

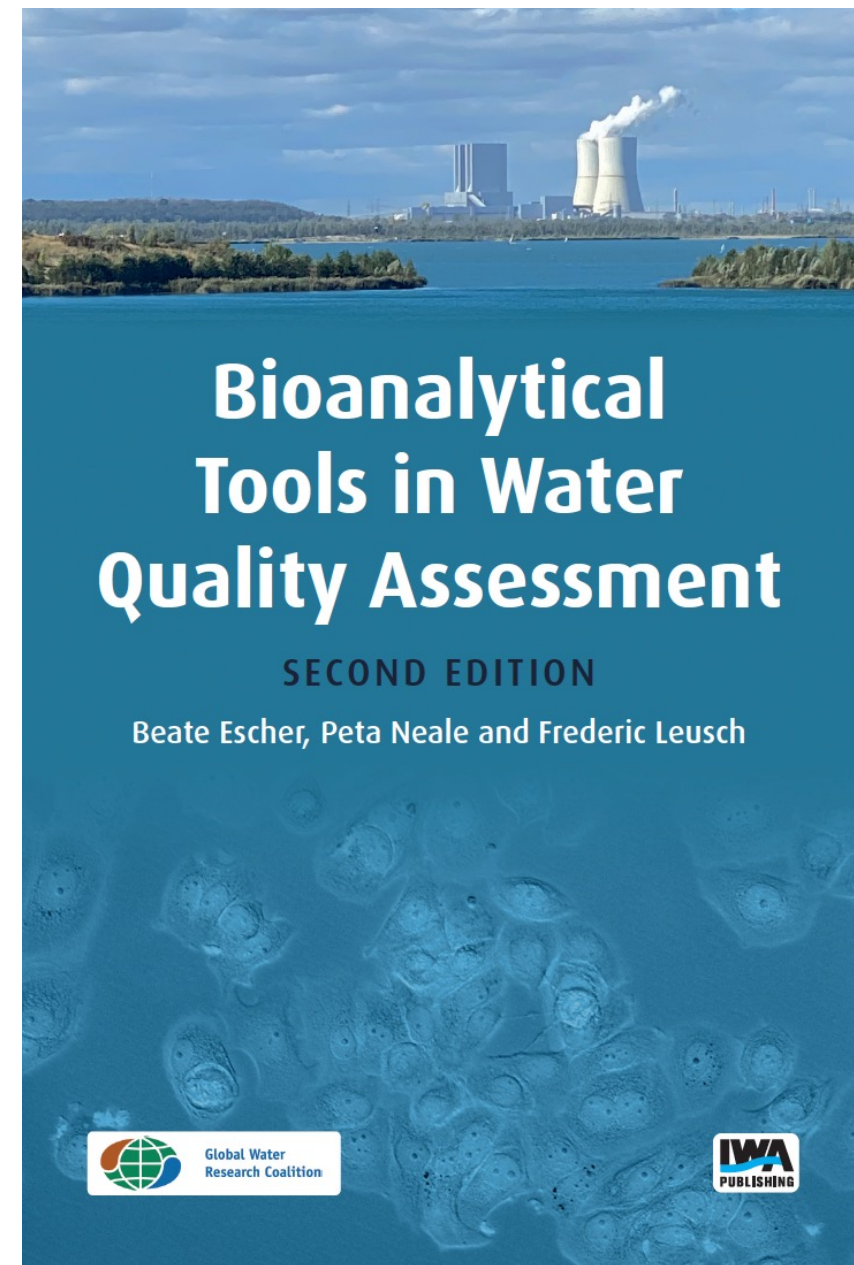
Chapter 12

Sampling, sample preparation and dosing

This presentation accompanies Chapter 12 of “Bioanalytical Tools in Water Quality Assessment”
<https://www.iwapublishing.com/books/9781789061970/bioanalytical-tools-water-quality-assessment-2nd-edition>

Exercises and more material can be found at
www.ufz.de/bioanalytical-tools.

