



Ex situ conservation of *Pinus koraiensis* can preserve genetic diversity but homogenizes population structure

Yuewei Tong^{a,b}, Walter Durka^{c,d}, Wangming Zhou^a, Li Zhou^a, Dapao Yu^a, Limin Dai^{a,*}

^a CAS Key Laboratory of Forest Ecology and Management, Institute of Applied Ecology, Shenyang 110016, China

^b University of Chinese Academy of Sciences, Beijing 100049, China

^c Department of Community Ecology (BZF), Helmholtz Centre for Environmental Research-UFZ, Halle 06120, Germany

^d German Centre for Integrative Biodiversity Research (iDiv), Halle-Jena-Leipzig, Deutscher Platz 5e, Leipzig D-04103, Germany



ARTICLE INFO

Keywords:

Pinus koraiensis
Ex situ conservation
Clonal seed orchard
Genetic diversity
Population structure

ABSTRACT

Pinus koraiensis is a conifer species of ecological and economic importance in northeast China that has been excessively exploited in recent years. A clonal seed orchard (CSO) was established including potentially genetically differentiated provenances from the whole distribution area, we applied nine simple sequence repeat (SSR) makers to study the genetic diversity and population structure of six maternal populations as well as their progeny populations. The results showed a high genetic diversity in both maternal and progeny populations, with the average expected heterozygosity of 0.617 and 0.632, respectively. The level of genetic diversity in the progeny populations was slightly higher than that in the maternal populations, and almost all diversity descriptors were correlated between maternal and progeny populations, indicating that the CSO could preserve the established species' gene pool. An overall low level of genetic differentiation of *P. koraiensis* ($F_{ST} = 0.029$ and 0.025 for maternal and progeny populations) was found. The six maternal populations clustered into two groups by Bayesian cluster analysis with the two northernmost populations comprising one group and the other four populations as another genetically differentiated group, potentially indicating regional adaptation. The northern group had significantly higher levels of within population diversity. Population structure was not evident anymore in the progeny populations, suggesting that mating among differentiated provenances within the CSO homogenized the different gene pools, although differences in diversity were still maintained. These results will inform efforts for the conservation and management of *P. koraiensis* and provide guidance for future studies of population genetics and breeding programs.

1. Introduction

Forests are considered one of the most complex terrestrial ecosystems due to the high level of biodiversity, and play an important role in the function of forest ecosystems (Geburek and Konrad, 2008; Litkowiec et al., 2018). Genetic diversity, widely recognized as a key component of biodiversity (Millennium Ecosystem Assessment, 2005), provides the essential basis for the adaptation and resilience of species to environmental stress and change (Potter et al., 2017). As long-lived, immobile, and often widespread life forms, forest tree species need high levels of genetic diversity to adapt to changing environmental influences (Loo et al., 2014; Potter et al., 2017). Unfortunately, the genetic resources of forest trees are threatened by an increased use of timber and reduction of the area of forestland (Rajora and Mosseler, 2001). For example, deforestation, which directly eliminates numbers and sizes of the locally adapted populations is an immediate threat to forest tree

genetic diversity (Ledig, 1992). Therefore, maintaining or enhancing genetic diversity in forest tree species is an urgent global necessity for genetic resource conservation (Ratnam et al., 2014; Holliday et al., 2017). This may require the integration of both *in situ* (on site) and *ex situ* (off site) conservation strategies, especially in high-value species and those with small and vulnerable populations (Dawson et al., 2013; Pritchard et al., 2014; Schwartz et al., 2017).

Ex situ conservation strategies preserve plants or plant germplasm away from their areas of natural occurrence (Given, 1994). A clonal seed orchard (CSO), one of these strategies, often consist of trees selected from a single provenance, however, also multiple provenances may be combined, thus broadening the genetic basis (Zobel et al., 1958; Giertych, 1975). The main goals of CSOs are to achieve genetic gain in economically relevant traits due to initial selection, and to maintain genetic variation by enhancing panmixia (Chaloupkova et al., 2019). Because CSOs represent the link between breeding programs and

* Corresponding author.

E-mail address: lmdai@iae.ac.cn (L. Dai).

reforestation activities, inadequate genetic diversity in seed crops may result in accumulation of inbreeding, which directly reduces potential productivity (Kang et al., 2001; Kang et al., 2005; Sonstebo et al., 2018; Chaloupkova et al., 2019). Generally, the genetic diversity of seeds produced in a CSO is determined by that of provenances and parent trees which, however, was unknown during the establishment. Moreover, the diversity of seeds will depend on planting design (Yuan et al., 2016) and the pollen dispersal distance. Although most forest temperate forest trees are wind pollinated, pollen dispersal distance is limited and may be non-random, e.g. due to preferential wind direction (Feng et al., 2010). If provenances are genetically differentiated and seeds are produced by random mating in a seed orchard, higher genetic diversity is expected for seeds compared with the average source provenance. However, while mixing genetically differentiated populations will increase genetic diversity, potentially with hybrid vigor, this is connected with a potential risk of outbreeding depression (hybrid breakdown) by disrupting coadapted gene complexes if provenances are adapted to different environmental conditions (Johansen-Morris and Latta, 2006). Therefore, as CSOs are a tool for breeding of numerous tree species around the world, studying the genetic diversity of maternal provenances and offspring produced in a CSO can provide a reliable basis for evaluating the potential risks in the management of seed orchards and plantations (El-Kassaby and Ritland, 1996; Nielsen and Hansen, 2012; Sonstebo et al., 2018).

Pinus koraiensis Siebold et Zucc. (Korean pine, Pinaceae), has its centre of distribution in Northeastern China, and ranges from South Korea into Russia and Japan (Aizawa et al., 2012). It is one of the most ecologically and economically important conifer species in the mountainous area of Northeastern China (Ma et al., 1992). The species is a dominant forest tree species in its habitat and produces high-quality timber and edible pine nuts (Chen et al., 2010; Aizawa et al., 2012). In recent decades, the populations of natural broad-leaved Korean pine mixed forests have been sharply reduced (Yu et al., 2011), and the species has been listed as nationally endangered in China (<http://www.plant.csdb.cn/endangeredplants>). Thus, the protection of *P. koraiensis* has become urgent, especially for the genetic resources. There are several hypotheses about the origin of Korean pine populations in continental Asia ranging from a single glacial refugium, substantiated by the genetically depauperate plastid genomes that lack variation (Aizawa et al., 2012) to the hypothesis of several continental *in situ* relic populations (Potenko and Velikov, 1998). For the nuclear genome of *P. koraiensis*, a gradient of genetic variation decreasing from South (Korea) to North (Russia) (Kim et al., 2005) indicates however that populations are not homogenous and likely display genetic structure.

