Visualizing the chemical landscape of planktonic photosymbioses using single-cell chemical imaging





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## Marine plankton is highly diverse and has complex trophic modes and life cycles











The ocean is mainly oligotrophic

(N, P, and trace metals)

#### Small cells dominates the plankton community

#### Metabolic strategies of large cells in the open ocean



#### mixotrophy, nutrient storage, and metabolic symbiosis

#### Photosymbiosis between unicellular organisms in the oceanic plankton

Heterotrophic hosts + intracellular microalgae



#### A wide diversity of hosts:

(100-400 µm in size)

A-F: Radiolarians

(G-H) Foraminiferans(I) Dinoflagellates

Symbiosis is obligatory for the host

Benefits for the symbiont?

Decelle J, Colin S and Foster R. 2015

#### Acantharia host the microalga *Phaeocystis*







Decelle et al PNAS 2012

#### Collodaria host the microalga *Brandtodinium*







Free-living form of the dinoflagellate *Brandtodinium* 



Probert et al J Phycol 2014

### **Ecological importance of Radiolaria**



### Eukaryotic plankton diversity in the sunlit ocean

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#### Science 348, (2015); DOI: 10.1126/science.1261605

#### LETTER

doi:10.1038/nature17652

### *In situ* imaging reveals the biomass of giant protists in the global ocean

Tristan Biard<sup>1,2</sup>, Lars Stemmann<sup>2</sup>, Marc Picheral<sup>2</sup>, Nicolas Mayot<sup>2</sup>, Pieter Vandromme<sup>3</sup>, Helena Hauss<sup>3</sup>, Gabriel Gorsky<sup>2</sup>, Lionel Guidi<sup>2</sup>, Rainer Kiko<sup>3</sup> & Fabrice Not<sup>1</sup>

#### Radiolaria represent a <u>high biomass</u> in the plankton community

#### ARTICLE

doi:10.1038/nature16942

## Plankton networks driving carbon export in the oligotrophic ocean

Lionel Guidi<sup>1,2</sup>\*, Samuel Chaffron<sup>3,4,5</sup>\*, Lucie Bittner<sup>6,7,8</sup>\*, Damien Eveillard<sup>9</sup>\*, Abdelhalim Larhlimi<sup>9</sup>, Simon Roux<sup>10</sup><sup>†</sup>, Youssef Darzi<sup>3,4</sup>, Stephane Audic<sup>8</sup>, Léo Berline<sup>1</sup>+, Jennifer R. Brum<sup>10</sup><sup>†</sup>, Luis Pedro Coelho<sup>11</sup>, Julio Cesar Ignacio Espinoza<sup>10</sup>, Shruti Malviya<sup>7</sup>+, Shinichi Sunagawa<sup>11</sup>, Céline Dimier<sup>8</sup>, Stefanie Kandels–Lewis<sup>11,12</sup>, Marc Pichera<sup>11</sup>, Julie Poulain<sup>13</sup>, Sarah Searson<sup>1,2</sup>, *Tara* Oceans Consortium Coordinators<sup>‡</sup>, Lars Stemmann<sup>1</sup>, Fabrice Not<sup>8</sup>, Pascal Hingamp<sup>14</sup>, Sabrina Speich<sup>15</sup>, Mick Follows<sup>16</sup>, Lee Karp–Boss<sup>17</sup>, Emmanuel Boss<sup>17</sup>, Hiroyuki Ogata<sup>18</sup>, Stephane Pesant<sup>19,20</sup>, Jean Weissenbach<sup>13,21,22</sup>, Patrick Wincker<sup>13,21,22</sup>, Silvia G. Acinas<sup>23</sup>, Peer Bork<sup>11,24</sup>, Colomban de Vargas<sup>8</sup>, Daniele Iudicone<sup>25</sup>, Matthew B. Sullivan<sup>10</sup><sup>†</sup>, Jeroen Raes<sup>3,4,5</sup>, Eric Karsent<sup>7,12</sup>, Chris Bowler<sup>7</sup> & Gabriel Gorsky<sup>+</sup>

Radiolaria are stongly involved in <u>carbon export</u> in oligotrophic waters

The physiology and functioning of photosymbiosis in plankton remains unknown



Their ecological success must rely on their efficiency to acquire, transfer and recycle nutrients



Questions

What are the metabolic strategies of the host ?

What is the metabolic role and needs of each partner ?

What is the metabolism of the microalga between the symbiotic and free-living stage (outside the host)?

# Studying physiology of uncultured microbial cells is highly challenging



#### **Bulk analyses**

(transcriptomics, metabolomics, lipidomics)

High cell biomass No spatial information



Fessenden M, Nature 2016

### → <u>single-cell chemical imaging</u>

Single-cell approach: no need to have cultures Maintain <u>physical integrity</u> (relevant for symbioses) <u>Spatial information</u>: Localization of a metabolite or element

#### Sampling in the Mediterranean Sea

(Bay of Villefranche sur Mer, France)



- → Sampling in surface waters with a plankton net
- → Rapid isolation of individual <u>host cells</u> in natural seawater

Cultures of *Phaeocystis* and *Brandtodinium* (free-living symbionts)

Collaboration with Sophie Marro and John Dolan (LOV: CNRS/UPMC)