For questions please send e-mail to bioanalytical-tools@ufz.de



Learning goals

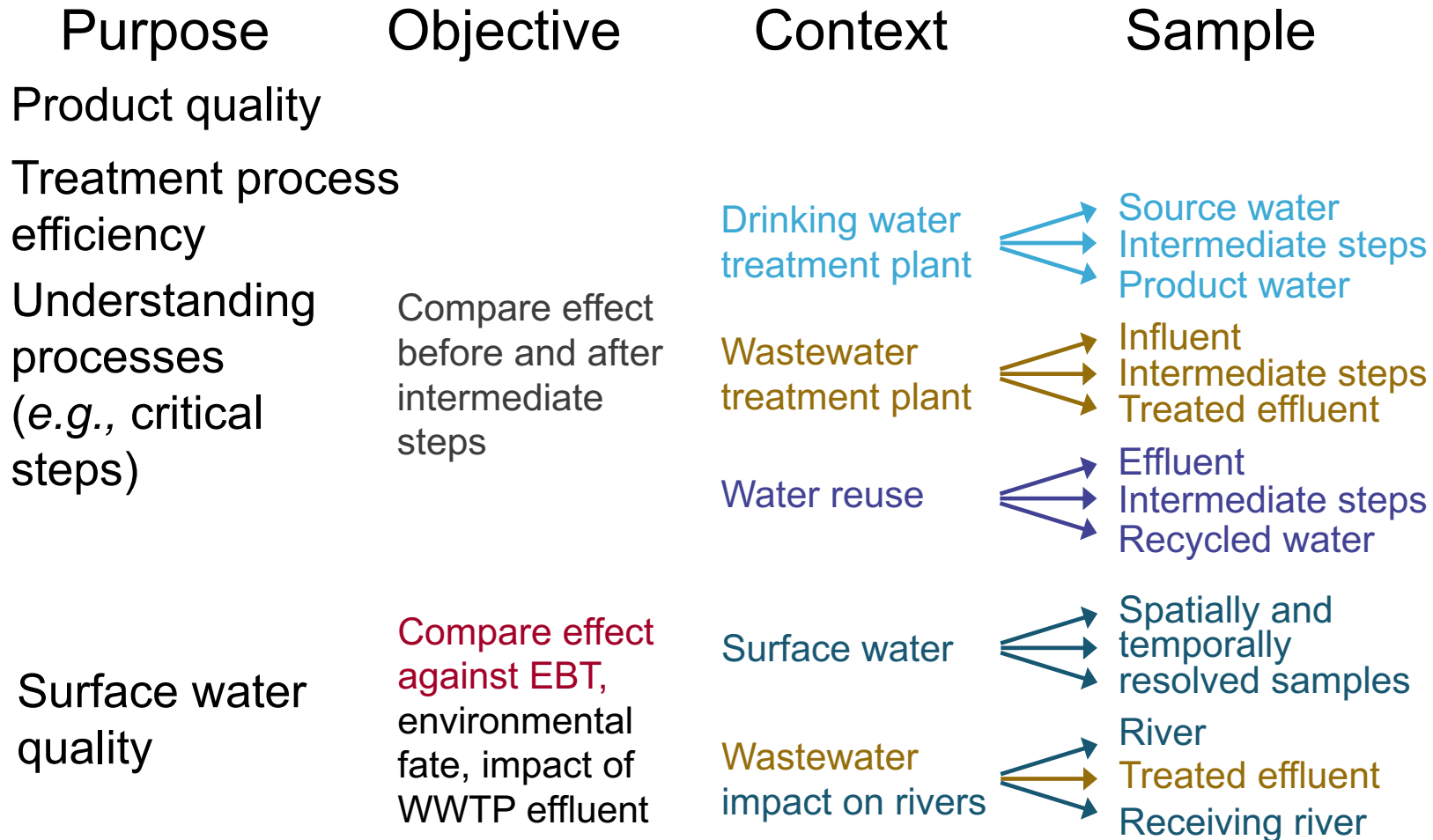
- Appreciate the importance of sampling and sample preparation
Even the best bioassay cannot rectify problems created by poor sample collection and preparation
- Validation of extraction ideally not only with bioassay but also with chemical analysis
- The bioassay is not forgiving (contaminated solvents and equipment will show, no internal standards for recovery etc. possible)
- Sampling and dosing need to go hand in hand