In the 1980s, a breeding program to protect the genetic resources of *P. koraiensis* was started in China and a clonal seed orchard (CSO) was established including provenances from the whole distribution area. However, the genetic diversity and population structure of the different provenances had not been assessed as well as their progeny. Therefore, our objectives of this study were, based on six populations of *P. koraiensis* in the CSO, to (i) estimate genetic diversity and population structure of *P. koraiensis* provenances, and (ii) to compare the genetic variation of maternal populations and offspring populations that represent first generation hybrids produced in the CSO.

2. Materials and methods

2.1. Study site and plant material

We studied the Hongwei Seed Orchard which is located in Lushuihe town, northeast China (42° 28' 30" N 127° 47' 00" E, 760 ~ 800 m a.s.l., 2.7°C mean annual temperature, 871 mm mean annual precipitation) and was established in 1989. This clonal seed orchard (CSO) was established from cuttings of 660 *Pinus koraiensis* mother trees which were selected by phenotypic evaluation (volume, height, diameter at breast height, bole straightness, and branching habits) from 17 provenance

Table 1

Names, population ID, and original site coordinates of *Pinus koraiensis* populations analysed.

Population	ID	Latitude (°)	Longitude (°)	Annual Mean Temperature (°C)	Annual Mean Precipitation (mm)
Lushuihe	LSH	42.53	127.80	4.12	783.68
Huangnihe	HNH	43.55	128.01	3.76	635.42
Dahailin	DHL	44.52	128.86	1.14	619.72
Dongfanghong	DFH	46.58	133.58	3.17	648.99
Dailin	DL	47.18	128.85	0.30	574.59
Fenglin	FL	48.13	129.19	0.04	596.15

areas in Northeast China, thus covering large parts of the species' range. Each provenance has 20 to 60 mother trees (clones) and each clone is represented by 10 to 20 ramets. The CSO has a total area of 16.33 ha with rectangular planting scheme with 3 m distance between trees in 17 plots. All ramets were distributed using random block designs, and avoiding close vicinity of ramets from the same clone.

We selected six provenances, henceforward called "populations" (Table 1), representing a S-N cline with the two northernmost populations (DL, FL) separated from the southern populations by the river Heilong Jiang (Amur). The minimum and maximum geographic distances among populations ranged from 100 km to 650 km. For each population, we collected both needles (henceforward referred to as "mothers") and seeds from one random ramet per clone for each of 25 clones (Table 1). Seeds were germinated and then the needles of one random seedling per clone were sampled. All samples were stored at -80 °C before DNA extraction.

2.2. Genotyping

Total genomic DNA was extracted from leaf tissue using the DNAsecure Plant Kit DP320 (TIANGEN) according to the manufacturer's instructions. The quality and concentration of genomic DNA were assessed by agarose gel electrophoresis. Samples were genotyped at eight dinucleotide and one trinucleotide SSR markers (P5, P6, P29, P45, P51, P52, P62, P63 and P79; see supplement Table S1) (Yu et al., 2012; Jia et al., 2017). PCRs for all loci were performed separately in a 20 µL reaction volume containing 40 ng of genomic DNA, 0.3 µM concentrations of each primer (the forward primers were labelled with a fluorescent dye (FAM or HEX)), 0.1 mM dNTP (TransGen Biotech, Beijing, China), 2 µL 10× buffer and 1.0 U Taq DNA Polymerase (TransGen Biotech, Beijing, China). Cycling parameters were initial denaturation at 94 °C for 5 min; followed by 35 cycles of denaturation at 94 °C for 30 s, annealing for 35 s (for annealing temperature, see supplement Table S1), extension at 72 °C for 40 s; and final extension at 72 °C for 3 min. The amplification products were separated on the ABI3730XL DNA analyzer (Applied Biosystems) using GS-500 LIZ (Applied Biosystems) as internal size standard. Allele binning and genotyping were performed with GeneMarker® software (SoftGenetics LLC, State College, PA, USA).

2.3. Data analysis

2.3.1. Genetic diversity indices

We calculated a set of population genetic diversity estimators using GeneALEX 6.5.1 (Peakall and Smouse, 2012). The observed (A) and effective (A_e) number of alleles, the observed (H_o) and expected (H_e) heterozygosity and the Shannon diversity index (I) were all calculated for each locus, over all loci, and for each population. In addition, we calculated allelic richness (A_r) and the inbreeding coefficient F_{IS} (Weir and Cockerham, 1984) in FSTAT 2.9.3 (Goudet, 1995).

2.3.2. Population structure analysis

Genetic population structure was assessed and quantified in several steps. First, Principal coordinates analysis (PCoA) was used to illustrate genetic distances between individuals. Next, analysis of molecular variance (AMOVA) (1000 permutations) based on the degree of genetic divergence among populations was performed using GeneAEx 6.5.1 (Peakall and Smouse, 2012). In addition, genetic structure was assessed with a Bayesian clustering algorithm using STRUCTURE version 2.3.4 (Hubisz et al., 2009). The range of possible number of clusters (K) tested was from 1 to 13 for all populations and 7 for mother and seed populations (number of populations plus 1), for which a series of ten independent runs was performed with a burn-in period of 10 000 steps followed by 100 000 MCMC replicates. We used provenance as location prior (LOCPRIOR) (Hubisz et al., 2009) under the admixture model. Note that using a location prior will enable detection of weak genetic differentiation among locations only when differentiation is actually present (Manual of Structure software). The *ad hoc* statistic ΔK (Evanno et al., 2010) together with inspecting the log-likelihood values was used to identify the most likely number of clusters. The ten runs from the most probable K were averaged by applying a FullSearch algorithm by CLUMPP 1.1.2 (Jakobsson and Rosenberg, 2007).

2.3.3. Statistical analysis

Mean values of genetic diversity descriptors were compared among sample types or populations by T-test in R (R Core Team, 2013). We tested for clinal variation of genetic diversity descriptors with latitude with linear models in R.

