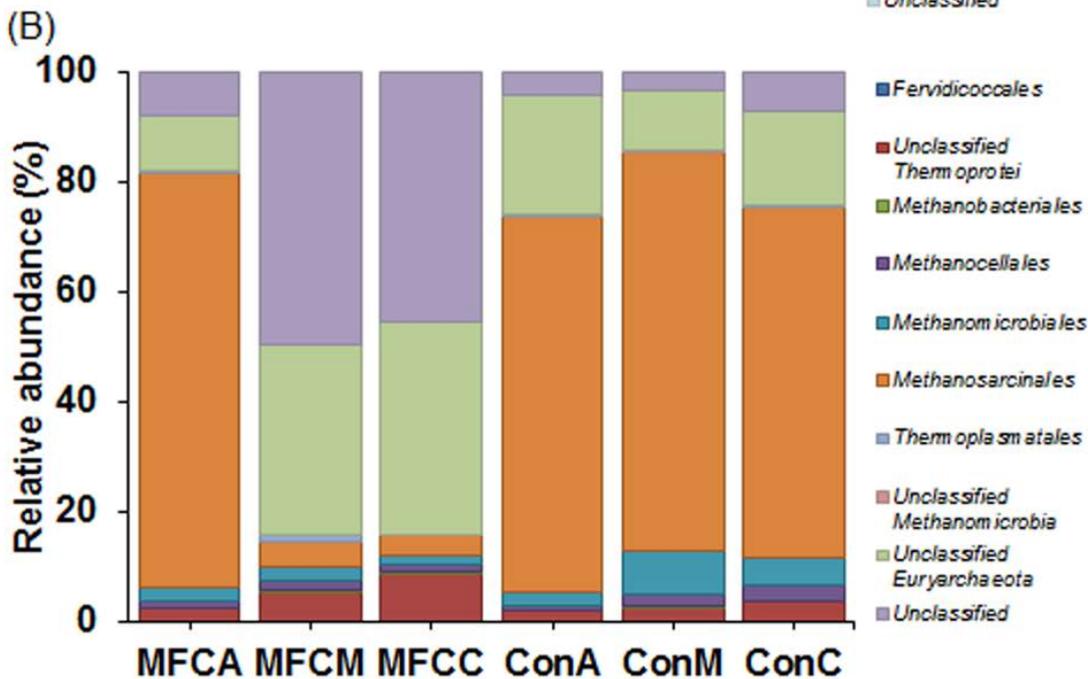
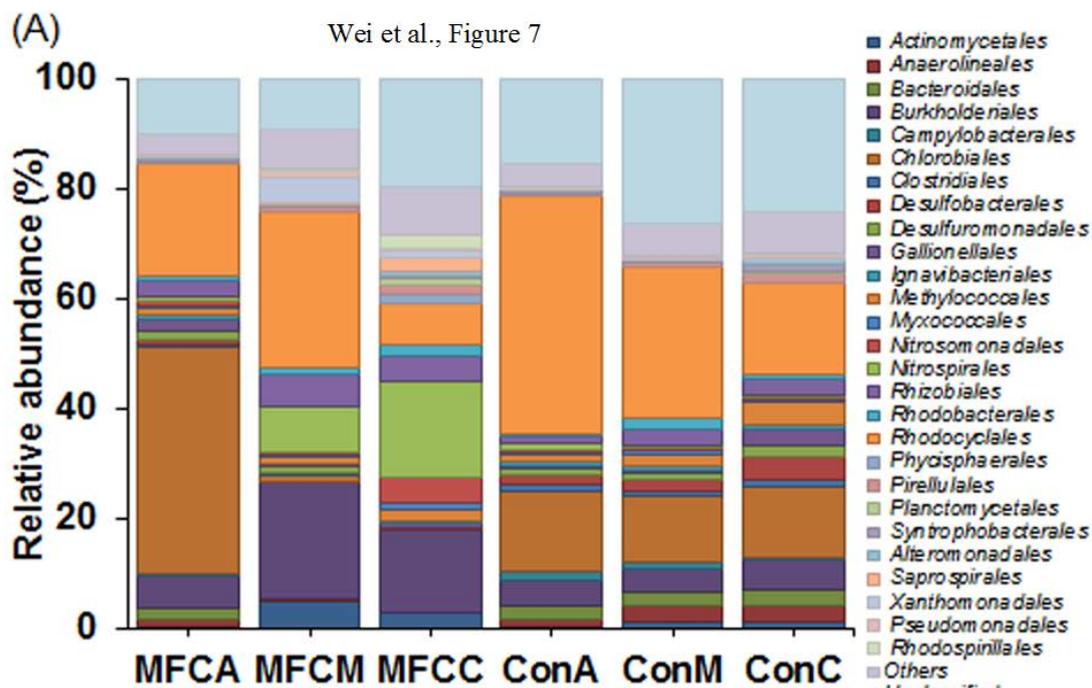
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