Water sampling strategy

Purpose	Objective	Context	Sample
Product quality	Compare effect against EBT	Drinking water treatment plant	→ Product water
		Wastewater treatment plant	→ Treated effluent
		Water reuse	→ Recycled water
Treatment process efficiency	Compare the effect of inlet and outlet water	Drinking water treatment plant	→ Source water
			→ Product water
		Wastewater treatment plant	→ Influent
			→ Treated effluent
		Water reuse	→ Effluent
			→ Recycled water

Water sampling strategy



Time resolution of sampling

- Grab water samples: drinking water, recycled water
- 24h composite samples (diurnal variation): wastewater treatment plants
- 7 consecutive 24h composite samples (weekly variation)
- Seasonal sampling (4 times a year)
- Event-triggered sampling:
 - rain and storm events
 - night/day variation

Sampling

- Sample volume

Rule of thumb: 0.5 L wastewater or 1 L surface water or 2 L drinking water on one 200 mg HLB SPE cartridge

- Sample preservation

The best is to extract immediately (<48h). Lowering pH to 3 might help extraction of organic acids and preserve sample

Chlorinated drinking water: quench with Na thiosulphate

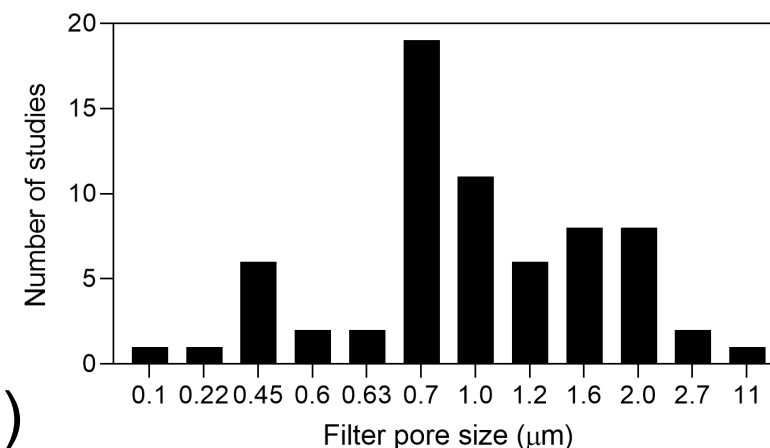
- Sample filtration

Glass fibre filter 0.7 to 2 μm if NTU > 5

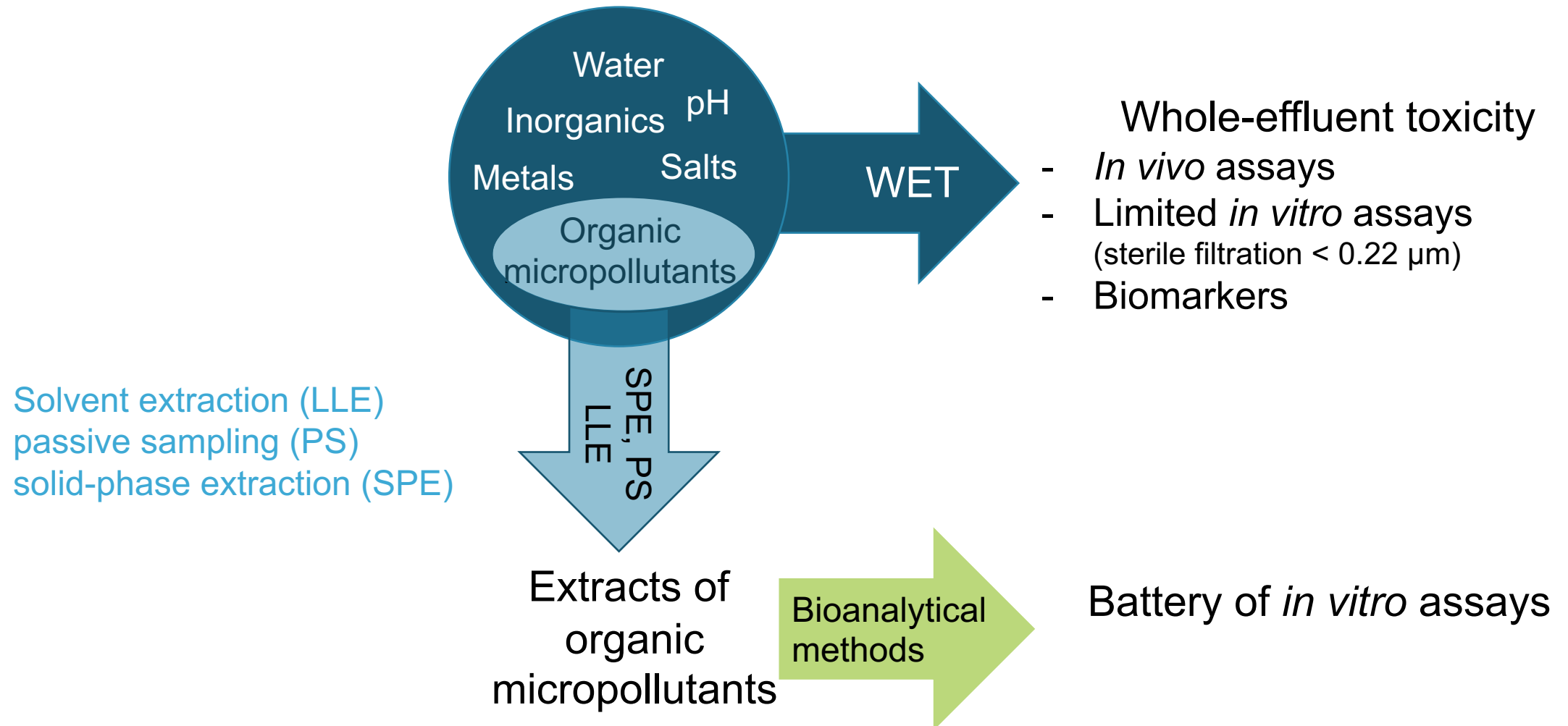
Caution: a lot of bioactivity on SPM!