3. Results

3.1. Genetic diversity

The nine SSR loci assayed were all polymorphic in *Pinus koraiensis*. Overall, there were a total of 73 different alleles with an average of $A = 6.0$ alleles per locus. Among loci, locus P51 revealed the highest level of polymorphism across all diversity descriptors, while P5 and P29 showed lowest values (Tab. S2). Among loci, observed heterozygosity (H_o) varied from 0.457 to 0.890 with an average of 0.685, expected heterozygosity (H_e) was just slightly lower than H_o , and ranged from 0.422 (P29) to 0.853 (P51), with an average of 0.624 (Tab. S2). A_r ranged from 2.5 (P5) to 11.3 (P51), and the average A_r was 6.7 (Table S2). The F_{IS} showed negative and significant values in loci P6, P45, P51, P52 and P63, and the overall F_{IS} calculated across 9 loci was -0.109 (Table S2, $P < 0.05$), indicating the existence of heterozygosity excess.

Genetic diversity parameters and inbreeding coefficients at population level are given in Table 2. In mothers, for all the diversity descriptors, the two northernmost populations had the highest values. When grouped into northern and southern populations, the northern group had significantly higher values for A , I , H_e , uH_e and A_r (Table 2). When testing for clinal variation with latitude, A , I and A_r showed increasing values towards North (Table 2). The F_{IS} showed significantly negative values in four populations indicating slight departure from Hardy-Weinberg equilibrium with heterozygosity excess. Almost all diversity descriptors were correlated between mothers and seeds (H_e , uH_e , I : $p < 0.05$; A , A_r : $p < 0.1$). For all diversity descriptors values were higher in seeds than in mothers, significantly so in A , I , and A_r . The spatial patterns found for mothers were similarly found, however with reduced strength, in seeds as the two northern populations had significantly higher values than the southern populations for I , H_o , H_e and uH_e , but only marginally significantly higher values for A and A_r . Clinal variation with latitude was only marginally significant for the same parameters as in mothers (A , I , A_r). The inbreeding coefficient was significantly negative in four seed populations, but F_{IS} was not correlated between mothers and seeds.

3.2. Genetic differentiation and population structure

The non-hierarchical AMOVA (Table 3) showed that 4.25%, 3.49% and 2.27% of genetic variation was among populations ($P < 0.001$) for all, mother, and seed populations, respectively, with the rest residing within populations. Pairwise F_{ST} values were low (0.009 to 0.035), but often significant (Tables 4 and S3). For the mother populations, nine out of 15 pairwise F_{ST} values were significant, while for the seed populations six were significant (Table 4). Pairwise F_{ST} values were significant mostly for comparisons including the two northern populations.

Furthermore, a hierarchical AMOVA analysis revealed significant differentiation (2.56%) between groups of mother and seeds ($P < 0.001$), and a similar amount of variation (2.80%) among populations within groups ($P < 0.001$) (Table 3). Similarly, a hierarchical AMOVA was carried out according to the results of the genetic diversity analysis which indicated that the two northern and four southern populations could be considered as two geographic groups. For the mother populations, genetic variation between groups was 2.37% ($P < 0.001$) and variation among populations within groups was 2.19% ($P < 0.001$, Table 3). However, in the seed populations, the variation between groups was not significant and 2.19% resided among populations within groups (Table 3).

The subsequent STRUCTURE analysis provided additional information on number of gene pools and the level of genomic mixture and admixture among populations. For mothers, the most probable division with the strongest support in terms of log-likelihood values was at $K = 2$ (Fig. S1), with southern populations LSH and HNH representing pure cluster I (blue), northern populations DL and FL pure cluster II (red) and intermediate populations DHL and DFH being admixed (Fig. 1). For seeds, only a single cluster was found, with L(K) being highest at $K = 1$ and no geographic separation of populations whatsoever (Fig. S2, S3). Analyzing all populations, the ΔK value reached a maximum for $K = 2$ with mothers (blue) and seed (red) populations representing the two clusters (Fig. 2, S4).

4. Discussion

4.1. Genetic diversity

The main task of seed orchards is to produce a large number of high genetic quality seeds for reforestation without reducing its genetic diversity (Chaloupkova et al., 2019). In this paper, we analysed genetic diversity and population structure of *Pinus koraiensis* in a CSO, to assess the degree of genetic differentiation among provenances and the changes in genetic variation between mothers and seeds.

Firstly, the average expected heterozygosity of the mother populations of *Pinus koraiensis* in this study was 0.617, indicating that genetic diversity was high. This result is similar to most current research revealing high levels of genetic diversity in *Pinus* species when assessed with SSR markers. For example, H_e was 0.782 for *P. thunbergii* (Iwaizumi et al., 2018), 0.601 in *P. koraiensis* (Feng et al., 2010), 0.55 in *P. sylvestris* (Bernhardsson et al., 2016), 0.586 also in *P. sylvestris* (Toth et al., 2017), 0.531 in *P. strobus* (Mandak et al., 2013), and 0.428 in *P. yunnanensis* (Xu et al., 2016). Meanwhile, we found a pattern of genetic diversity for *P. koraiensis* in this study, that is, the genetic diversity of the northern populations was higher than that of the southern populations. This is in contrast to results of Kim (Kim et al., 2005) who found a large-scale decline of variation from south (North Korea) to north (Russia). Thus, large scale patterns are not necessarily true on a more regional scale. This pattern went also undetected in Kim et al. (2005), potentially due to biased sample sizes or choice of genetic markers or of populations. Anyway, the results indicate that against a general cline of northward reduction of genetic diversity, populations in Xiaoxinganling mountains maintained an increased level of diversity.

Secondly, the level of genetic diversity of the seed populations in

Table 2

Genetic diversity parameters for the mother and seed populations of six *P. koraiensis* analyzed at nine microsatellite loci: mean number of alleles (*A*), mean number of effective alleles (*A_e*), Shannon's index (*I*), observed heterozygosity (*H_o*), expected heterozygosity (*H_e*), unbiased expected heterozygosity (*uH_e*), allelic richness (*A_r*) and inbreeding coefficient (*F_{IS}*). In addition, correlation coefficients and p-value of correlation with latitude and mean values for mother and seed populations, and p-value of an ANOVA testing for differences between northern and southern populations (N vs. S) as well as mothers and seeds.