### Sample preparation for chemical imaging

#### **1- Cryo-fixation with High-Pressure Freezing** (Leica HPM100)

The best method for preserving the <u>ultrastructure</u> and <u>native chemistry</u> of cells





**2- Freeze substitution** -90°C to -30°C for 5 days with Acetone + osmium tetroxide (Leica AFS2)

**3- Resin Embedding** (Room temperature)

#### **4- Ultra-Sectioning**





Collaboration with IBS, Grenoble (Benoit Gallet)

## Correlated approach between electron microscopy and chemical imaging



# Results - I

**Ultratructure organisation** of the host-symbiont integration

Morphology of the symbionts in the host vs free-living

Electron microscopy (SEM/TEM)

### Acantharia- *Phaeocystis* (symbiont)



#### Acantharia- Phaeocystis (symbiont)

TEM

## **Inside the host Outside the host** 8-12 μm 3-4 µm Morphological transformation of the symbiont towards a powerful photosynthetic machinery 0.5 µn

In the host: the volume of the microalga increases with more chloroplasts and thylakoids



#### Collodaria – *Brandtodinium* (symbiont)



#### Collodaria – *Brandtodinium* (symbiont)

#### **Outside the host**

#### Inside the host



#### Morphological transformation of the symbiont towards a powerful photosynthetic machinery

In the host: increase of the size of the symbiont and surface area of chloroplasts

# Results-II

#### Metabolic transformation of the symbionts ?

Metabolic costs for the host ?

## Correlated approach between electron microscopy and chemical imaging



ProVIS Centre Leipzig





#### **NanoSIMS<sup>4</sup>**



#### Mass species <sup>12</sup>C<sup>14</sup>N <sup>31</sup>P<sup>16</sup>O2 <sup>31</sup>P <sup>16</sup>O <sup>12</sup>C2 <sup>32</sup>S

Analyses with look@nanosims software (Polerecky et al., 2012)







Analysis beam: Bi3 Sputter beam Ar-cluster

#### Mass spectrum (0-800 Da)

IonTof Surface Lab 6 software + reference database from literature

#### Synchrotron X-ray fluorescence to visualize and quantify elements in cells

Synchrotron ESRF, Grenoble



Beam lines: ID21 & ID16B

Κ

3.0

In

3.5

4.0



## Visualization of the **ionome**

(elemental composition of a cell: macronutrients + trace metals)



The ionome provides an additional view of the phenotypic state The ionome can reflect the metabolic capacity and needs of a cell The ionome provides information about the biogeochemical impact

#### Nitrogen: a zoom-in into a single symbiont cell

High N content in chloroplasts → light-harvesting proteins and pigments And carbon-fixation enzymes (e.g. Rubisco) in pyrenoid (Geider and LaRoche 2002)

#### SEM

A metabolic cost for the host as N is poorly available in the ocean



#### Phosphorous: a zoom-in into single symbiont cell

#### Acantharia- *Phaeocystis*

nanoSIMS

 $PO_2$ 

#### Symbionts (chloroplasts) are poor in P (RNA, DNA, phospholipids)



SEN

10 20 30 40 50



#### **Phosphorous:** a zoom-in into a single symbiont cell

#### Symbiotic vs free-living stage



P limitation can block cell division but does not inhibit the photosynt

#### Intracellular photosynthesis (numerous chloroplasts ) source of ROS

existence of antioxidant mechanisms ?

## Sulfur metabolism

The chloroplast has a key role in sulfate reduction for the production of:

- The amino acids <u>cysteine</u> and <u>methionine</u>
- <u>Glutathione</u> and <u>phytochelatins</u>.
- <u>DMSP</u>(Dimethylsulfopropionate), <u>DMS</u> (Dimethylsulfide) and <u>DMSO</u> (Dimethylsulfoxide)

These molecules play a role in <u>antioxidant</u> protection and global <u>sulfur cycle</u>

#### An antioxidant function for DMSP and DMS in marine algae

W. Sunda\*, D. J. Kieber+, R. P. Kiene‡ & S. Huntsman\*



Takahashi et al 2011; Meyer and Weis 2012; Sunda et al 2002; Deschaseaux, et al 2014

Subcellular mapping of sulfur

Acantharia - Phaeocystis



#### → High sulfur content in symbionts (1.7 times more than in the host)

Collaboration with Giulia Veronesi (CEA – ESRF- Synchrotron Grenoble)



#### → High sulfur content in symbionts (2.8 times more than in the host)

Collaboration with Giulia Veronesi (CEA – ESRF- Synchrotron Grenoble)

#### Subcellular mapping of sulfur in symbionts

<sup>32</sup>S with nanoSIMS



#### Symbiont Brandtodinium

#### Sulfur in thylakoid membranes, pyrenoid, vacuoles

#### High sulfur content in vacuoles



#### Vacuoles contain up to 6.5 times more S than in chloroplasts



#### 

#### Summary

#### **Morphological** and metabolic reconfiguration of the symbiont



...towards a more dynamic view of the metabolism

### Perspectives

- Visualize and quantify the uptake and transfer of C, N and S with stable isotopes (SIP-nanoSIMS) in correlation with ToF-SIMS



<sup>13</sup>C uptake - 5h incubation

### Thank you for your attention

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