- Sample extraction

Solvent extraction (LLE) or solid-phase extraction (SPE)

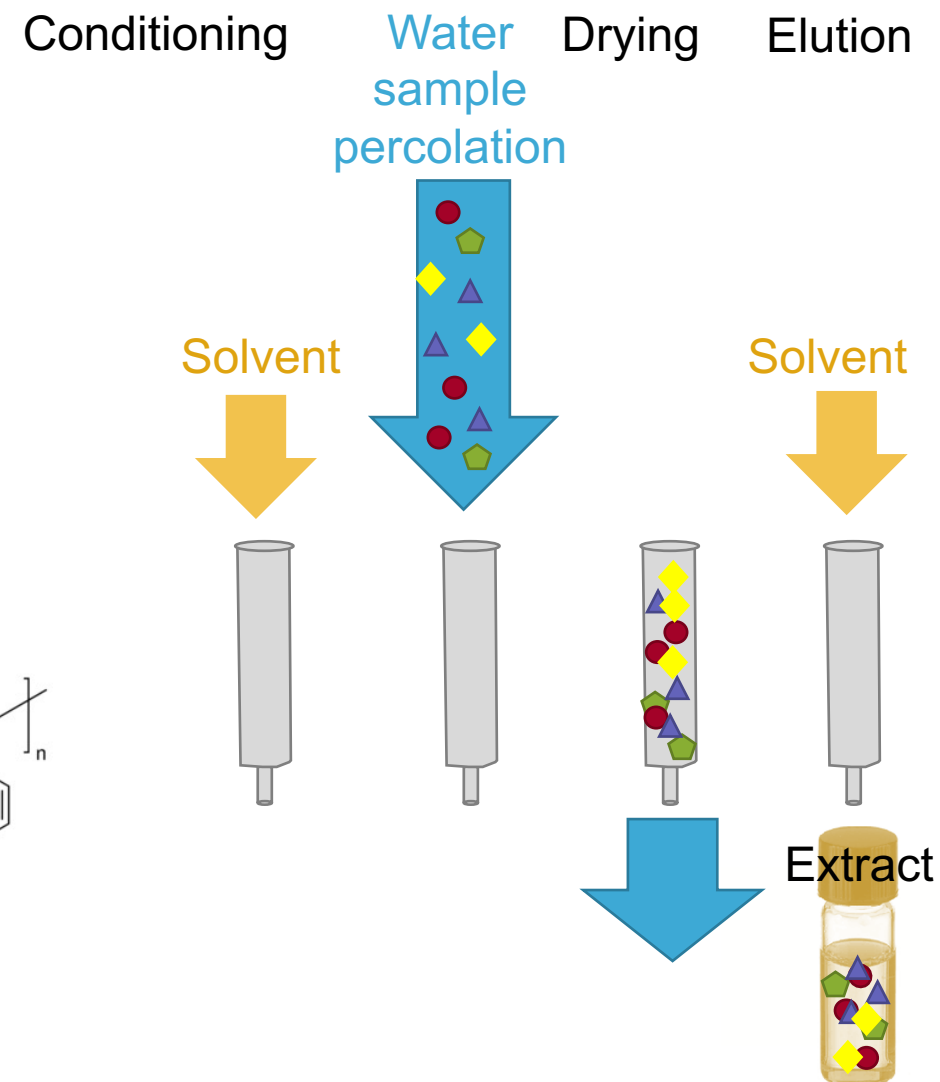
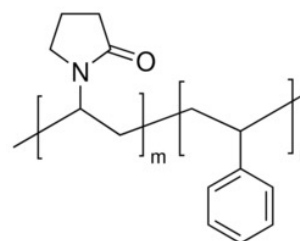