Population	Type	<i>A</i>	<i>A_e</i>	<i>I</i>	<i>H_o</i>	<i>H_e</i>	<i>uH_e</i>	<i>A_r</i>	<i>F_{IS}</i>	
LSH	Mother	5.2	3.3	1.212	0.649	0.603	0.616	5.222	-0.055	
HNH	Mother	5.2	3.2	1.210	0.680	0.610	0.622	5.222	-0.095 *	
DHL	Mother	5.3	3.3	1.220	0.693	0.612	0.624	5.333	-0.113 *	
DFH	Mother	5.3	3.7	1.235	0.600	0.593	0.605	5.333	0.009	
DL	Mother	5.8	3.6	1.333	0.720	0.638	0.652	5.778	-0.108 *	
FL	Mother	5.8	3.4	1.319	0.716	0.643	0.656	5.778	-0.092 *	
Correlation mother vs. latitude		r	0.854	0.677	0.852	0.304	0.628	0.626	0.866	0.014
		p	0.030	0.139	0.031	0.558	0.181	0.184	0.026	0.979
ANOVA N vs. S		p	< 0.001	0.520	< 0.001	0.114	0.006	0.005	< 0.001	0.422
LSH	Seed	6.3	3.6	1.305	0.707	0.622	0.635	6.333	-0.116 *	
HNH	Seed	6.4	3.3	1.279	0.640	0.623	0.636	6.444	-0.007	
DHL	Seed	6.1	3.8	1.320	0.644	0.633	0.646	6.111	0.002	
DFH	Seed	6.6	3.6	1.303	0.667	0.612	0.625	6.556	-0.069 *	
DL	Seed	6.8	3.6	1.366	0.724	0.646	0.660	6.778	-0.100 *	
FL	Seed	6.7	3.5	1.364	0.782	0.657	0.671	6.667	-0.170 *	
Correlation seed vs. latitude		r	0.792	0.06	0.772	0.609	0.616	0.623	0.732	0.509
		p	0.061	0.910	0.072	0.199	0.193	0.187	0.098	0.302
ANOVA N vs. S		p	0.065	0.882	0.008	0.038	0.016	0.014	0.069	0.135
Mean	Mother	5.4	3.4	1.255	0.676	0.617	0.629	5.444	-0.076	
Mean	Seed	6.5	3.6	1.323	0.694	0.632	0.645	6.481	-0.077	
Correlation mother-seed		r	0.773	0.273	0.937	0.440	0.978	0.977	0.744	-0.142
		p	0.071	0.600	0.006	0.382	< 0.001	< 0.001	0.090	0.789
ANOVA mothers vs. seeds		p	< 0.001	0.178	0.031	0.557	0.170	0.165	< 0.001	0.976

Table 3

Non-hierarchical AMOVA for all, mother and seed populations of *P. koraiensis*, and a hierarchical AMOVA with mothers and seeds as groups for all populations, as well as with the two northern and four southern populations as two geographic groups for mother populations and seed populations; significant values are indicated with * ($P < 0.05$).

Populations	Source of variation	df	Sum of square	Variance components	Percentage of variation	
All	Among Pops	11	122.680	0.235	4.25%*	
	Within Pops	288	1522.160	5.285	95.75%*	
	Total	299	1644.840	5.520	100%	
Mother	Among Pops	5	49.707	0.189	3.49%*	
	Within Pops	144	752.000	5.222	96.51%*	
	Total	149	801.707	5.411	100%	
Seed	Among Pops	5	42.293	0.124	2.27%*	
	Within Pops	144	770.160	5.348	98.73%*	
	Total	149	812.453	5.473	100%	
All groups: mothers vs. seeds	Between groups	1	30.680	0.143	2.56%*	
	Among Pops	10	92.000	0.157	2.80%*	
	Within Pops	288	1522.160	5.285	94.63%*	
	Total	299	1644.840	5.585	100.00%	
	Mother groups: N vs. S	Between groups	1	16.857	0.130	2.37%*
		Among pops	4	32.850	0.120	2.19%*
		Within pops	144	752.000	5.222	95.44%*
		Total	149	801.707	5.471	100.00%
		Seed groups: N vs. S	Between groups	1	8.883	0.008
	Among pops		4	33.410	0.120	2.19%*
Within pops	144		770.160	5.348	97.66%*	
Total	149		812.453	5.476	100.00%	

our study was actually higher than that of the mother populations. The number of alleles increased on average across loci and populations by one allele from 5.4 to 6.5 and *H_e* increased from 0.617 to 0.632. Thus, the progeny populations not only maintain but actually surpass the maternal level of genetic diversity. This confirms the hypothesis that

the *ex situ* conservation can preserve the species' gene pool, provided that a diverse wild gene pool is encompassed. In contrast, both in natural forests and orchards, usually a constant genetic diversity was found across generations. Roberds & Conkle found that there was no significant change allele frequencies between the maternal and progeny populations of *Pinus taeda* (Roberds and Conkle, 1984). Furthermore, for *Pinus massoniana* (Ai et al., 2006) and *Pseudotsuga menziesii* (Prat and Arnal, 1994) in CSOs is was shown that genetic diversity is maintained across generations. The underlying causes for the observed increase of diversity in our study likely is first the rich gene pool of 17 source populations representing the whole species range in China, second the efficient pollen dispersal of *P. koraiensis* up to 60 m (Feng et al., 2010) and third the planting strategy in this CSO that maximizes the probability of crossing between accessions. Therefore, the genetic diversity of the seed populations of *P. koraiensis* can not only be maintained, but increased in this CSO. Still, the geographic pattern of increased variation in northern populations is maintained, although with reduced effect in the seed populations.

4.2. Genetic structure and genetic differentiation

The population structure is important for establishing the appropriate scale and subunits for conservation management (Moritz, 1999). It is affected by mutation, gene flow, natural selection and genetic drift, and thus is related to the evolutionary history and biological characteristics of the species (Loveless and Hamrick, 1984; Schneller and Liebste, 2007). Population structure is manifested mainly by genetic differentiation among populations (Frankham et al., 2002). As expected for conifer tree species, we found an overall low level of population differentiation ($F_{ST} = 0.029$ and 0.025 for mother and seed populations), which is consistent with the findings of most species with outcrossing breeding system and wind pollination (Belletti et al., 2012; Iwaizumi et al., 2013; Mandak et al., 2013; Durka et al., 2017). *Pinus koraiensis* is wind pollinated and outcrossing (Feng et al., 2010), and its seeds are dispersed mainly by birds and rodents (Miyaki, 1987). Such pollination and seed dispersal syndromes are beneficial to increase gene flow within and among populations (Nybom, 2004; Petit et al., 2005), and consequently to reduce genetic differentiation among populations.