To extract or not to extract?

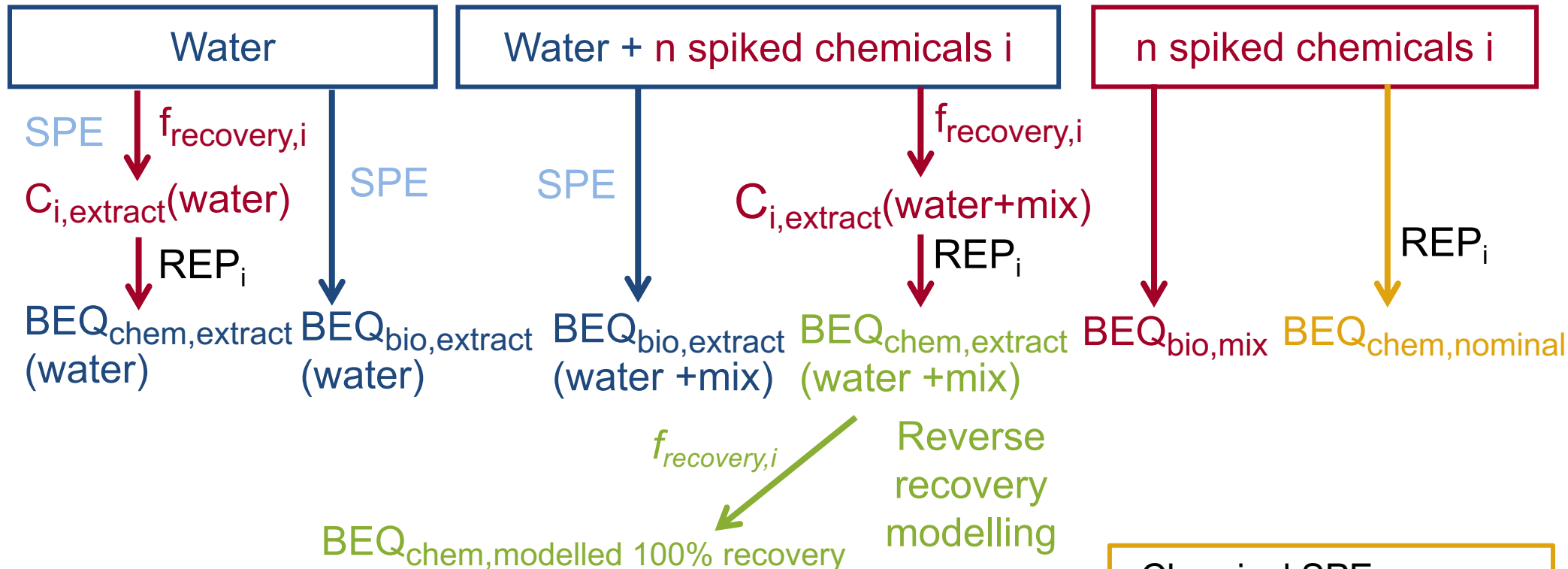


Solid-phase extraction

- Purpose
 - Chemical analysis: selective and specific for target analytes
 - Bioassays: as broad as possible
- Materials
 - Silica-based (C2, C8, C18)
 - Poly(divinylbenzene-co-N-vinylpyrrolidone) copolymers
 - Graphitised carbon black, coconut charcoal
 - Ion-exchange materials (WAX, WCX)
- Solvents:
 - Methanol, ethyl acetate
 - Elution: methanol, ethyl acetate, acetonitrile, dichloromethane, MTBE, hexane:acetone
 - Final solvent: methanol, DMSO
- Capacity: 1-5% of sorbent mass (2-10 mg/200mg_{sorbent})



Effect recovery by SPE



Predicted loss of effect after SPE

$$= 1 - \frac{\text{BEQ}_{\text{chem,extract}}(\text{water+mix})}{\text{BEQ}_{\text{chem, modelled 100\% recovery}}}$$

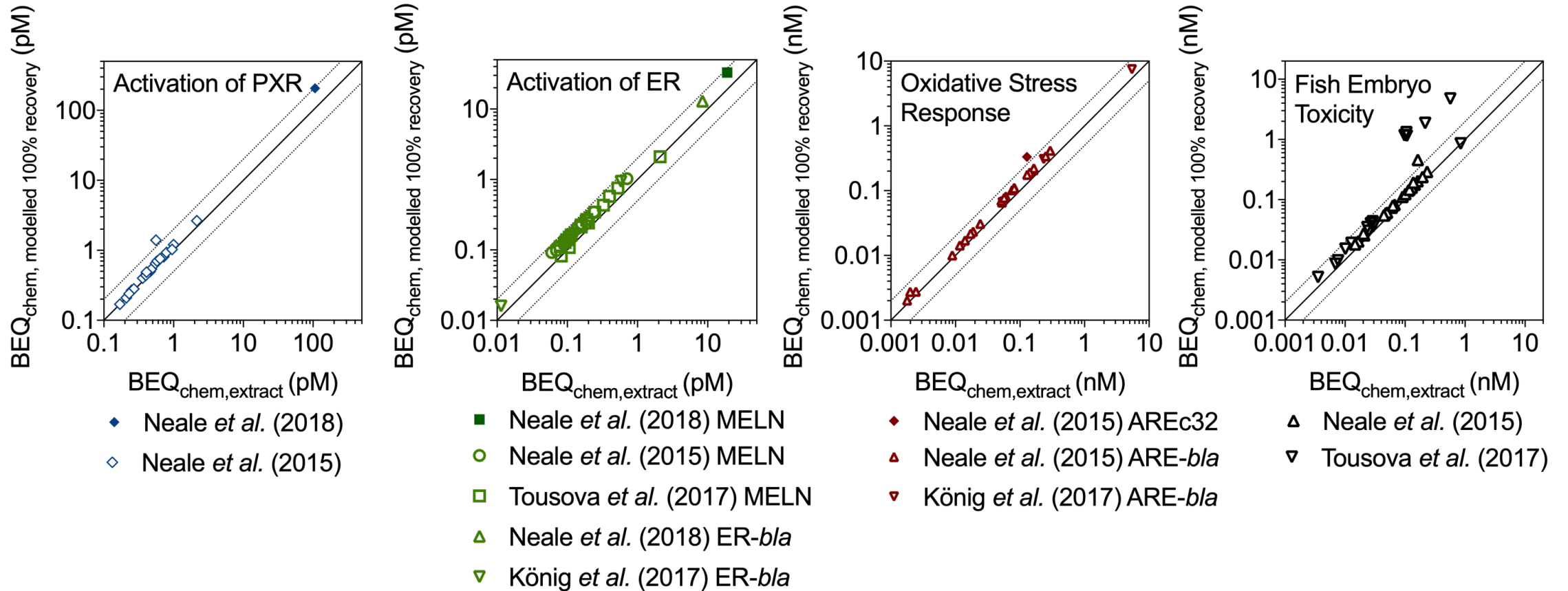
Chemical SPE recovery expressed as effect