Table 4

Pairwise F_{ST} for all populations of *P. koraiensis*, significant values are indicated with * ($P < 0.05$).

mothers						seeds						
LSH	HNH	DHL	DFH	DL	FL	LSH	HNH	DHL	DFH	DL	FL	
0.000											LSH	
0.011	0.000										HNH	
0.015*	0.012	0.000									DHL	
0.011	0.014	0.013	0.000								DFH	
0.033*	0.023*	0.018*	0.032*	0.000							DL	
0.017*	0.014*	0.012	0.016*	0.019*	0.000						FL	
0.016	0.019	0.019	0.023	0.031	0.018	0.000					LSH	
0.029	0.021	0.014	0.027	0.020	0.023	0.021*	0.000				HNH	
0.020	0.019	0.011	0.018	0.019	0.018	0.014	0.013	0.000			DHL	
0.014	0.020	0.018	0.016	0.035	0.019	0.011	0.028*	0.015	0.000		DFH	
0.021	0.021	0.017	0.025	0.020	0.016	0.009	0.017*	0.012	0.013	0.000	DL	
0.029	0.029	0.023	0.028	0.027	0.019	0.014	0.014*	0.016*	0.023*	0.010	0.000	FL

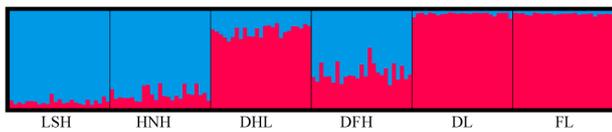


Fig. 1. Results of the STRUCTURE analysis of mother populations at $K = 2$. Each individual is represented by a single vertical bar, which is partitioned among gene pools. Colours represent genetic clusters and the coloured segments show the individual's estimated ancestry proportion.

According to the genetic diversity parameters and the STRUCTURE analysis, the six mother populations can be divided into two groups, with the two northern populations in the Xiaoxinganling Mountains as one group displaying higher genetic diversity and the southern four populations in the Changbai Mountains and its adjacent highlands as another group with lower genetic diversity. These groups are separated by the large Heilong Jiang river valley which may have restricted gene flow between these geographic regions. While populations in the Xiaoxinganling Mountains have always been part of natural forests with low human disturbance, the southern region has long been subjected to strong human disturbance. In addition, the regions also differ climatically which may have resulted in adaptive divergence among these regions. It is important to mention that one of the nine genetic markers used are EST-SSR (P79), i.e. it is part of the expressed genome. Thus unlike random genomic SSRs, which are typically considered to be neutral (Shi et al., 2011; Vieira et al., 2016), the EST-SSR markers are more likely to be under selection. Thus our data suggest that adaptive divergence may be at least partially responsible for genetic divergence between the two gene pools.

Both adaptive divergence and differences in human intervention are unlikely to be responsible for differences in genetic variation between the two population groups. However, the maternal population genetic structure and level of diversity also reflects the glacial refugia and postglacial expansion processes to a certain extent. A comprehensive study of the phylogeography of *Pinus koraiensis* is lacking and both a multiple refugia hypothesis (Potenko and Velikov, 1998) and a single-refugium hypothesis with a latitudinal cline of genetic variation (Kim et al., 2005; Aizawa et al., 2012) have been stated. Our data suggest that a scenario of a single refugium with later northward expansion resulting in constant decrease of genetic variation from south to north likely is too simple, as has been found in many Chinese forest species

(Shi et al., 2014). More complex scenarios have to be considered and the high diversity detected in the Xiaoxinganling Mountains may indicate a role as a local refuge.

However, for the seed populations in the CSO, the genetic population structure of the six populations became similar and clustered into just one group. This occurred due to the efficient mixing of gene pools among mother populations in the CSO. Thus, while seeds produced in the CSO have efficiently mixed gene pools resulting in increased diversity, but still maintaining regional differences in diversity, the primary geographically organized population differentiation, which may be in part due to regional adaptation, was wiped out.

4.3. Implications for forest management and conservation of *Pinus koraiensis*

Pinus koraiensis is a second-class protected precious timber species in China, it has formed rich genetic variation by long-term geographical isolation and natural selection (Feng et al., 2010; Park et al., 2017). At present, natural populations of *P. koraiensis* are declining due to habitat fragmentation and reduced population size entailing also long-term risks for its genetic resources. On the basis of the genetic diversity and population structure of *P. koraiensis* in this study, several strategies for more effectively protecting this species may be proposed: (1) The population genetics of the Korean pine should be studied and integrated with analysis techniques of molecular biology (e.g. transcriptome), physiology (e.g. rates of photosynthesis, water use efficiency, frost resistance) (Hofmann et al., 2015), and ecology to evaluate the ability to survive future changes and planning conservation strategies. (2) Strengthening the protection of existing natural forests by conserving their habitats, prohibiting timber harvest and installing sustainable management regimes are essential actions for maintaining the genetic diversity of this species. (3) In this paper, we have shown that the genetic diversity of the progeny population in the CSO is slightly increased compared with the maternal population, providing evidence that the gene pool of *P. koraiensis* as present in the CSO can be preserved by the *ex situ* conservation. Therefore, strengthening the *ex situ* conservation efforts and combining them with *in situ* conservation, can not only protects the *P. koraiensis* resources, but also enriches its genetic diversity. Lastly, however, our finding that *P. koraiensis* in China is divided into a Southern and a Northern gene pool suggests that the two clusters represent regionally adapted gene pools. Thus the question

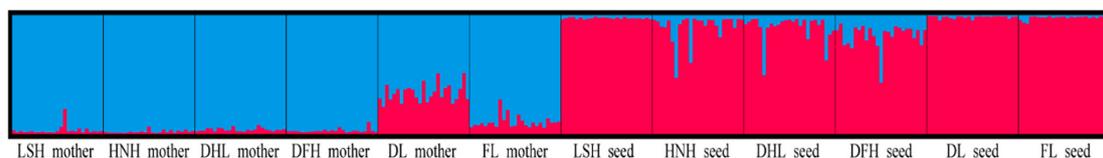


Fig. 2. Results of the STRUCTURE analysis of all populations at $K = 2$.

arises whether hybrid offspring with higher diversity and potential hybrid fitness effects, e.g. heterosis (Agrawal, 2009; Oakley et al., 2015), actually show higher fitness in the field compared to local seed that may have lower diversity, but is regionally/locally adapted and therefore may outperform foreign genotypes. To assess the relevance of the “Mix or Match” debate (Lesica and Allendorf, 1999; Bucharova et al., 2019), more extensive experimental research should be carried out.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

This work is supported by CFERN & BEIJING TECHNO SOLUTIONS Award Funds on excellent academic achievements and the Project of National Natural Science Foundation of China (NSFC 41877549 and 41701052). We would like to acknowledge Chengrui Mao, Yan Wang, Yang Tang, Hong Chen and all the people from the Forestry Bureau in Lushuihe Town that helped us with the sampling.