$$= \frac{\text{BEQ}_{\text{bio,mix}}}{\text{BEQ}_{\text{chem,nominal}}}$$

Effect recovery by SPE

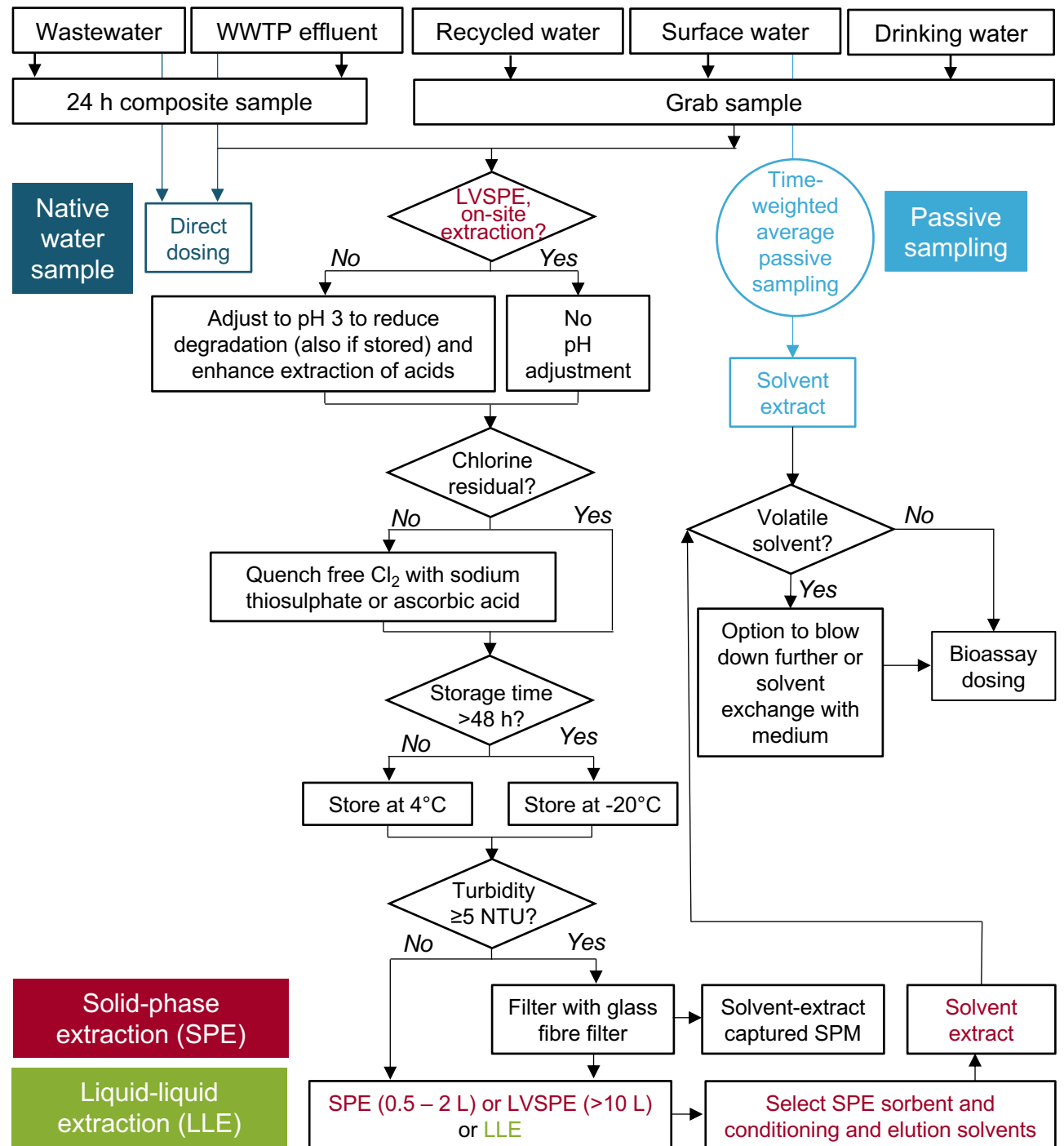
$$= 1 - \frac{\text{BEQ}_{\text{bio,extract}}(\text{water+mix}) - \text{BEQ}_{\text{bio,extract}}(\text{water})}{\text{BEQ}_{\text{bio}}(\text{mix})}$$

Effect recovery by SPE

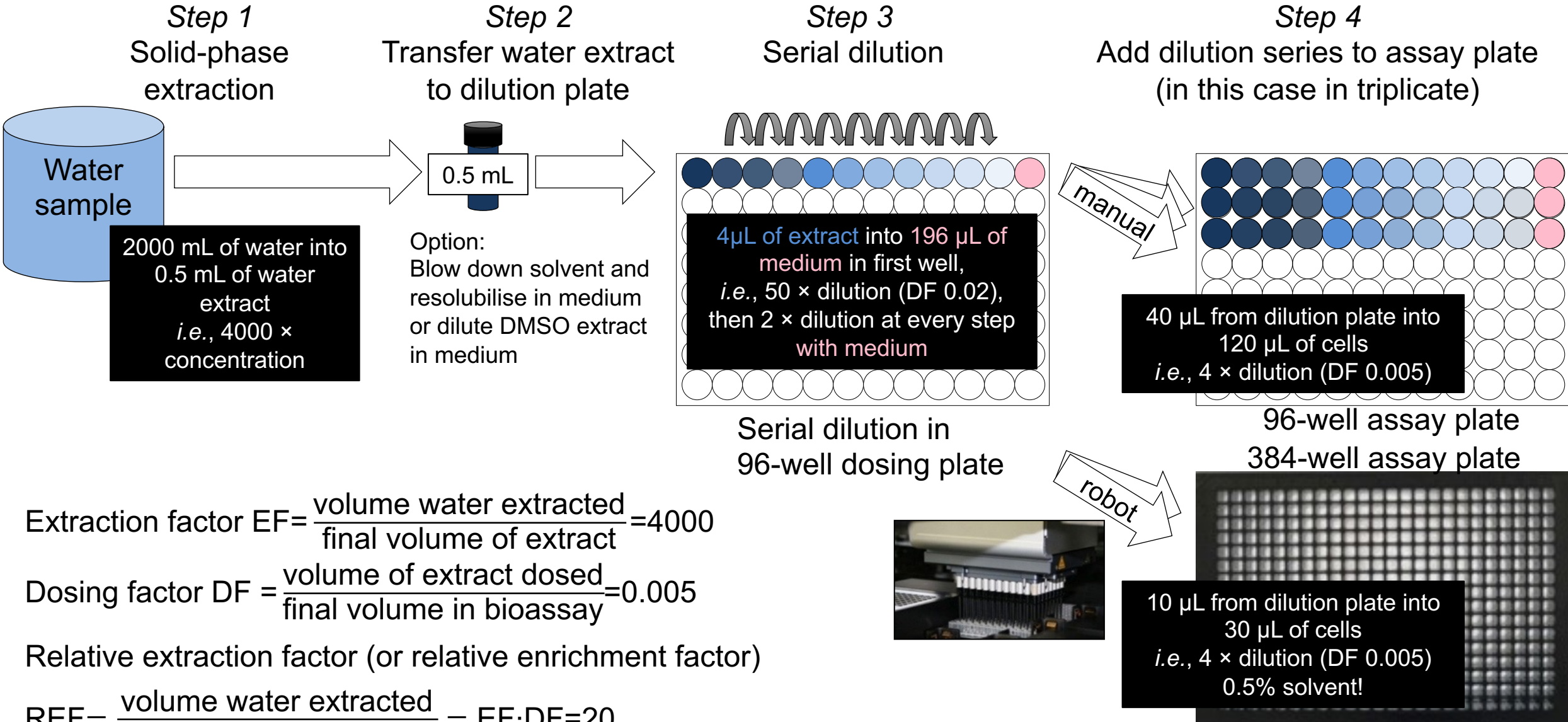


Within a factor of 2!

What to consider.....



Dosing into bioassays



$$\text{Extraction factor } EF = \frac{\text{volume water extracted}}{\text{final volume of extract}} = 4000$$

$$\text{Dosing factor } DF = \frac{\text{volume of extract dosed}}{\text{final volume in bioassay}} = 0.005$$

Relative extraction factor (or relative enrichment factor)

$$REF = \frac{\text{volume water extracted}}{\text{final volume in bioassay}} = EF \cdot DF = 20$$

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