Author statement

All the co-authors have read and approved the submitted version of the manuscript, and all those entitled to authorship have been so named. The content of this manuscript is original and no other articles using this database have been submitted (or are going to be submitted in the next three months) elsewhere. All the materials, raw data, and protocols used in the article are available upon request and without any restriction, and will be published in a public repository if the article is accepted for publication.

References

- Agrawal, A.F., 2009. Spatial heterogeneity and the evolution of sex in diploids. *Am. Nat.* 174, S54–S70. <https://doi.org/10.1086/599082>.
- Ai, C., Xu, L.A., Lai, H.L., Huang, M.R., Wang, Z.R., 2006. Genetic diversity and paternity analysis of a seed orchard in *Pinus massoniana*. *Scientia Silvae Sinicae*. 42, 146–150 (In Chinese).
- Aizawa, M., Kim, Z.S., Yoshimaru, H., 2012. Phylogeography of the Korean pine (*Pinus koraiensis*) in northeast Asia: inferences from organelle gene sequences. *J. Plant Res.* 125, 713–723. <https://doi.org/10.1007/s10265-012-0488-4>.
- Belletti, P., Ferrazzini, D., Piotti, A., Monteleone, I., Ducci, F., 2012. Genetic variation and divergence in Scots pine (*Pinus sylvestris* L.) within its natural range in Italy. *Eur. J. Forest Res.* 131, 1127–1138. <https://doi.org/10.1007/s10342-011-0584-3>.
- Bernhardsson, C., Floran, V., Ganea, S.L., Garcia-Gil, M.R., 2016. Present genetic structure is congruent with the common origin of distant Scots pine populations in its Romanian distribution. *For. Ecol. Manage.* 361, 131–143. <https://doi.org/10.1016/j.foreco.2015.10.047>.
- Bucharova, A., Bossdorf, O., Hoelzel, N., Kollmann, J., Prasse, R., Durka, W., 2019. Mix and match: regional admixture provenancing strikes a balance among different seed-sourcing strategies for ecological restoration. *Conserv. Genet.* 20, 7–17. <https://doi.org/10.1007/s10592-018-1067-6>.
- Chaloupkova, K., Stejskal, J., El-Kassaby, Y.A., Frampton, J., Lstiburek, M., 2019. Current advances in seed orchard layouts: two case studies in conifers. *Forests* 10, 93. <https://doi.org/10.3390/f10020093>.
- Chen, M.M., Feng, F.J., Sui, X., Li, M.H., Zhao, D., Han, S.J., 2010. Construction of a framework map for *Pinus koraiensis* Sieb. et Zucc. using SRAP, SSR and ISSR markers. *Trees-Struct. Funct.* 24, 685–693. <https://doi.org/10.1007/s00468-010-0438-5>.
- Dawson, I.K., Guariguata, M.R., Loo, J., Weber, J.C., Lengkeek, A., Bush, D., Cornelius, J., Guarino, L., Kindt, R., Orwa, C., Russell, J., Jannadass, R., 2013. What is the relevance of smallholders' agroforestry systems for conserving tropical tree species and genetic diversity in circa situ, in situ and ex situ settings? A review. *Biodiv. Conserv.* 22, 301–324. <https://doi.org/10.1007/s10531-012-0429-5>.
- Durka, W., Michalski, S.G., Berendzen, K.W., Bossdorf, O., Bucharova, A., Hermann, J.-M., Hoelzel, N., Kollmann, J., 2017. Genetic differentiation within multiple common grassland plants supports seed transfer zones for ecological restoration. *J. Appl. Ecol.* 54, 116–126. <https://doi.org/10.1111/1365-2664.12636>.
- El-Kassaby, Y.A., Ritland, K., 1996. Genetic variation in low elevation Douglas-fir of British Columbia and its relevance to gene conservation. *Biodivers. Conserv.* 5, 779–794. <https://doi.org/10.1007/bf00051786>.
- Evanno, G., Regnaut, S., Goudet, J., 2010. Detecting the number of clusters of individuals using the software STRUCTURE: a simulation study. *Mol. Ecol.* 14, 2611–2620. <https://doi.org/10.1111/j.1365-294X.2005.02553.x>.
- Feng, F.J., Sui, X., Chen, M.M., Zhao, D., Han, S.J., Li, M.H., 2010. Mode of pollen spread in clonal seed orchard of *Pinus koraiensis*. *J. Biophys. Chem.* 1, 33–39. <https://doi.org/10.4236/jbpc.2010.11004>.
- Frankham, R., Ballou, J.D., Briscoe, D.A., 2002. Introduction to conservation genetics. Cambridge University Press, Cambridge, UK.
- Geburek, T., Konrad, H., 2008. Why the conservation of forest genetic resources has not worked. *Conserv. Biol.* 22, 267–274. <https://doi.org/10.1111/j.1523-1739.2008.00900.x>.
- Giertych, M., 1975. Seed orchard designs. *Forestry Commission Bull.* 25–37.
- Given, D.R., 1994. Principles and practice of plant conservation. Timber Press, Portland.
- Goudet, J., 1995. FSTAT (version 1.2): A computer program to calculate f-statistics. *J. Hered.* 86, 485–486. <https://doi.org/10.1093/oxfordjournals.jhered.a111627>.
- Hofmann, M., Durka, W., Liesebach, M., Bruelheide, H., 2015. Intraspecific variability in frost hardiness of *Fagus sylvatica* L. *Eur. J. Forest Res.* 134, 433–441. <https://doi.org/10.1007/s10342-015-0862-6>.
- Holliday, J.A., Aitken, S.N., Cooke, J.E.K., Fady, B., Gonzalez-Martinez, S.C., Heuertz, M., Jaramillo-Correa, J.-P., Lexer, C., Staton, M., Whetten, R.W., Plomion, C., 2017. Advances in ecological genomics in forest trees and applications to genetic resources conservation and breeding. *Mol. Ecol.* 26, 706–717. <https://doi.org/10.1111/mec.13963>.
- Hubisz, M.J., Falush, D., Stephens, M., Pritchard, J.K., 2009. Inferring weak population structure with the assistance of sample group information. *Mol. Ecol. Resour.* 9, 1322–1332. <https://doi.org/10.1111/j.1755-0998.2009.02591.x>.
- Iwaizumi, M.G., Miyata, S., Hirao, T., Tamura, M., Watanabe, A., 2018. Historical seed use and transfer affects geographic specificity in genetic diversity and structure of old planted *Pinus thunbergii* populations. *For. Ecol. Manage.* 408, 211–219. <https://doi.org/10.1016/j.foreco.2017.10.026>.
- Iwaizumi, M.G., Tsuda, Y., Ohtani, M., Tsumura, Y., Takahashi, M., 2013. Recent distribution changes affect geographic clines in genetic diversity and structure of *Pinus densiflora* natural populations in Japan. *For. Ecol. Manage.* 304, 407–416. <https://doi.org/10.1016/j.foreco.2013.05.026>.
- Jakobsson, M., Rosenberg, N.A., 2007. CLUMPP: a cluster matching and permutation program for dealing with label switching and multimodality in analysis of population structure. *Bioinformatics* 23, 1801. <https://doi.org/10.1093/bioinformatics/btm233>.
- Jia, D., Zhen, Z., Zhang, H.G., Tang, J.H., 2017. EST-SSR marker development and transcriptome sequencing analysis of different tissues of Korean pine (*Pinus koraiensis* Sieb. et Zucc.). *Biotechnol. Biotechnol. Equip.* 31, 679–689. <https://doi.org/10.1080/13102818.2017.1331755>.
- Johansen-Morris, A.D., Latta, R.G., 2006. Fitness consequences of hybridization between ecotypes of *Avena barbata*: Hybrid breakdown, hybrid vigor, and transgressive segregation. *Evolution* 60, 1585–1595. <https://doi.org/10.1554/05-680.1>.
- Kang, K.S., Lindgren, D., Mullin, T.J., 2001. Prediction of genetic gain and gene diversity in seed orchard crops under alternative management strategies. *Theor. Appl. Genet.* 103, 1099–1107. <https://doi.org/10.1007/s001220100700>.
- Kang, K.S., Lindgren, D., Mullin, T.J., Choi, W.Y., Han, S.U., 2005. Genetic gain and diversity of orchard crops under alternative management options in a clonal seed orchard of *Pinus thunbergii*. *Silvae Genetica*. 54, 93–96. <https://doi.org/10.1515/sg-2005-0014>.
- Kim, Z.S., Hwang, J.W., Lee, S.W., Yang, C., Gorovoy, P.G., 2005. Genetic variation of Korean pine (*Pinus koraiensis* Sieb. et Zucc.) at allozyme and RAPD markers in Korea, China and Russia. *Silvae Genetica* 54, 235–246. <https://doi.org/10.1515/sg-2005-0034>.
- Ledig, F.T., 1992. Human impacts on genetic diversity in forest ecosystems. *Oikos* 63, 87–108. <https://doi.org/10.2307/3545518>.
- Lesica, P., Allendorf, F.W., 1999. Ecological genetics and the restoration of plant communities: mix or match? *Restor. Ecol.* 7, 42–50. <https://doi.org/10.1046/j.1526-100X.1999.07105.x>.
- Litkowiec, M., Lewandowski, A., Wachowiak, W., 2018. Genetic variation in *Taxus baccata* L.: a case study supporting Poland's protection and restoration program. *Forest Ecol. Manage.* 409, 148–160. <https://doi.org/10.1016/j.foreco.2017.11.026>.
- Loo, J., Souvannavong, O., Dawson, L.K., 2014. Seeing the trees as well as the forest: the importance of managing forest genetic resources. *For. Ecol. Manage.* 333, 1–8. <https://doi.org/10.1016/j.foreco.2014.08.014>.
- Loveless, M.D., Hamrick, J.L., 1984. Ecological determinants of genetic-structure in plant-populations. *Annu. Rev. Ecol. Syst.* 15, 65–95. <https://doi.org/10.1146/annurev.es.15.110184.000433>.
- Ma, J.L., Zhang, L.W., Cheng, D., Li, J.W., 1992. Geographic distribution of *Pinus koraiensis* in the world. *J. Northeast Forestry Univ.* 20, 40–47 (In Chinese).
- Mandak, B., Hadincova, V., Mahelka, V., Wildova, R., 2013. European invasion of North American *Pinus strobus* at large and fine scales: high genetic diversity and fine-scale genetic clustering over time in the adventive range. *PLoS ONE* 8. <https://doi.org/10.1371/journal.pone.0068514>.
- Millennium Ecosystem Assessment, 2005. *Ecosystems and Human Well-being: Synthesis*. Island Press, Washington, DC.
- Miyaki, M., 1987. Seed dispersal of the Korean pine, *Pinus koraiensis*, by the red squirrel, *Sciurus vulgaris*. *Ecol. Res.* 2, 147–157. <https://doi.org/10.1007/BF02346923>.
- Moritz, C., 1999. Conservation units and translocations: strategies for conserving evolutionary processes. *Hereditas* 130, 217–228. <https://doi.org/10.1111/j.1601-5223.1999.00217.x>.
- Nielsen, U.B., Hansen, O.K., 2012. Genetic worth and diversity across 18 years in a Nordmann fir clonal seed orchard. *Annals Forest Sci.* 69, 69–80. <https://doi.org/10.1007/s13595-011-0159-y>.
- Nybom, H., 2004. Comparison of different nuclear DNA markers for estimating intra-specific genetic diversity in plants. *Mol. Ecol.* 13, 1143–1155. <https://doi.org/10.1111/j.1365-294X.2004.02141.x>.
- Oakley, C.G., Agren, J., Schemske, D.W., 2015. Heterosis and outbreeding depression in

- crosses between natural populations of *Arabidopsis thaliana*. *Heredity* 115, 73–82. <https://doi.org/10.1038/hdy.2015.18>.
- Park, J.M., Kwon, S.H., Lee, H.J., Na, S.J., El-Kassaby, Y.A., Kang, K.S., 2017. Integrating fecundity variation and genetic relatedness in estimating the gene diversity of seed crops: *Pinus koraiensis* seed orchard as an example. *Can. J. For. Res.* 47, 366–370. <https://doi.org/10.1139/cjfr-2016-0223>.
- Peakall, R., Smouse, P.E., 2012. GenAlEx 6.5: genetic analysis in Excel. Population genetic software for teaching and research - an update. *Bioinformatics* 28, 2537–2539. <https://doi.org/10.1093/bioinformatics/bts460>.
- Petit, R.J., Duminil, J., Fineschi, S., Hampe, A., Salvini, D., Vendramin, G.G., 2005. Comparative organization of chloroplast, mitochondrial and nuclear diversity in plant populations. *Mol. Ecol.* 14, 689–701. <https://doi.org/10.1111/j.1365-294X.2004.02410.x>.
- Potenko, V.V., Velikov, A.V., 1998. Genetic diversity and differentiation of natural populations of *Pinus koraiensis* (Sieb. et Zucc.) in Russia. *Silvae Genetica* 47, 202–208. <https://doi.org/0000-0002-8442-4825>.
- Potter, K.M., Jetton, R.M., Bower, A., Jacobs, D.F., Man, G., Hipkins, V.D., Westwood, M., 2017. Banking on the future: progress, challenges and opportunities for the genetic conservation of forest trees. *New Forest* 48, 153–180. <https://doi.org/10.1007/s11056-017-9582-8>.
- Prat, D., Arnal, S., 1994. Allozyme variation and mating system in 3 artificial stands of Douglas-fir (*Pseudotsuga menziesii* (mirb) franco) planted in Europe. *Silvae Genetica* 43, 199–206.
- Pritchard, H.W., Moat, J.F., Ferraz, J.B.S., Marks, T.R., Camargo, J.L.C., Nadarajan, J., Ferraz, I.D.K., 2014. Innovative approaches to the preservation of forest trees. *For. Ecol. Manage.* 333, 88–98. <https://doi.org/10.1016/j.foreco.2014.08.012>.
- Rajora, O.P., Mosseler, A., 2001. Challenges and opportunities for conservation of forest genetic resources. *Euphytica* 118, 197–212. <https://doi.org/10.1023/a:1004150525384>.
- Ratnam, W., Rajora, O.P., Finkeldey, R., Aravanopoulos, F., Bouvet, J.-M., Vaillancourt, R.E., Kanashiro, M., Fady, B., Tomita, M., Vinson, C., 2014. Genetic effects of forest management practices: Global synthesis and perspectives. *For. Ecol. Manage.* 333, 52–65. <https://doi.org/10.1016/j.foreco.2014.06.008>.
- R Core Team, 2013. R: A language and environment for statistical computing.
- Roberds, J.H., Conkle, M.T., 1984. Genetic-structure in loblolly - pine stands - Allozyme variation in parents and progeny. *Forest Sci.* 30, 319–329.
- Schneller, J., Liebst, B., 2007. Patterns of variation of a common fern (*Athyrium filix-femina*; Woodsiaaceae): population structure along and between altitudinal gradients. *Am. J. Bot.* 94, 965–971. <https://doi.org/10.3732/ajb.94.6.965>.
- Schwartz, K.R., Parsons, E.C.M., Rockwood, L., Wood, T.C., 2017. Integrating *in-situ* and *ex-situ* data management processes for biodiversity conservation. *Front. Ecol. Evol.* 5, 120. <https://doi.org/10.3389/fevo.2017.00120>.
- Shi, M.M., Michalski, S.G., Chen, X.Y., Durka, W., 2011. Isolation by elevation: genetic structure at neutral and putatively non-neutral loci in a dominant tree of subtropical forests, *Castanopsis eyrei*. *PLoS ONE* 6. <https://doi.org/10.1371/journal.pone.0021302>.
- Shi, M.M., Michalski, S.G., Welk, E., Chen, X.Y., Durka, W., 2014. Phylogeography of a widespread Asian subtropical tree: genetic east-west differentiation and climate envelope modelling suggest multiple glacial refugia. *J. Biogeogr.* 41, 1710–1720. <https://doi.org/10.1111/jbi.12322>.
- Sonstebo, J.H., Tollefsrud, M.M., Myking, T., Steffenrem, A., Nilsen, A.E., Edvardsen, O.M., Johnskas, O.R., El-Kassaby, Y.A., 2018. Genetic diversity of Norway spruce (*Picea abies* (L.) Karst.) seed orchard crops: effects of number of parents, seed year, and pollen contamination. *For. Ecol. Manage.* 411, 132–141. <https://doi.org/10.1016/j.foreco.2018.01.009>.
- Toth, E.G., Vendramin, G.G., Bagnoli, F., Cseke, K., Hoehn, M., 2017. High genetic diversity and distinct origin of recently fragmented Scots pine (*Pinus sylvestris* L.) populations along the Carpathians and the Pannonian Basin. *Tree Genet. Genomes* 13. <https://doi.org/10.1007/s11295-017-1137-9>.
- Vieira, M.L.C., Santini, L., Diniz, A.L., Munhoz, C.d.F., 2016. Microsatellite markers: what they mean and why they are so useful. *Genet. Mol. Biol.* 39, 312–328. <https://doi.org/10.1590/1678-4685-gmb-2016-0027>.
- Weir, B.S., Cockerham, C.C., 1984. Estimating F-statistics for the analysis of population structure. *Evolution* 38, 1358–1370. <https://doi.org/10.2307/2408641>.
- Xu, Y.L., Cai, N.H., Woeste, K., Kang, X.Y., He, C.Z., Li, G.Q., Chen, S., Duan, A.A., 2016. Genetic diversity and population structure of *Pinus yunnanensis* by simple sequence repeat markers. *Forest Sci.* 62, 38–47. <https://doi.org/10.5849/forsci.15-064>.
- Yu, D.P., Zhou, L., Zhou, W.M., Ding, H., Wang, Q.W., Wang, Y., Wu, X.Q., Dai, L.M., 2011. Forest management in Northeast China: history, problems, and challenges. *Environ. Manage.* 48, 1122–1135. <https://doi.org/10.1007/s00267-011-9633-4>.
- Yu, J.H., Chen, C.M., Tang, Z.H., Yuan, S.S., Wang, C.J., Zu, Y.G., 2012. Isolation and characterization of 13 novel polymorphic microsatellite markers for *Pinus koraiensis* (Pinaceae). *Am. J. Bot.* 99, 421–424. <https://doi.org/10.3732/ajb.1200145>.
- Yuan, H., Niu, S., El-Kassaby, Y.A., Li, Y., Li, W., 2016. Simple genetic distance-optimized field deployments for clonal seed orchards based on microsatellite markers: as a case of Chinese pine seed orchard. *PLoS ONE* 11. <https://doi.org/10.1371/journal.pone.0157646>.
- Zobel, B.J., Barber, J., Brown, C.L., Perry, T.O., 1958. Seed orchards—their concept and management. *J. Forest.* 56, 815–825.